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Abstract: The circadian clock is a self-sustained time-keeping system which controls behavioral, biochemical and physiological rhythmic processes. In mammals, the cogwheels of this clock are the so-called clock genes which control their own expression via several feedback loops. One of these genes is hPER1, a clock gene which disposes of a glucocorticoid-responsive element and might therefore be influenced by glucocorticoids. In humans, stress is associated with an increase in the glucocorticoid cortisol and is seen as a major factor in the etiology of numerous mental health problems. For this reason, our goal was to investigate the putative cortisol-mediated influence of acute and chronic psychosocial stress on the gene expression of hPER1 as well as hPER2, another related clock gene from the same family. We therefore applied laboratory psychosocial stress to thirty-one healthy men and measured cortisol as well as mRNA levels of hPER1 and hPER2. Our main findings suggest that acute psychosocial stress influences the expression of hPER1 and hPER2 dependent on the subjective experience of chronic stress. We therefore conclude that the reactivity to acute stress on the gene expression level of these two genes differs significantly between subjects with high chronic stress compared to subjects with low chronic stress.

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Effects of Psychosocial Stress on the Gene Expression of the Clock Genes \( hPER1 \) and \( hPER2 \) in Humans

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The circadian clock is a self-sustained time-keeping system which controls behavioral, biochemical and physiological rhythmic processes. In mammals, the cogwheels of this clock are the so-called clock genes which control their own expression via several feedback loops. One of these genes is \( hPER1 \), a clock gene which possesses a glucocorticoid-responsive element and might therefore be influenced by glucocorticoids. In humans, stress is associated with an increase in the glucocorticoid cortisol and is seen as a major factor in the etiology of numerous mental health problems. For this reason, our goal was to investigate the putative cortisol-mediated influence of acute and chronic psychosocial stress on the gene expression of \( hPER1 \) as well as \( hPER2 \), another related clock gene from the same family. We therefore applied laboratory psychosocial stress to thirty-one healthy men and measured cortisol as well as mRNA levels of \( hPER1 \) and \( hPER2 \). Our main findings suggest that acute psychosocial stress influences the expression of \( hPER1 \) and \( hPER2 \) dependent on the subjective experience of chronic stress. We therefore conclude that the reactivity to acute stress on the gene expression level of these two genes differs significantly between subjects with high chronic stress compared to subjects with low chronic stress.

Keywords: Circadian Rhythm; Clock Genes; Cortisol; Gene Expression; \( hPER1 \); \( hPER2 \); Psychosocial Stress; Biological Clock

Introduction

All biological processes recurring daily, also known as circadian-controlled processes, are governed by the so-called circadian clock, a self-sustained time-keeping system controlling rhythmic behavioral, biochemical and physiological processes (Panda et al., 2002; Reppert & Weaver, 2002). In other words, the circadian clock affects the sleep-wake cycle, body temperature, hormone secretion, enzyme activity, renal blood flow, heart rate, and various other physiological activities as well as the regulation of the cell cycle and of apoptosis (Ko & Takahashi, 2006; Albrecht, 2004; Lévi et al., 2007). The neuronal structure of the suprachiasmatic nuclei (SCN) within the hypothalamus acts as central pacemaker in mammals. Its neurons oscillate in a self-sustained fashion and synchronize the expression of clock genes in the peripheral cells (Ralph et al., 1990; Czeisler et al., 1999). The main Zeitgeber for this vital circadian process connecting the outside world with the pacemaker is light: photic information is transmitted via specific ganglion cells of the retina over the retino-hypothalamic tract to the SCN (Lucas et al., 1999), where the neurons begin to oscillate. Within the single cells, the mechanism of the molecular clockwork is based on transcriptional/translational feedback loops (TTFLs) of so-called clock genes. These TTFLs exist not only in the SCN but in almost every cell of the body (Lévi & Schibler, 2007). The inner clock is therefore composed of many different and tissue-specific clocks within single cells (Albrecht & Eichele, 2003), which show a great inter-individual variety of expression patterns in humans (Teboul et al., 2005). The synchronization between the central clock (SCN) and the periphery seems to be performed by neuronal and/or hormonal signaling (Kriegsfeld & Silver, 2006; Bujs et al., 2003). Along with this synchronization, there appear to be more entrainment factors for peripheral oscillators. Increasing evidence has demonstrated, for example, that the timing of food intake can “adjust” the peripheral clocks and therefore act as a strong Zeitgeber (Damiola et al., 2000; Stokkan et al., 2001; Schibler et al., 2003). Likewise, it has been shown that glucocorticoids (e.g. dexamethasone) are capable of “resetting” the clock in peripheral tissue (Balsalobre et al., 2000). It has been proposed that amongst other pathways glucocorticoids and genes of the \( Per \) family (in particular \( Per1 \)) interact via glucocorticoid-responsive elements (GRE) (Yamamoto et al., 2005). Also the gene expression of other core clock genes as e.g. \( Rev-ERBa \), \( Npas2 \), could be stimulated in response to the synthetic glucocorticoid dexamethasone in primary mesenchymal cells, the mechanism of the molecular clockwork is based on...
stem cells of mice. These clock genes showed significant changes of their transcript levels within 4 hours after dexamethasone exposure. In Per2, there could be 8 different identified glucocorticoid binding sites (GBS's) of which one seems to be constantly occupied during Per2’s rhythmic expression and is furthermore essential for the regulation of glucocorticoids (So et al., 2009). In addition, the CLOCK protein has been shown to act on glucocorticoid receptors thereby reducing their DNA-binding capacity to GRE-sites (Nader et al., 2009).

One of the best explored glucocorticoids in humans is cortisol. Its presence varies according to a circadian rhythm, peaking in the morning immediately after an individual’s awakening and decreasing as the day progresses. The synthesis of cortisol is induced by the activation of the hypothalamus-pituitary-adrenal (HPA) axis. The corticotropin-releasing hormone (CRH) is released from the paraventricular nucleus (PVN) of the hypothalamus, inducing the synthesis of adrenocorticotropic (ACTH). This in turn stimulates the adrenal cortex, resulting in the synthesis of cortisol (Kirschbaum et al., 2007). There is abundant evidence that the HPA axis is strongly activated in situations of psychosocial stress, and after which there is a considerable increase in cortisol levels (Kudielka et al., 2003). Moreover, increasing evidence shows that cortisol present in the morning is not exclusively induced through an activation of the HPA axis, but also through light-induced activation of the adrenal glands (Scheer & Buijs, 1999; Ishida et al., 2005). This may imply an even stronger relationship between the circadian clock and glucocorticoids than hitherto assumed. Furthermore recent research could show a variation of gene expression in the hippocampus according to a promoter polymorphism in the Per3 gene and its association to the stress response in mice: there was a significantly higher Per3 expression in stressed animals compared to non stressed animals (Wang et al., 2012).

It appears that a disrupted circadian clock is not only clearly associated with physical illness, e.g. cancer (Stevens, 2005; Fu et al., 2002), diabetes (Woon et al., 2007) or sleeping disorders (Viola et al., 2007), but is also involved in several mental health problems such as affective disorders (Roybal et al., 2007; Benedetti et al., 2008) or schizophrenia (Peng et al., 2007). Due to the fact that stress is considered as a factor strongly involved to the fact that sleeping habits might change with age (Yamazaki et al., 2002). The investigation took place on two consecutive days in the laboratories of the Department of Clinical Psychology and Psychotherapy of the University of Zurich.

Exclusion criteria were jetlag, medication, sleeping disorders, intake of psychotropic substances, psychological or physical illness, hospitalization, smoking and shift work within four months prior to the data acquisition. The study participants were instructed to adhere to a strict schedule for one week before they participated in the investigation: they were asked to sleep and eat on a regular basis according to their own, inherent rhythm.

Procedure, Sampling Intervals and Stress/Control Condition: The study participants were instructed not to exercise and to avoid stressful psychological experiences if possible during the two study days. They were asked to arrive at the laboratory at 1930h. On one of the two evenings, the participants were required to undergo a psychosocial stress test (stress condition); on the other evening, they had no intervention at all (control condition). While the sequence of the conditions was randomly assigned, the sampling intervals for cortisol and hPER1 as well as hPER2 were identical on both evenings. For cortisol: 2000h (baseline), 2030h (sample immediately after psychosocial stress in the stress condition), 2040h, 2050h, 2100h, 2115h, 2130 h and 2200h. For hPER1 and hPER2: 2000h (baseline), 2030h (sample immediately after psychosocial stress in the stress condition), 2100h, 2130h, 2200h and 2210h (exactly 2 hours after the onset of psychosocial stress in the stress condition).

Sampling Methods and Analysis of mRNA and Cortisol: mRNA levels of hPER1 as well as hPER2 were assessed in oral mucosa (Bjarnason et al., 2001) as described previously by Cajochen and colleagues (Cajochen et al., 2006) and analyzed in our biochemical laboratory of the Psychological Institute of the University of Zurich. In brief, each sample was taken with a pipette tip (epDualfilter T.I.P.S. 50 - 1000, Eppendorf, Hamburg, Germany), immediately pipetted into a mixture of 0.7 µl of β-Mercaptoethanol and 100 µl Lysis buffer (Absolutely RNA Nanoprep Kit, Stratagene) and then frozen at −80°C. RNA extraction was performed using the Absolutely RNA Nanoprep Kit (Stratagene), followed by reverse transcription with Superscript III First Strand Synthesis Super Mix for qRT-PCR (Invitrogen). The quantitative real-time PCR was conducted using TaqMan Universal PCR Mastermix (Applied Biosystem) on an ABI 7700 Sequence Detection System (Applied Biosystems). The following primer and probes were used: Hs00429988 for hPER1, HS00256143_m1 for hPER2 and 4352934E for the endogenous control hGAPDH (all Applied Biosystems). The amount of target, normalized to the endogenous reference (GAPDH) and relative to a calibrator, was calculated by 2−ΔΔCT.

Cortisol samples were collected using Salivettes (Sarstedt, Sevelen, Switzerland) and subsequently frozen at −20°C. Saliva samples were assayed with Luminescence Immunoassay (IBL) in the Laboratory of Biopsychology of the Technical University of Dresden, Germany.
Assessment of Psychometric Data: Assessment of chronic stress within the three months prior to measurement was conducted using the Trier Inventory of Chronic Stress (TICS). This is a well-validated and reliable instrument used to retrospectively collect psychometric data relating to the subjective experience of stress (Schulz & Schlotz, 1999). This questionnaire measures six aspects of chronic stress: Work overload, worries, social stress, lack of social recognition, work discontent and intrusive memories. The answers are given on a five-point rating scale.

Psychosocial stress test: In the stress condition, the Trier Social Stress Test (TSST) was applied. The TSST is a standardized laboratory stress protocol which reliably induces psychosocial stress in humans and consists of a speech task and a mental arithmetic task, both of which must be accomplished in front of an audience and a video camera. The subject is informed ten minutes before the stress task that he has five minutes to prepare a short speech in which he should apply for a job. Speeches are made in front of two persons dressed in doctors’ overalls, who are instructed to be very serious and unfriendly. After five minutes the speeches are interrupted and subjects are told to count backwards beginning at 2047 with intervals of 17. After each mistake, the subject must start again at the beginning (Kirschbaum et al., 1993).

Statistics: Statistics were calculated using SPSS 15.0. No participant was excluded from the calculation. However, due to missing data in the measurement of hPER1 and hPER2, there were cases which could not be included (relevant for calculations with linear mixed models: for hPER1 30 subjects, for hPER2 22 subjects were included). The poor analyzability of hPER2 is difficult to explain. We assume that some of the specific primer probes unfortunately did not work well. Psychometric and cortisol data were complete.

To compare the total amount of cortisol for both control and stress condition, areas under the curve were calculated (Preusser et al., 2003) using the formula: AUCt = \((m_2 - m_1)/m_1 \times \text{ttotal}\). While \(m_2\) stands for the height of cortisol at a specific measurement time point, \(m_1\) describes the lengths in minutes between two measurement time points. The total amount of increase/decrease of cortisol was determined using the formula: AUCI = AUCt – \(m_1 \times \text{ttotal}\). In order to compare the amount of cortisol that increased between subjects, the area under the curve is calculated in relation to the first measurement time point; therefore, the product of the first measured cortisol value and the overall measurement period (in minutes: ttotal) is subtracted from the total amount of cortisol.

Examinations considering repeated measurements were calculated using linear mixed models (LMM). Due to missing data in the measurement of hPER1 and hPER2, this procedure was more appropriate than the general linear model for repeated measures (ANOVA) since it also includes incomplete cases in the analysis. Moreover, general linear models assume independence of repeated measurements, while linear mixed models are appropriate for dependent and therefore nested data. For these reasons we chose to calculate the data using this hierarchical linear model procedure (see e.g. Peugh & Enders, 2005). Correlations were calculated using Pearson’s correlation (two-tailed).

Results

The focus of this study was to investigate the link between the well-researched glucocorticoid cortisol and the clock genes hPER1 and hPER2. Furthermore, it was our goal to explore the influence of chronic as well as acute psychosocial stress on the expression of hPER1 and hPER2 in one group of subjects, namely 31 healthy men aged 20 to 30. Data concerning both gene expression and cortisol were collected at different time points in the evenings of two consecutive days. We distinguished between a stress and a control condition and compared the levels of cortisol as well as the levels of hPER1 and hPER2 (always in relation to hGAPDH) and psychometric data.

Diurnal Decrease and Stress-Induced Increase of Cortisol in the Evenings

Subjects participated under a different condition each evening. On the first evening, subjects were randomly assigned to the control or the stress condition and were then placed in the opposite group on the second evening. While in the control condition, cortisol levels decreased by 20% (mean) between 2000h and 2030h due to the regular circadian decline. In the stress condition a stress-induced cortisol increase of 244% (mean) was observed. The diurnal decrease/stress-induced increase (AUCi) and the total amount of cortisol (AUCt) in the evening were calculated using the formula for the area under the curve. This was conducted for both control and stress conditions: In the control condition, the cortisol decrease in the evening ranged from −735.25 to −3.98 nmol/l (mean = −234.93; sd = 201.88), while in the stress condition cortisol increase varied from −545.08 to 1812.25 nmol/l (mean = 462.03; sd = 521.00). The total amount of cortisol (AUCt) of the control evening ranged from 75.65 nmol/l to 798.75 nmol/l (mean = 334.45; sd = 200.63), while the total amount of cortisol of the stress evening displayed a range from 206.43 nmol/l to 2089.45 nmol/l (mean = 960.96; sd = 494.81). The stress-induced difference is highly significant (\(F = 173.422; p = 0.0004\)) and can be seen as a successful induction of psychosocial stress.

Stress Matters: The Effect of Chronic Stress on Cortisol

To assess the perception of chronic stress levels, subjects were required to fill in a questionnaire (screening scale of the Trier Inventory of Chronic Stress, TICS) which recorded the level of subjectively experienced chronic stress over the three months immediately preceding data collection. The scores varied from 26 to 63 (mean = 46.87, sd = 10.51). The scores were median-split, providing two groups of subjects: one group with low levels of subjectively perceived chronic stress (n = 18) and the other with high levels (n = 13).

Subjects who reported higher levels of chronic stress showed significantly lower cortisol levels in the evening (control condition: \(F = 19.637, p < 0.000; \) see Figure 1(a); stress condition: \(F = 11.715, p = 0.001; \) see Figure 1(b)).

Subjects with Chronic Stress Show Higher Levels of hPER1 and hPER2

The baseline amounts of mRNA of hPER1 and hPER2 were measured at 2000h (baseline) and did not differ between day one and day two. All mRNA measurements were calculated in
Figure 1.
The influence of chronic stress on cortisol levels and gene expression levels of hPER1 and hPER2 for both, control and stress condition.

ratio to the house-keeping gene hGAPDH. In order to assess the influence of chronic stress levels on the mRNA levels of hPER1 and hPER2, we used the above described psychometric data of the TICS comparing the subjects of the two median-split groups as described above. In contrast to the inverse relationship between chronic stress and cortisol, higher chronic stress seems to be associated with higher levels of hPER1. This effect is significant in the stress situation (F = 8.502; p = 0.004; see Figure 1(d)), but not in the control condition (F = 0.502; p = 0.480; see Figure 1(c)). This indicates significantly differing gene expression levels of hPER1 between subjects with high respectively low chronic stress levels immediately after acute psychosocial stress. While subjects with low chronic stress levels show an immediate suppression of mRNA levels of hPER1 with small standard deviations, there are heightened mRNA levels of hPER1 and greater interindividual differences (higher standard deviation) in subjects with high chronic stress levels after acute stress. Similarly, higher hPER2 levels after psychosocial stress seem to be associated with higher levels of chronic stress (F = 3.305; p = 0.075; trend; see Figure 1(f)), whereas this is not the case in the control condition (F = 1.197; p = 0.287; see Figure 1(e)).

Subjects with high levels of perceived chronic stress show lower levels of cortisol in control as well as stress condition, but significantly higher hPER1 levels in the stress condition (p = 0.004). In addition, the same subjects display higher hPER2 levels in the stress condition (p = 0.075). Interestingly also standard deviations of hPER1 and hPER2 seem to be larger in subjects with high levels of perceived chronic stress.

The Rise of Cortisol after Acute Psychosocial Stress Seems to Be Associated with the Gene Expression of hPER1 and hPER2

As reported above, we assessed the simultaneously measured association between the total amount of cortisol and the gene expression levels of hPER1 and hPER2 in the evening between 2000h and 2210h. This relationship was tested separately for the stress and the control conditions, as the total amounts of cortisol were expected to differ between these conditions. Indeed, there was a highly significant difference between the two conditions in cortisol (see above). Such a marked difference between stress and control condition was not found either for hPER1 (F = 0.307; p = 0.580) or for hPER2 (F = 1.864; p = 0.175).

However after the induction of psychosocial stress the levels of hPER1 as well as hPER2 measured begin to correlate strongly with their baseline levels (all p < 0.0001 for hPER1; see also Table 1). This effect was not seen in the control condition (all p > 0.050). This finding contrasts with the correlations between cortisol levels: due to the constant decrease in cortisol in the evening, there is a strong association between the cortisol levels measured in the control condition, but not in the stress condition. Based on the above-mentioned sharp increase in cortisol levels after the stress task, none of the subsequent cortisol levels correlate with its baseline. It might therefore be deduced that a sudden increase in cortisol (relative change of
cortisol) is involved in “resetting” or “flattening” the fluctuation between the measurements of hPER1 and, to a lesser extent, also of hPER2.

Time points compared to their baseline at 2000 h (compared within conditions): hPER1 and hPER2 seem to react to dramatic changes in levels of cortisol: As soon as cortisol deviates from its circadian decreasing course by means of a stress-induced, sudden increase, hPER1 begins to correlate significantly with its baseline indicating a possible “flattening“ of its course. The same effect, although to a lesser extent, is observed in hPER2 (statistical analysis was calculated using Pearson’s correlation, two-tailed). This effect is not observed in the control condition where cortisol decreases slowly due to its circadian course.

Discussion

To our knowledge, this study is the first to examine the interaction of psychosocial stress and the gene expression of hPER1 and hPER2 in humans.

Regarding the impact of chronic stress, our data show that the levels of hPER1 and hPER2 differ in subjects with high chronic stress compared to subjects with low chronic stress. Similar results have been found in the gene expression of Per3 of stressed animals (Wang et al., 2012). Although reports on the interaction of chronic stress and cortisol are still conflicting (Kudielka et al., 2009), in our sample, high levels of chronic stress are unambiguously associated with lower basal cortisol.

In spite of the fact that there is still no reliable reference for “healthy”, or in the statistical sense “normal”, gene expression levels for either hPER1 or hPER2, or any other clock gene, we can draw the conclusion from the data of our sample that subjects with high chronic stress levels show blunted basal cortisol in the evening. At the same time and in accordance with findings reported above, the expression of hPER1 and also hPER2 is heightened in subjects with higher chronic stress compared to subjects with low reported chronic stress. This interaction is even more pronounced after acute psychosocial stress and might suggest that chronically stressed humans react differently and more instable respectively regarding the influence on the circadian clock. However, we did not find the expected increase of hPER1 and hPER2 after the increase of cortisol due to psychosocial stress as expected according to data from animal studies (e.g. Balsalobre et al., 2000).

While subjects with low chronic stress show a more coherent “flattening” of the mRNA levels of hPER1 and hPER2, subjects with high chronic stress experience show a far greater standard deviation of the measured time points after the acute psychosocial stress, which might indicate a less deterministic and therefore more inefficient physiological reactivity to acute stress. Recent findings that postulate an increased vulnerability to circadian rhythm disruption after experienced stress in the presence of CLOCK and PER3 polymorphisms in women, support our data (Antypa et al., 2012). Our findings report about the change within the first two hours after psychosocial stress and therefore might describe a very first reaction to psychosocial stress at the cost of not fully monitoring the amplitude and frequency of the circadian oscillation of hPER1 and hPER2, which is a short-coming of our study. On the other side, the fact that we “put the spyglass” on the 120 minutes immediately after a stressor occurred, might give new insights and hypotheses about important processes of the first minutes of a stress-reaction. Nevertheless, 120 minutes might be too short to detect large effects since the peak of transcribed mRNA accumulation depends on the half-life of the mRNA (and many post-transcriptional processes are not known yet or is still unknown how they influence the half life of mRNA), which on average is about two hours after the inducing signal. Usually, the influence of glucocorticoids on the clock gene expression was measured in large intervals of several hours, which better provides for the circadian oscillation of clock genes, but might miss the very first changes of sensitively interacting systems. Balsalobre et al. (2000) postulated that glucocorticoids might reset the clocks in the periphery and therefore play an important role for the synchronization between the SCN and the periphery, but they cannot be the only signals to entrain the periphery.

Recent results could show that acute systemic inflammatory processes, which involve the activation of pro-inflammatory cytokines, illustrated a suppressive effect on the gene expression of several clock genes amongst others Per1 and Per2.

Okada and colleagues could show that rats which received an intravenous injection of lipopolysaccharide demonstrated notable increases of TNFα, IL-6 and also corticosterone, while the gene expression of Per1 and Per2 in the SCN were slightly suppressed during the day of injection. The same was seen in the cells of the liver with a latency of 4 to 6 hours. Furthermore, a suppressed locomotor activity could be observed, which is also known as “sickness-behavior”. Cavadini et al. (2007) also reported a significant reduction in locomotor activity, prolonged rest time, and impaired gene expression of clock-related genes after the infusion of TNFα. Data from our research group report that immediately after a psychosocial stressor (TSST) cortisol is elevated, while cytokines are significantly decreased. Moreover, within the first 60 minutes post-stress, LPS-stimulated TNFα and IL-6 were predicted by the subjective “control expectancy” as well as the experienced “challenge” (Wirtz et al.,

<table>
<thead>
<tr>
<th>Condition</th>
<th>Compared to baseline measurement at 0800 h</th>
<th>Cortisol correlation (p)</th>
<th>hPER1 correlation (p)</th>
<th>hPER2 correlation (p)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0830 h</td>
<td>0.973 (&lt;0.000)**</td>
<td>0.163 (0.546)</td>
<td>0.087 (0.889)</td>
</tr>
<tr>
<td></td>
<td>0900 h</td>
<td>0.912 (&lt;0.000)**</td>
<td>0.496 (0.060)</td>
<td>−0.214 (0.863)</td>
</tr>
<tr>
<td></td>
<td>0930 h</td>
<td>0.730 (&lt;0.000)**</td>
<td>−0.005 (0.984)</td>
<td>−0.440 (0.458)</td>
</tr>
<tr>
<td></td>
<td>1000 h</td>
<td>0.796 (&lt;0.000)**</td>
<td>0.339 (0.216)</td>
<td>0.980 (0.128)</td>
</tr>
<tr>
<td>Stress</td>
<td>0830 h</td>
<td>0.379 (0.035)†</td>
<td>0.928 (&lt;0.000)**</td>
<td>0.666 (0.071)†</td>
</tr>
<tr>
<td></td>
<td>0900 h</td>
<td>0.107 (0.568)</td>
<td>0.912 (&lt;0.000)**</td>
<td>0.725 (0.042)</td>
</tr>
<tr>
<td></td>
<td>0930 h</td>
<td>0.217 (0.242)</td>
<td>0.887 (&lt;0.000)**</td>
<td>0.805 (0.229)*</td>
</tr>
<tr>
<td></td>
<td>1000 h</td>
<td>0.296 (0.143)</td>
<td>0.815 (&lt;0.000)**</td>
<td>−0.497 (0.503)</td>
</tr>
</tbody>
</table>

Table 1. Highly significant correlations between measurement time points of hPER1 as well as hPER2 after stress-induced increase of cortisol.
was observed in the increase of cortisol. The same effect, although to a lesser extent, was evident after the stress-induced and considerable increase in hPER1 and hPER2 (as shown: Okada et al., 2006; Haimovich et al., 2010) would not be so efficient. This association might be explanatory for our findings and might explain why physical, but also stress-induced mental illness affects the experience of fatigue and/or disturbed sleep patterns on the long term.

In any case, a sudden increase of cortisol seems to affect the mRNA levels of both hPER1 and hPER2 in the periphery or at least in oral mucosa, albeit the effect is not as pronounced as in animal studies which report an increase of clock gene expression (Balsalobre et al., 2000; Takahashi et al., 2001; Fukuoka et al., 2005). First, this might be due to the fact that in animal studies, high dosage rates of glucocorticoids are administered in order to induce a phase shift of clock genes (e.g. so called “dexamethasone-shock”; Balsalobre et al., 2000). Such a high dosage in relation to the animals’ body weight is, of course, not comparable to a relatively short-lived and natural stress-induced rise of cortisol. Therefore, it seems difficult to compare data from human studies with animal studies. Second, it would probably be very dangerous for an organism’s homeostasis if a relatively short-lasting acute stressor would have the impact to severely disturb and/or disrupt the circadian rhythm. Lastly, we did not monitor the development of gene expression over two hours post-stress; therefore, we might have missed a possible later increase or change of the gene expression of hPER1 and hPER2. Nevertheless, we were able to show that acute psychosocial stress in humans has an impact on the gene expression of hPER1 as well as hPER2. Even a natural increase of cortisol—induced by an acute psychosocial stressor—seems to flatten the fluctuation of hPER1 and hPER2. This is evident based on the highly significant correlations between measurements of hPER1 after the stress-induced and considerable increase of cortisol. The same effect, although to a lesser extent, was observed in hPER2. This effect failed to occur in the control condition, when cortisol levels were following their steady circadian decrease. Hence, it might be concluded that humans, depending on their individual stress reactivity, minimally “reset” their clocks after subjectively experienced stressful situations. This might be an important result given that mental and physical health strongly depends on an organism’s ability to restore physiological homeostasis (e.g. McEwan & Lasley, 2003).

In general, it might be misleading to talk about chronic stress in a cross-sectional study since it is very difficult to retrospectively investigate the onset of the so-called chronic stress; therefore, the only appropriate design would be a prospective longitudinal study to adequately investigate chronic stress and its correlates because it is about the time-related process of a maladaptive change (as also alluded in the term “chronic”). This may be the reason why there are many conflicting biopsychological data with large interindividual differences when it comes to chronic stress (Kudielka et al., 2009). In allusion to McEwan et al. (2003), chronic stress might be seen as an ongoing or repeated strain which can lead to an allostatic load. This means that the sensitive equilibrium of highly adaptive and interacting systems of an organism is disturbed and not able to restore homeostasis any more, which might lead to the deregulation of one or more of the interacting psycho-physiological systems. Concerning our data, this could lead to the conclusion that the interaction between cortisol and the gene expression of hPER1 and hPER2 might be additionally mediated by other pathways such as the immune system. Recent findings suggest a transient, but strong suppressive effect of pro-inflammatory cytokines on the gene expression of clock genes (Okada et al., 2006; Haimovich et al., 2010; see above). It seems important to us to emphasize the influence of the time aspect of chronic stress and to focus the process of change over time since the organism’s interacting systems will most probably display different allostatic regulation patterns according to the length of stress endurance. Therefore, cross-sectional studies might be limited in their statements about chronic stress and should not be simplified to universally and generally accepted facts, but at the same time might explain findings which are not conform to the postulated hypotheses. Nevertheless, it is interesting and important to see that subjects who reported about high experienced chronic stress in our study react differently compared to subjects who report low experienced chronic stress. This might indicate the beginning of the shift from allostatic to an allostatic load that could lead to severe illness in the long term.

Cortisol, as part of the endocrine HPA axis pathway, generally seems to have a regulatory role in a putative feedback-mechanism between the inner clock and the endocrine system (Dallmann et al., 2006; Dickmeis et al., 2007), and therefore it seems to be conclusive that even sudden stress-induced increases of cortisol level directly as well as indirectly (via other systems mediated), induce short-period effects in the gene expression of hPER1 and hPER2. On the basis of a strong interaction between the circadian clock and the HPA axis, our finding may be a first meaningful step to explain the disruption of sleep-wake cycles in stress-induced disorders.

Shortcomings of our study are the relatively small number of subjects and the broad range of mRNA levels of hPER1 and hPER2, which makes interpretation of the statistical analyses difficult. The latter is not a new problem and it has been addressed before (Brown et al., 2005; Teboul et al., 2005). One aim of future research should be to examine more subjects in order to raise statistical power. A hardly solvable problem in human research concerning the biological clock seems to be the question, if peripherally measured clock gene expression is significantly correlated with the central gene expression in the brain. Recent findings suggest that peripheral measurements can approximately indicate an individual inner period and body time (Pagani et al., 2010; Kasukawa et al., 2012).

Furthermore, it would be recommendable to take measurements at more points spread evenly over the day in order to better account for the circadian rhythms of the parameters measured. Nevertheless our data fortify the importance to further research the association between stress and the inert biological clock in humans and might indicate a possible mecha-
nism how psychosocial stress might influence or even disrupt an individual’s circadian rhythm on the long term.

Three important questions remain and should be addressed in future research: 1) Which systems are actively involved in the stress reaction, which also constitute signaling pathways that therefore affect the inner clock? 2) Which are the molecular mechanisms that mediate the interaction between these systems and the inner clock? 3) How do these processes allostatically change over time in the presence of an ongoing strain?

One of the crucial goals for future research will be to determine the effect of short-term changes of psychosocial stress and cortisol on the long-term “ticking of the clock”.

REFERENCES


