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1 **Breastfed at Tiffany's**

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7

8 **Keywords**

9 Breast milk, microbiota, IgA, lactose, lactase, oligosaccharide

10

11 **Abstract**

12 The importance of breast milk for the growing infant is undisputed; breastfeeding decreases
13 infantile mortality by 10-fold and decreases the incidence of infectious diseases. Despite its
14 recognized benefits, the structural richness of breast milk has also impeded the
15 characterization of the multiple roles of milk components on infant physiology. However,
16 the important roles of some components of breast milk are beginning to be dissected. For
17 instance, molecules such as immunoglobulin A and milk oligosaccharides protect from
18 gastrointestinal infections and influence the development of the gut microbiota. Deciphering
19 the complex composition of breast milk brings to light multifaceted contributions that sum
20 up to make breast milk the ultimate personalized medicine.

21 **Breastfeeding protects child and mother**

22 Breast milk is often described as being the gold standard of infant nutrition, because breast
23 milk provides all macronutrients and vitamins required for the optimal development of the
24 suckling infant. Recent meta-analyses underline the beneficial effects of breastfeeding on
25 short-term child health by decreasing infant mortality and morbidity [1], and on long-term
26 development by reducing the risk for obesity [2]. Health benefits also extend to the nursing
27 mother, as breastfeeding protects against breast cancer [3]. Considering the strong impact
28 of breast milk on child health, the WHO recommends exclusive breastfeeding for the first six
29 months of life and breast milk as a complement to solid food for at least an additional year
30 [4].

31 More than just a gold standard, breast milk is a jewel shaped by millions of years of
32 evolution that chiseled a perfect multi-functional fluid. In fact, beyond the supply of
33 nutrients and vitamins, breast milk provides bioactive factors including immunoglobulins,
34 cytokines, antimicrobial proteins, hormones, and oligosaccharides, which work in concert to
35 fortify mucosal immunity, shape the gut microbiota, stimulate body growth, and even to
36 regulate birth spacing in mothers. Breast milk is a rich fluid that fulfills multiple tasks as
37 discussed in this Review.

38 **Breast milk is a meal**

39 Let us begin by crunching some numbers. At the beginning of lactation, each human breast
40 produces on average 450 g of milk daily. After 15 months, the daily output still reaches up to
41 200 g of milk, although the amount largely depends on the intensity of breastfeeding [5]. To
42 accommodate this increased energy expenditure the nursing mother has to increase her
43 daily caloric intake of about 2000 kcal [6] by taking up an additional 500 kcal. This

44 supplement nearly compensates for the 625 kcal required for the daily production of 700 to
45 900 g of breast milk. The process itself is very efficient as the conversion of dietary energy to
46 milk energy has been estimated to reach 80% [7]. Altogether, the energy expenditure bound
47 to milk production is considerable and is comparable to the daily caloric uptake of the brain
48 [8].

49 The true structural and functional richness of breast milk emanates from the multitude of
50 components included in the fat, protein, and carbohydrate fractions. The composition of
51 breast milk differs largely between mammals. For example, marine mammals have a milk
52 rich in fat, fast-growing mammals have a milk rich in proteins, marsupials and primates have
53 a milk rich in carbohydrates [9]. In humans, the ethnicity and age of nursing mothers have
54 little impact on the overall milk composition [10], but the stage of lactation has by far the
55 largest effect on the individual classes of macronutrient. In general, **colostrum** (see Glossary)
56 has high concentrations of bioactive proteins and oligosaccharides, whereas mature milk has
57 proportionally high levels of lipids and caseins. The maternal diet has little effect on most
58 macronutrient classes, although dietary lipids definitively influence the fatty acid
59 composition of breast milk [11]. Lipids are the largest source of calories, yielding 40-50% of
60 the total dietary energy of breast milk [12]. In addition to triglycerides and cholesterol, the
61 lipid fraction of early milk includes several lipid mediators, such as anti-inflammatory lipoxins
62 and resolvins [13]. Milk proteins are often subdivided into insoluble caseins that build
63 micelles, and soluble whey proteins, which include bioactive proteins such as secreted
64 immunoglobulin A (sIgA), lactoferrin, lysozyme, and α -lactalbumin. The carbohydrate
65 fraction consists of lactose (50-70 g/l) and complex oligosaccharides (5-10 g/l). Despite its
66 structural simplicity and the universal occurrence of glucose (Glc) and galactose (Gal) in

67 living organisms, the disaccharide lactose that combines Glc and Gal is only found in
68 mammals.

69 **Breast milk is a clock**

70 Lactose is synthesized in the secretory epithelium of the mammary gland by coupling Gal in a
71 β 1-4 linkage to Glc. Lactose synthase (EC 2.4.1.22) is a dimer comprising the β 1-4
72 galactosyltransferase B4GALT1 [14] found in the Golgi apparatus of all cells and α -
73 lactalbumin [15], which is specifically expressed in the mammary gland. In the absence of α -
74 lactalbumin, B4GALT1 has a low affinity for Glc as acceptor substrate and preferentially
75 transfers Gal to N-acetylglucosamine (GlcNAc). While associated with α -lactalbumin, the
76 affinity of B4GALT1 for Glc increases by 1000-fold [16], thereby enabling the formation of
77 lactose. During pregnancy the expression of α -lactalbumin is inhibited by high levels of
78 circulating progesterone [17] that counteracts the stimulatory effect of the pituitary
79 hormone prolactin, the levels of which rise strongly in the second half of gestation. At
80 parturition, progesterone drops while sustained prolactin secretion induces α -lactalbumin
81 expression, hence stimulating milk production. In addition to its role as nutrient, lactose is
82 also used as an acceptor substrate for the synthesis of a multitude of oligosaccharides,
83 which will be addressed in the next section.

84 After ingestion of breast milk, lactose must be cleaved back to Gal and Glc in order to be
85 absorbed and used as a source of energy by the suckling infant. The enzyme responsible for
86 lactose cleavage is the β -galactosidase lactase [18], which is expressed at the brush border
87 membrane of the small intestine. Lactase expression is tightly regulated and is progressively
88 turned off in the majority of children around two to three years of age (Fig. 1). Decreased
89 lactase activity leads to the passage of lactose to the large intestine, where it is metabolized

90 by microbes, thereby releasing hydrogen, methane, carbon dioxide, and lactate [19]. These
91 fermentation products cause bloating, abdominal cramps, and nausea, which are the typical
92 symptoms of lactose intolerance. The emergence of such symptoms will lead the nursed
93 child to reject breast milk and eventually to natural weaning.

94 Ovarian follicle maturation is suppressed during lactation because of the elevated prolactin
95 and low gonadotropin levels in nursing mothers [20]. This phenomenon prevents a new
96 pregnancy when a mother dedicates a major fraction of her energy expenditure to
97 breastfeeding. Accordingly, lactase repression and the transition to a state of lactose
98 intolerance can be seen as a natural clock regulating weaning and thereby the return to
99 fertility for the mother (Fig. 1). Therefore, the lactose-lactase system has been suggested to
100 act as a biological timer controlling birth spacing in humans [21].

101 Whereas the majority of mankind loses lactase expression during early childhood, about 40%
102 of the human population shows lifelong lactase persistence. The geographical distribution of
103 lactase persistence is striking as it is mainly localized to Europe, West Africa, the Middle East,
104 and Pakistan/West India. Lactase persistence is in fact a recent trait in human evolution, as
105 the dominant mutation conferring persisting lactase expression appeared about 7500 years
106 ago in Eastern Europe [22]. The high frequency of this lactase haplotype in the European
107 population indicates a strong selection pressure, which coincided with the emergence of
108 dairy cultures across Europe. Distinct mutations in the promoter region of the lactase gene
109 have been reported in West Africa and Asia, indicating that lactase persistence spread across
110 the globe through convergent evolution. The rise of lactase persistence certainly contributed
111 to the expansion of dairy farming and milk consumption in lactose-tolerant populations. The
112 increasing availability of cow's milk introduced alternatives to breast milk for young children
113 and thereby lowered the age of weaning. The resulting shortening of the nursing period also

114 yielded a faster return to fertility in women and thus increased the birth rate in Neolithic
115 farming societies.

116 **Breast milk is a fertilizer**

117 Breast milk is the first fluid ingested by the newborn; it is the first food for the infant but it is
118 also a strong conditioner for the gut microbiota, which develops swiftly in the days following
119 birth. As documented by numerous recent studies, the gut microbiota is emerging as a
120 critical organ involved not just in intestinal physiology, but in influencing general metabolism
121 and affecting the severity of diseases such as diabetes [23] and atherosclerosis [24]. Breast
122 milk, as the product of million years of evolution, provides the optimal seeding ground for
123 the development of a healthy gut microbiota. A better understanding of the coordinated
124 action of breast milk constituents in shaping the gut microbiota will lead to a definition of
125 treatments aimed at restoring a healthy gut microbiota in diseases.

126 In addition to lactose, human breast milk comprises a large number of complex
127 oligosaccharide structures, consisting of three or more monosaccharides, which are also
128 produced in the lactating mammary epithelium. In contrast to lactose that functions as an
129 energy source to the infant, milk oligosaccharides cannot be digested by the suckling infant.
130 Human colostrum, the milk produced during the first few days after the birth of the baby,
131 contains about 22 g/l of oligosaccharides and mature milk still about 12 g/l [25]. Human milk
132 comprises close to 200 distinct oligosaccharides [26], a number that is by far the highest
133 among mammalian milks. This diversity is achieved by combining the four carbohydrates Gal,
134 GlcNAc, fucose (Fuc) and **sialic acid** (Sia) on a lactose core (Fig. 2). The term sialic acid covers
135 in fact a large family of acidic carbohydrates; N-acetylneuraminic acid (Neu5Ac) is the only
136 sialic acid found in humans, whereas Neu5Ac and N-glycolylneuraminic acid (Neu5Gc) are
137 found in most mammals. The glycosyltransferases involved in the assembly of milk

138 oligosaccharides are the same enzymes that build the glycans decorating glycoproteins and
139 glycolipids. The production of milk oligosaccharides is solely regulated by glycosyltransferase
140 expression in the mammary epithelium. As the program of glycosyltransferase gene
141 expression varies between mothers, the amounts of some milk oligosaccharides shows a
142 large degree of inter-individual variability [27].

143 Mammals lack the glycosidase machinery required to cleave milk oligosaccharides in the
144 gastrointestinal tract. Therefore, unaltered milk oligosaccharides reach the large intestine,
145 where they are consumed by selected bacterial taxa. The assimilation of oligosaccharides
146 requires glycosidase enzymes such as fucosidases and sialidases to break down the
147 oligosaccharides into monosaccharides, and carbohydrate transporters in order to use the
148 released monosaccharides as carbon source. Some intestinal bacteria including
149 *Bifidobacterium* spp. and *Bacteroides* spp. are well-equipped to degrade and utilize milk
150 oligosaccharides [28]. Because bacteria can have preferences for specific milk
151 oligosaccharides, differences in the composition of milk oligosaccharides impact the
152 colonization of the gut by individual bacterial groups. For example, “non-secretor” mothers,
153 who lack the fucosyltransferase FUT2, produce milk oligosaccharides devoid of α 1,2-linked
154 Fuc. Infants of such “non-secretor” mothers show delayed intestinal colonization with
155 bifidobacteria [29], which include Fuc consumers such as *Bifidobacterium*
156 *longum* subsp. *infantis* and *Bifidobacterium bifidum*. Compositional shifts in the gut
157 microbiota induced by different milk oligosaccharide mixtures may have long-term effects
158 on the course of inflammatory diseases. For instance, elevated amounts of the milk
159 oligosaccharide Sia(α 2-3)lactose promote the formation of a niche for **Enterobacteriaceae**
160 during lactation, which extensively expands during disease and exacerbates intestinal
161 inflammation in colitis [30]. Surprisingly, Enterobacteriaceae cannot feed on Sia(α 2-3)lactose

162 as they lack the sialidase enzymes able to cleave the capping Sia units. In fact,
163 Enterobacteriaceae rely on sialidases released by other intestinal bacteria for that task, such
164 as members of the *Bacteroides* genus [31]. As milk oligosaccharides are structurally similar
165 to intestinal **mucin** O-glycans, bacterial glycosidases also digest carbohydrates from the
166 protective mucin layer lining the intestine. The release of carbohydrates from milk
167 oligosaccharides and intestinal mucins mediated by bacterial glycosidases eventually
168 supports the “cross-feeding” of **pathobionts** such as Enterobacteriaceae. In the recent years
169 milk oligosaccharides and intestinal glycans have thus been recognized as key players
170 influencing the composition of the gut microbiota under healthy conditions and during
171 disease [32].

172 Maternal sIgA are another component of breast milk controlling the bacterial colonization of
173 the gut. Bacterial antigens are among the epitopes recognized by maternal sIgA and these
174 antibodies bind to intestinal bacteria once they reach the infant gut. Some bacterial taxa,
175 such as Enterobacteriaceae, are more widely coated than others, such as *Prevotella* and
176 *Bacteroides* [33]. Coating of bacteria with sIgA hampers their proliferation in the gut,
177 thereby preventing the expansion of colitogenic bacteria [34]. The importance of maternal
178 sIgA in shaping the gut microbiota has also been demonstrated in newborn mice nursed by
179 mothers unable to transfer sIgA into their milk because of a polymeric Ig receptor defect.
180 Mice fed with antibody-deficient milk presented long-lasting and detrimental changes in
181 their gut microbiota, as exemplified by increasing Pasteurellaceae and Lachnospiraceae
182 levels, and increased susceptibility towards colitis induced by dextran sulfate sodium [35].
183 The milk proteins lysozyme and lactoferrin also influence the gut microbiota by cleaving cell
184 wall polysaccharides and by chelating iron, respectively. Colostrum is especially rich in

185 lactoferrin [36], which binds with high affinity to iron, thereby restricting its availability for
186 the growth of pathobionts, such as Enterobacteriaceae [37].

187 **Breast milk is an umbrella**

188 Breast milk contains physiologically relevant amounts of bioactive proteins including
189 immunoglobulins, cytokines, **defensins** and lactoferrin that contribute to the immune
190 protection of the infant [38] (Fig. 3). Some of these immunomodulatory factors, such as
191 macrophage colony-stimulating factor [39], are produced by epithelial cells in mammary
192 ducts, whereas others, such as transforming growth factor β (TGF β) [40], are produced by
193 leukocytes present in the breast milk. Importantly, these bioactive proteins remain active
194 after passage through the stomach because of a higher gastric pH in infants of about 3-5
195 compared to a gastric pH of 1-2 in adults. The stability of milk proteins is further maintained
196 by α 1-antitrypsin in early milk, which protects other proteins from gastric proteolysis [41].

197 The first bioactive proteins identified in the breast milk were immunoglobulins. Transfer of
198 immunity from mother to child was described in 1903 and this effect was linked to
199 antibodies contained in the milk. The vast majority of immunoglobulins in the breast milk
200 belong to the IgA class. Levels of sIgA reaching 12 g/l are commonly detected in the
201 colostrum, while mature milk contains about 1 g/l [42]. Since the intestinal immune system
202 at birth is immature with a low production of sIgA in the first weeks of life, the high levels of
203 sIgA in the colostrum significantly contribute to the immune protection of an infant. Thus, a
204 transfer of adaptive secretory immunity from mother to an infant in the form of sIgA
205 provides a direct protection against a variety of pathogens until the infant immune system
206 takes over by producing sufficient sIgA levels around a month after birth [43]. Lactoferrin,
207 which reaches concentrations of 1-3 g/l in breast milk, is another immune protective factor.
208 Lactoferrin efficiently chelates iron, at the same time reducing the growth of certain bacteria

209 relying on iron and increasing absorption of iron by the infant through binding to the
210 intestinal lactoferrin receptor ITLN1 [44]. Importantly, the cleavage of lactoferrin by pepsin
211 in the stomach yields lactoferricin, which acts as an antimicrobial peptide by disrupting the
212 membrane of gram-negative bacteria [45]. In addition, lactoferrin induces macrophage
213 phagocytosis, thereby promoting the elimination of certain bacteria [46]. The major milk
214 protein α -lactalbumin also shares antimicrobial properties when partially unfolded and
215 associated with oleic acid [47]. The resulting complex furthermore induces apoptosis in
216 tumor cells, and has therefore been called HAMLET, for human α -lactalbumin made lethal to
217 tumor cells. The antitumor activity of the HAMLET complex demonstrates that milk proteins
218 have therapeutic potential, as for example in treating colon cancer [48].

219 During the first weeks of life, the two anti-inflammatory cytokines interleukin-10 (IL-10) and
220 TGF β that are transferred through breast milk contribute to the maturation of mucosal
221 immunity [38]. Indeed, the milk levels of TGF β correlate with sIgA production in breastfed
222 infants [49], and with a decreased risk for child diseases including allergy [50]. Further
223 studies performed in mice showed that milk TGF β promotes immune tolerance to oral
224 antigens during mucosal maturation [51]. Similarly, targeted deletion of IL-10 in mice leads
225 to spontaneous enterocolitis under conventional housing conditions, which can be
226 prevented by parenteral administration of either IL-10 [52] or TGF β [53].

227 In addition to cytokines, breast milk provides passive immune-protective factors such as
228 lysozyme, defensins [54], and soluble CD14 (sCD14) [55], which assist the infant innate
229 immune system in coping with infections (Fig. 3). The concentration of sCD14 in breast milk
230 is 20-fold higher than in the serum of mothers. Milk α -lactalbumin binds sCD14 and thereby
231 protects it from degradation when passing the stomach. The elevated epithelial permeability
232 in the neonate intestine enables an efficient absorption of sCD14, which sensitizes the

233 innate immune system towards Gram-negative bacteria, thus contributing to the
234 maintenance of microbial homeostasis in the neonatal intestine.

235 Beside their prebiotic action discussed above, milk oligosaccharides also exert anti-microbial
236 functions by acting as soluble receptors for pathogens. For example, H2 type
237 oligosaccharides (Fig. 2) inhibit the adhesion of *Campylobacter jejuni* to the intestinal
238 epithelium [56], and fucosylated milk oligosaccharides from secretor mothers inhibit
239 norovirus infection [57]. Oligosaccharides carrying Lewis X antigens are recognized by the
240 DC-SIGN lectin on intestinal dendritic cells. Such oligosaccharides prevent the binding of HIV
241 through DC-SIGN, thereby decreasing the presentation of the virus to CD4⁺ T cells [58]. Some
242 milk oligosaccharides have been shown to directly regulate immune cells. For example, the
243 oligosaccharide lacto-N-fucopentaose III induces the production of IL-10 in spleen cells [59].
244 Also, oral supplementation of mice with Sia(α 2-3)lactose increases activation of intestinal
245 CD11c⁺ dendritic cells [60]. The activating properties of specific milk oligosaccharides may be
246 related to their structural similarity with carbohydrate epitopes found on pathogens. Indeed,
247 α 2,3-linked Sia is present on surfaces of various pathogenic bacteria, such as group B
248 *Streptococcus* [61], *Campylobacter jejuni* [62], *Haemophilus influenzae*, and *Neisseria*
249 *meningitidis* [63].

250 **Breast milk is a remote control**

251 Besides contributing to the development of gut microbiota and the maturation of the
252 mucosal immune system, breast milk also affects metabolic pathways and supports the
253 growth of the suckling infant. Several hormones occurring in breast milk likely mediate the
254 same functions as they do as endocrine factors. Accordingly, leptin in breast milk [64] is
255 probably involved in controlling satiety and fat storage; insulin-like growth factor 1 (IGF1)
256 [65] is probably involved in stimulating body growth; and adiponectin [66] is probably

257 involved in regulating blood glucose levels and fatty acid oxidation. Whereas these
258 hormones certainly play a role in the early growth and development of breastfed infants, the
259 true significance of milk-borne hormones is elusive as clear experimental support is difficult
260 to obtain.

261 Assessing the biological contribution of breast milk hormones is doubtless a challenging task
262 when trying to differentiate their effects from those mediated by the same hormones
263 produced endogenously. Also, the pleiotropic actions of several hormones render the
264 identification of specific effects quite difficult and can lead to ambiguous conclusions. For
265 example, some studies have attributed behavioral functions to breast milk cortisol by
266 associating cortisol levels in maternal milk with human infant temperament [67], and with
267 reduced anxiety as investigated in rats [68]. Far from discrediting such studies, it must be
268 reminded that cortisol, as the main glucocorticoid hormone, exerts multiple actions and that
269 behavioral changes may be indirect and consecutive to numerous metabolic and
270 immunologic effects. In fact, cortisol is an important factor controlling intestinal immunity
271 [69]. Accordingly, cortisol delivered through breast milk probably contributes to the
272 maintenance of anti-inflammatory conditions in the early phase of intestinal microbial
273 colonization in infants.

274 **Breast milk is a waste basket**

275 Breast milk provides various protective compounds as discussed above, but breast milk also
276 conveys lipophilic **xenobiotics** that accumulate in the maternal breast tissue. The list of
277 environmental contaminants is long, featuring heavy metals, pesticides, synthetic additives,
278 and **endocrine disruptors** among others. Thanks to vigilant scientists such as Rachel Carson
279 [70], who raised awareness in the general community of the health risks of such
280 contaminants, several xenobiotics have been banned over the past 20 years. For example,

281 dichlorodiphenyltrichloroethane (DDT) was broadly used as pesticide in agriculture before
282 the warning call of Rachel Carson, which eventually led to the ban of DDT in 49 countries by
283 1995. DDT and its metabolites affect bird reproduction and are highly toxic to fish. In
284 humans, exposure to DDT has been associated with preterm birth [71] and increased risk for
285 breast cancer [72]. The United States did ban the use of DDT in 1972 and several European
286 countries already restricted the pesticide in 1970. The levels of DDT measured in the breast
287 milk of Swedish mothers peaked by 1970 at 3 µg/g lipids and steadily declined to zero by the
288 end of the twentieth century [73]. Additional studies addressing the accumulation of DDT in
289 the human body revealed a half-life for DDT in human fatty tissues, such as breast tissue, of
290 about four years.

291 Whereas several xenobiotics have been black-listed, others are still widely used. For example,
292 phthalates are non-covalent additives found in plastics, textiles, personal-care products and
293 so forth. Phthalates are released in the environment and accumulate in fat tissues. They are
294 found in dietary products such as in butter but also in breast milk. Phthalates have been
295 claimed to act as endocrine disruptors [74]. Positive correlations have been described
296 between specific phthalates in breast milk and altered levels of sexual hormones in suckling
297 infant boys at three months of age. Especially noteworthy was the detection in such infants
298 of a higher ratio of luteinizing hormone to testosterone than is normal, which is indicative of
299 an anti-androgenic action of some phthalates [75].

300 Besides xenobiotics, breast milk is also involved in the transmission of pathogens, such as
301 HIV and cytomegalovirus, to the suckling infant. Newborns from cytomegalovirus-positive
302 mothers are protected prenatally by the transfer of anti-virus IgG through the placenta [76].
303 The situation is not as positive in the case of HIV, as transmission of the virus has been
304 documented in 10 to 40% of mother-infant pairs [77]. Consequently, the CDC recommends

305 avoiding breast-feeding for HIV-positive mothers [78]. In general, only a few maternal
306 viruses are transmitted through breast milk, which underlines the general safety of
307 breastfeeding.

308 **Concluding remarks**

309 Beyond the biological functions of breast milk addressed in this review, the act of
310 breastfeeding itself is the topic of emotional discussions related to the philosophical
311 question of motherhood. Should society encourage breastfeeding simply because it is
312 "natural"? Is a woman who stops nursing her baby after three months a bad mother? Does
313 breastfeeding depreciate the economic and social status of women [79]? Similar provocative
314 questions keep the general debate on breastfeeding alive and remind us that the discussion
315 on breast milk transcends biology (see Outstanding Questions). Breast milk is ultimately why
316 Carolus Linnaeus, as the father of seven children, chose the term *Mammalia* to define our
317 own class of animals in the tree of life.

318

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323 **Glossary**

324 **Colostrum:** The first milk; it is produced by the end of pregnancy and is secreted in the first
325 four days postpartum. Colostrum is rich in sIgA and milk oligosaccharides, thereby providing
326 a first line of immune defense to the newborn. Bovine colostrum was used as a source of
327 anti-microbial immunoglobulins against infections before the emergence of antibiotics-
328 based therapies.

329 **Defensin:** Short cationic antimicrobial peptides that bind to bacterial and fungal cell walls
330 and kill microbes by destabilizing their membrane integrity. Defensins are mainly produced
331 by leukocytes and by Paneth cells in the crypts of the small intestine. Colonization of the gut
332 by microbiota stimulates the production of defensins.

333 **Endocrine disruptor:** Chemicals that share structural features with hormones and interfere
334 with endocrine pathways. Animals are exposed to endocrine disruptors through different
335 modes, ranging from skin contact to oral ingestion. Some endocrine disruptors are
336 environmental pollutants, such as dioxin, while others are additives to food and materials, as
337 for example bisphenol A, which is found in plastics.

338 **Enterobacteriaceae:** Family of Gram-negative, facultative anaerobic rod-shaped bacteria
339 encompassing *Escherichia coli*, *Shigella*, *Klebsiella*, *Salmonella*, and *Yersinia*. Most
340 Enterobacteriaceae reside in the intestine of animals. The Enterobacteriaceae family
341 includes commensals, pathobionts and pathogens. Enterobacteriaceae cannot process
342 oligosaccharides and large polysaccharides.

343 **Mucin:** Mucins are a family of highly O-glycosylated hydrophilic proteins that are the main
344 constituents of the mucus that protects epithelial layers on mucosal surfaces. Mucins are
345 also found in body fluids such as saliva and phlegm. Mucins are large proteins that are either

346 anchored to cells through a transmembrane domain, or secreted as massive gel-like
347 aggregates. The mucin MUC2, secreted by goblet cells, is the main constituent of the thick
348 mucus layer lining the gastrointestinal tract.

349 **Pathobiont:** Organism that normally lives in symbiosis with a host, but can become
350 pathogenic under specific conditions such as when becoming a dominant taxa in a complex
351 environment. Typical pathobionts among gastrointestinal bacteria are *Helicobacter pylori*,
352 *Clostridium difficile*, and *Escherichia coli*.

353 **Sialic acid:** Family of 9-carbon carboxylated carbohydrates found in vertebrates and some
354 bacteria. Sialic acids are mainly found as terminal monosaccharides on glycan chains and are
355 part of carbohydrate epitopes used as receptors for viruses, such as influenza viruses, and
356 toxins, such as cholera toxin. The main forms of sialic acid found in vertebrates are N-
357 acetylneuraminic acid (NeuAc) and N-glycolylneuraminic acid (NeuGc). Humans have lost the
358 ability to synthesize NeuGc because of inactivating mutations in the *CMAH* gene encoding
359 the cytidine monophosphate-N-acetylneuraminic acid hydroxylase enzyme.

360 **Xenobiotic:** A chemical compound detected in an organism that does not synthesize it.
361 Xenobiotics can mediate pharmacological and endocrine effects that can range from toxic to
362 harmless. Xenobiotics include drugs such as antibiotics and their metabolites, but also
363 environmental pollutants that accumulate through the food chain.

364 **References**

- 365 1 Ip, S. *et al.* (2007) Breastfeeding and maternal and infant health outcomes in developed
366 countries. *Evid Rep Technol Assess (Full Rep)*, 1-186
- 367 2 Horta, B.L. *et al.* (2015) Long-term consequences of breastfeeding on cholesterol,
368 obesity, systolic blood pressure and type 2 diabetes: a systematic review and meta-
369 analysis. *Acta Paediatr* 104, 30-37
- 370 3 Chowdhury, R. *et al.* (2015) Breastfeeding and maternal health outcomes: a systematic
371 review and meta-analysis. *Acta Paediatr* 104, 96-113
- 372 4 World Health Organization (2003) *Global strategy for infant and young child feeding*.
373 WHO Press, Geneva.
- 374 5 Dewey, K.G. (1997) Energy and protein requirements during lactation. *Annu Rev Nutr* 17,
375 19-36
- 376 6 Hautvast, J.G. *et al.* (1989) Recommended dietary allowances for Europe. *Lancet* 2, 1220
- 377 7 Prentice, A.M. *et al.* (1996) Energy requirements of pregnant and lactating women. *Eur J*
378 *Clin Nutr* 50 Suppl 1, S82-110; discussion S110-111
- 379 8 Clark, D.D. and Sokoloff, L. (1999) Circulation and Energy Metabolism of the Brain. In
380 *Basic Neurochemistry: Molecular, Cellular, and Medical Aspects* (6th edn) (Siegel, G.J. *et*
381 *al.*, eds), pp. 637-670, Lippincott
- 382 9 Lemay, D.G. *et al.* (2009) The bovine lactation genome: insights into the evolution of
383 mammalian milk. *Genome Biol* 10, R43
- 384 10 Jenness, R. (1979) The composition of human milk. *Semin Perinatol* 3, 225-239
- 385 11 Insull, W., Jr. *et al.* (1959) The fatty acids of human milk. II. Alterations produced by
386 manipulation of caloric balance and exchange of dietary fats. *J Clin Invest* 38, 443-450

- 387 12 Koletzko, B. *et al.* (2001) Physiological aspects of human milk lipids. *Early Hum Dev* 65
388 Suppl, S3-S18
- 389 13 Weiss, G.A. *et al.* (2013) High levels of anti-inflammatory and pro-resolving lipid
390 mediators lipoxins and resolvins and declining docosahexaenoic acid levels in human
391 milk during the first month of lactation. *Lipids Health Dis* 12, 89
- 392 14 Strous, G.J. and Berger, E.G. (1982) Biosynthesis, intracellular transport, and release of
393 the Golgi enzyme galactosyltransferase (lactose synthetase A protein) in HeLa cells. *J*
394 *Biol Chem* 257, 7623-7628
- 395 15 Brodbeck, U. *et al.* (1967) The isolation and identification of the B protein of lactose
396 synthetase as alpha-lactalbumin. *J Biol Chem* 242, 1391-1397
- 397 16 Ramakrishnan, B. and Qasba, P.K. (2001) Crystal structure of lactose synthase reveals a
398 large conformational change in its catalytic component, the beta1,4-
399 galactosyltransferase-I. *J Mol Biol* 310, 205-218
- 400 17 Turkington, R.W. and Hill, R.L. (1969) Lactose synthetase: progesterone inhibition of the
401 induction of alpha-lactalbumin. *Science* 163, 1458-1460
- 402 18 Skovbjerg, H. *et al.* (1981) Purification and characterisation of amphiphilic
403 lactase/phlorizin hydrolase from human small intestine. *Eur J Biochem* 114, 653-661
- 404 19 Hove, H. *et al.* (1999) Lactic acid bacteria and the human gastrointestinal tract. *Eur J Clin*
405 *Nutr* 53, 339-350
- 406 20 Taya, K. and Greenwald, G.S. (1982) Mechanisms of suppression of ovarian follicular
407 development during lactation in the rat. *Biol Reprod* 27, 1090-1101
- 408 21 Brüssow, H. (2007) *The quest for food : a natural history of eating*. Springer
- 409 22 Coelho, M. *et al.* (2005) Microsatellite variation and evolution of human lactase
410 persistence. *Hum Genet* 117, 329-339

- 411 23 Wen, L. *et al.* (2008) Innate immunity and intestinal microbiota in the development of
412 Type 1 diabetes. *Nature* 455, 1109-1113
- 413 24 Koeth, R.A. *et al.* (2013) Intestinal microbiota metabolism of L-carnitine, a nutrient in
414 red meat, promotes atherosclerosis. *Nat Med* 19, 576-585
- 415 25 Kunz, C. *et al.* (2000) Oligosaccharides in human milk: structural, functional, and
416 metabolic aspects. *Annu Rev Nutr* 20, 699-722
- 417 26 Ninonuevo, M.R. *et al.* (2006) A strategy for annotating the human milk glycome. *J Agric*
418 *Food Chem* 54, 7471-7480
- 419 27 Thurl, S. *et al.* (2010) Variation of human milk oligosaccharides in relation to milk groups
420 and lactational periods. *Br J Nutr* 104, 1261-1271
- 421 28 Marcobal, A. *et al.* (2010) Consumption of human milk oligosaccharides by gut-related
422 microbes. *J Agric Food Chem* 58, 5334-5340
- 423 29 Lewis, Z.T. *et al.* (2015) Maternal fucosyltransferase 2 status affects the gut
424 bifidobacterial communities of breastfed infants. *Microbiome* 3, 13
- 425 30 Fuhrer, A. *et al.* (2010) Milk sialyllactose influences colitis in mice through selective
426 intestinal bacterial colonization. *J Exp Med* 207, 2843-2854
- 427 31 Huang, Y.L. *et al.* (2015) Sialic acid catabolism drives intestinal inflammation and
428 microbial dysbiosis in mice. *Nat Commun* 6, 8141. doi: 8110.1038/ncomms9141.
- 429 32 Ng, K.M. *et al.* (2013) Microbiota-liberated host sugars facilitate post-antibiotic
430 expansion of enteric pathogens. *Nature* 502, 96-99
- 431 33 Tsuruta, T. *et al.* (2010) Development of a method for the identification of S-IgA-coated
432 bacterial composition in mouse and human feces. *Biosci Biotechnol Biochem* 74, 968-
433 973
- 434 34 Palm, N.W. *et al.* (2014) Immunoglobulin A coating identifies colitogenic bacteria in
435 inflammatory bowel disease. *Cell* 158, 1000-1010

- 436 35 Rogier, E.W. *et al.* (2014) Secretory antibodies in breast milk promote long-term
437 intestinal homeostasis by regulating the gut microbiota and host gene expression. *Proc*
438 *Natl Acad Sci U S A* 111, 3074-3079
- 439 36 Adamkin, D.H. (2012) Mother's milk, feeding strategies, and lactoferrin to prevent
440 necrotizing enterocolitis. *JPEN J Parenter Enteral Nutr* 36, 25S-29S
- 441 37 Bullen, J.J. *et al.* (1972) Iron-binding proteins in milk and resistance to *Escherichia coli*
442 infection in infants. *Br Med J* 1, 69-75
- 443 38 Laiho, K. *et al.* (2003) Breast milk fatty acids, eicosanoids, and cytokines in mothers with
444 and without allergic disease. *Pediatr Res* 53, 642-647
- 445 39 Hara, T. *et al.* (1995) Identification of macrophage colony-stimulating factor in human
446 milk and mammary gland epithelial cells. *Pediatr Res* 37, 437-443
- 447 40 Saito, S. *et al.* (1993) Transforming growth factor-beta (TGF-beta) in human milk. *Clin*
448 *Exp Immunol* 94, 220-224
- 449 41 Chowanadisai, W. and Lonnerdal, B. (2002) Alpha(1)-antitrypsin and antichymotrypsin in
450 human milk: origin, concentrations, and stability. *Am J Clin Nutr* 76, 828-833
- 451 42 Hanson, L.A. (1961) Comparative immunological studies of the immune globulins of
452 human milk and of blood serum. *Int Arch Allergy Appl Immunol* 18, 241-267
- 453 43 Brandtzaeg, P. (2007) Induction of secretory immunity and memory at mucosal surfaces.
454 *Vaccine* 25, 5467-5484
- 455 44 Suzuki, Y.A. *et al.* (2001) Molecular cloning and functional expression of a human
456 intestinal lactoferrin receptor. *Biochemistry* 40, 15771-15779
- 457 45 Kuwata, H. *et al.* (1998) Direct evidence of the generation in human stomach of an
458 antimicrobial peptide domain (lactoferricin) from ingested lactoferrin. *Biochim Biophys*
459 *Acta* 1429, 129-141

- 460 46 Lima, M.F. and Kierszenbaum, F. (1987) Lactoferrin effects of phagocytic cell function. II.
461 The presence of iron is required for the lactoferrin molecule to stimulate intracellular
462 killing by macrophages but not to enhance the uptake of particles and microorganisms. *J*
463 *Immunol* 139, 1647-1651
- 464 47 Marks, L.R. *et al.* (2012) The human milk protein-lipid complex HAMLET sensitizes
465 bacterial pathogens to traditional antimicrobial agents. *PLoS One* 7, e43514
- 466 48 Puthia, M. *et al.* (2014) Prevention and treatment of colon cancer by peroral
467 administration of HAMLET (human alpha-lactalbumin made lethal to tumour cells). *Gut*
468 63, 131-142
- 469 49 Ogawa, J. *et al.* (2004) Role of transforming growth factor-beta in breast milk for
470 initiation of IgA production in newborn infants. *Early Hum Dev* 77, 67-75
- 471 50 Oddy, W.H. and Rosales, F. (2010) A systematic review of the importance of milk TGF-
472 beta on immunological outcomes in the infant and young child. *Pediatr Allergy Immunol*
473 21, 47-59
- 474 51 Verhasselt, V. *et al.* (2008) Breast milk-mediated transfer of an antigen induces
475 tolerance and protection from allergic asthma. *Nat Med* 14, 170-175
- 476 52 Berg, D.J. *et al.* (1996) Enterocolitis and colon cancer in interleukin-10-deficient mice are
477 associated with aberrant cytokine production and CD4(+) TH1-like responses. *J Clin*
478 *Invest* 98, 1010-1020
- 479 53 Oz, H.S. *et al.* (2004) Efficacy of a transforming growth factor beta 2 containing
480 nutritional support formula in a murine model of inflammatory bowel disease. *J Am Coll*
481 *Nutr* 23, 220-226
- 482 54 Salzman, N.H. *et al.* (2003) Protection against enteric salmonellosis in transgenic mice
483 expressing a human intestinal defensin. *Nature* 422, 522-526

- 484 55 Labeta, M.O. *et al.* (2000) Innate recognition of bacteria in human milk is mediated by a
485 milk-derived highly expressed pattern recognition receptor, soluble CD14. *J Exp Med*
486 191, 1807-1812
- 487 56 Ruiz-Palacios, G.M. *et al.* (2003) *Campylobacter jejuni* binds intestinal H(O) antigen (Fuc
488 alpha 1, 2Gal beta 1, 4GlcNAc), and fucosyloligosaccharides of human milk inhibit its
489 binding and infection. *J Biol Chem* 278, 14112-14120
- 490 57 Jiang, X. *et al.* (2004) Human milk contains elements that block binding of noroviruses to
491 human histo-blood group antigens in saliva. *J Infect Dis* 190, 1850-1859
- 492 58 Naarding, M.A. *et al.* (2005) Lewis X component in human milk binds DC-SIGN and
493 inhibits HIV-1 transfer to CD4+ T lymphocytes. *J Clin Invest* 115, 3256-3264
- 494 59 Velupillai, P. and Harn, D.A. (1994) Oligosaccharide-specific induction of interleukin 10
495 production by B220+ cells from schistosome-infected mice: a mechanism for regulation
496 of CD4+ T-cell subsets. *Proc Natl Acad Sci U S A* 91, 18-22
- 497 60 Kurakevich, E. *et al.* (2013) Milk oligosaccharide sialyl(α 2,3)lactose activates intestinal
498 CD11c+ cells through TLR4. *Proc Natl Acad Sci U S A* 110, 17444-17449
- 499 61 Pritchard, D.G. *et al.* (1992) Murine monoclonal antibodies to type Ib polysaccharide of
500 group B streptococci bind to human milk oligosaccharides. *Infect Immun* 60, 1598-1602
- 501 62 Kuijf, M.L. *et al.* (2010) TLR4-mediated sensing of *Campylobacter jejuni* by dendritic cells
502 is determined by sialylation. *J Immunol* 185, 748-755
- 503 63 Vimr, E.R. *et al.* (2004) Diversity of microbial sialic acid metabolism. *Microbiol Mol Biol*
504 *Rev* 68, 132-153
- 505 64 Quinn, E.A. *et al.* (2015) Maternal characteristics associated with milk leptin content in a
506 sample of Filipino women and associations with infant weight for age. *J Hum Lact* 31,
507 273-281

508 65 Baxter, R.C. *et al.* (1984) Immunoreactive somatomedin-C/insulin-like growth factor I
509 and its binding protein in human milk. *J Clin Endocrinol Metab* 58, 955-959

510 66 Martin, L.J. *et al.* (2006) Adiponectin is present in human milk and is associated with
511 maternal factors. *Am J Clin Nutr* 83, 1106-1111

512 67 Grey, K.R. *et al.* (2013) Human milk cortisol is associated with infant temperament.
513 *Psychoneuroendocrinology* 38, 1178-1185

514 68 Catalani, A. *et al.* (2000) Maternal corticosterone during lactation permanently affects
515 brain corticosteroid receptors, stress response and behaviour in rat progeny.
516 *Neuroscience* 100, 319-325

517 69 Cima, I. *et al.* (2004) Intestinal epithelial cells synthesize glucocorticoids and regulate T
518 cell activation. *J Exp Med* 200, 1635-1646

519 70 Carson, R. (1962) *Silent spring*. Fawcett Crest

520 71 Longnecker, M.P. *et al.* (2001) Association between maternal serum concentration of
521 the DDT metabolite DDE and preterm and small-for-gestational-age babies at birth.
522 *Lancet* 358, 110-114

523 72 Cohn, B.A. *et al.* (2015) DDT Exposure in Utero and Breast Cancer. *J Clin Endocrinol*
524 *Metab* 100, 2865-2872

525 73 Noren, K. and Meironyte, D. (2000) Certain organochlorine and organobromine
526 contaminants in Swedish human milk in perspective of past 20-30 years. *Chemosphere*
527 40, 1111-1123

528 74 Albert, O. and Jegou, B. (2014) A critical assessment of the endocrine susceptibility of
529 the human testis to phthalates from fetal life to adulthood. *Hum Reprod Update* 20,
530 231-249

531 75 Main, K.M. *et al.* (2006) Human breast milk contamination with phthalates and
532 alterations of endogenous reproductive hormones in infants three months of age.
533 *Environ Health Perspect* 114, 270-276

534 76 Minamishima, I. *et al.* (1994) Role of breast milk in acquisition of cytomegalovirus
535 infection. *Microbiol Immunol* 38, 549-552

536 77 The Working Group on Mother-To-Child Transmission of HIV (1995) Rates of mother-to-
537 child transmission of HIV-1 in Africa, America, and Europe: results from 13 perinatal
538 studies. *J Acquir Immune Defic Syndr Hum Retrovirol* 8, 506-510

539 78 Centers for Disease Control (1985) Recommendations for assisting in the prevention of
540 perinatal transmission of human T-lymphotropic virus type III/lymphadenopathy-
541 associated virus and acquired immunodeficiency syndrome. *MMWR Morb Mortal Wkly*
542 *Rep* 34, 721-726, 731-722

543 79 Badinter, E. and Hunter, A. (2011) *The conflict : how modern motherhood undermines*
544 *the status of women*. Metropolitan Books/Henry Holt and Co.

545

546 **Figure legends**

547 **Figure 1.** Lactose biosynthesis and degradation. Lactose synthase (PDB: 1nhe [16]) is a
548 heterodimer comprising the β 1-4 galactosyltransferase B4GALT1 and α -lactalbumin. The
549 pituitary hormone prolactin stimulates the expression of α -lactalbumin in the lactating
550 mammary gland. In the small intestine of the suckling infant, lactose is cleaved by lactase,
551 which expression is age-dependent. The resulting monosaccharides glucose (Glc) and
552 galactose (Gal) are absorbed by the sodium-glucose linked transporter SGLT1. In the absence
553 of lactase, lactose reaches the colon where it is degraded by intestinal microbes. The
554 increase of bacterial fermentation products causes abdominal cramps, bloating and nausea,
555 which leads to cessation of breastfeeding.

556 **Figure 2.** Biosynthetic pathway of milk oligosaccharides. The lactose core (boxed structure) is
557 modified by addition of fucose (Fuc), sialic acid (Sia), N-acetylglucosamine (GlcNAc) and
558 galactose (Gal). The most common breast milk trisaccharides are fucose(α 1-2)lactose (2FL),
559 fucose(α 1-3)lactose (3FL), sialyl(α 2-3)lactose (3SL), and sialyl(α 2-6)lactose (6SL). In human
560 milk, the most common tetrasaccharide is lacto-N-tetraose (LNT), whereas lacto-N-
561 neotetraose (LNnT) dominates in other mammalian milks. In human milk, Lewis antigens (LeA,
562 LeB, LeX, LeY, sLeA, sLeX) are epitopes (blue shaded structures) frequently found on milk
563 oligosaccharides. A and B blood group antigens are absent in human milk oligosaccharides,
564 but O antigen type I (H1) and type II (H2) are common. The LNT and LNnT cores can be
565 further elongated (dashed arrows) to yield oligosaccharides consisting of more than 20
566 monosaccharides.

567 **Figure 3.** Immune-active compounds of breast milk. Human breast milk delivers cytokines
568 (cyan circles), such as TGF β , IL-10, and M-CSF, soluble CD14 (sCD14, blue triangles),
569 lactoferrin (red circle), human milk oligosaccharides (HMO), and secreted IgA (sIgA) to the

570 infant gut. Upon resorption by the intestinal mucosa, sCD14 contributes to innate immune
571 protection by recognizing microbe-associated molecular patterns. Milk lactoferrin, sIgA and
572 Paneth cell-derived defensins (orange circles) prevent bacterial (grey rods) overgrowth.
573 HMO act as receptor decoys, inhibiting bacterial adhesion to mucosal surfaces. HMO can be
574 also taken up by M-cells (blue cells) in Peyer's patches, which may contribute to the
575 induction of tolerogenic responses toward structurally-related mucosal glycans. In the
576 intestinal mucosa, dendritic cells (DC), macrophages (MΦ), T lymphocytes (T), B lymphocytes
577 (B) and plasma cells (PC) orchestrate mucosal immunity through the secretion of cytokines
578 (green circles).