



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2015

PYY3–36: Beyond food intake

Stadlbauer, Ulrike ; Woods, Stephen C ; Langhans, Wolfgang ; Meyer, Urs

DOI: <https://doi.org/10.1016/j.yfrne.2014.12.003>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-120370>

Journal Article

Accepted Version



The following work is licensed under a Creative Commons: Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.

Originally published at:

Stadlbauer, Ulrike; Woods, Stephen C; Langhans, Wolfgang; Meyer, Urs (2015). PYY3–36: Beyond food intake. *Frontiers in Neuroendocrinology*, 38:1-11.

DOI: <https://doi.org/10.1016/j.yfrne.2014.12.003>

PYY₃₋₃₆: Beyond food intake

Ulrike Stadlbauer¹, Stephen C. Woods², Wolfgang Langhans¹, Urs Meyer¹

¹Physiology and Behavior Laboratory, ETH Zurich, Switzerland.

²Department of Psychiatry and Behavioral Neuroscience, University of Cincinnati,
Cincinnati, Ohio, USA.

Correspondence:

Dr. Ulrike Stadlbauer

Physiology and Behavior Laboratory
Schorenstrasse 16, 8603 Schwerzenbach, Switzerland

E-mail: ulrike-stadlbauer@ethz.ch

Tel: +41 44 655 7486; *Fax:* +41 44 655 7206

Running title: Behavioral effects of PYY₃₋₃₆

Number of manuscript pages: 41

Number of tables: 1

Number of figures: 3

Abstract

The gastrointestinal hormone peptide tyrosine tyrosine 3-36 (PYY₃₋₃₆) has attained broad recognition with respect to its involvement in energy homeostasis and the control of food intake. It is mainly secreted by distal intestinal enteroendocrine L-cells in response to eating and exerts both neurally mediated paracrine and endocrine effects on various target organs. In addition to its gastrointestinal effects, PYY₃₋₃₆ has long been known to inhibit food intake. Recent closer examination of the effects of PYY₃₋₃₆ revealed that this gut-derived peptide also influences a wide spectrum of behavioral and cognitive functions that are pivotal for basic processes of perception and judgment, including central information processing, salience learning, working memory, and behavioral responding to novelty. Here, we review the effects of PYY₃₋₃₆ that go beyond food intake and provide a conceptual framework suggesting that several apparently unrelated behavioral actions of PYY₃₋₃₆ may actually reflect different manifestations of modulating the central dopamine system.

Key words: Dopamine; energy homeostasis; eating; incentive salience; gut peptide.

1. Introduction

Peptide tyrosine tyrosine (PYY) is a peptide hormone which, together with pancreatic polypeptide (PP) and neuropeptide Y (NPY), comprises the PP family of peptides (Berglund et al., 2003). The two existing forms of PYY differ by two amino acids (Grandt et al., 1994; Medeiros and Turner, 1994). PYY₁₋₃₆ is released from enteroendocrine L-cells in response to nutrient signals in the chyme. In the blood, PYY₁₋₃₆ is rapidly converted to PYY₃₋₃₆ by the ubiquitously expressed enzyme, dipeptidyl-peptidase IV (DPP-IV), which cleaves the two N-terminal amino acids (Mentlein et al., 1993). Hence, PYY₃₋₃₆ is the major circulating form of the peptide, known to exert different and sometimes opposite biological functions than PYY₁₋₃₆ (Grandt et al., 1994) (**Figure 1**).

As extensively reviewed elsewhere (Karra and Batterham, 2010; Schwartz and Holst, 2010; Walther et al., 2011), the distinct biological functions exerted by PYY₁₋₃₆ and PYY₃₋₃₆ have been explained by their different binding affinities for the five Y receptor subtypes in mammals, Y1, Y2, Y4, Y5 and Y6. All are inhibitory G-protein coupled receptors that reduce cyclic-AMP and the mobilization of intracellular calcium (Michel et al., 1998; Berglund et al., 2003). Whereas PYY₁₋₃₆ has similar affinities for the Y1 and Y2 receptor, PYY₃₋₃₆ is a high-affinity Y2 receptor ligand (Walther et al., 2011). In the periphery, the Y2 receptor is expressed by parasympathetic and sympathetic sensory neurons, in addition to intestinal and some vascular cells (Widdowson, 1993; Gehlert, 1994; Cabrele and Beck-Sickinger, 2000). The Y2 receptor is also abundantly expressed in several regions of the central nervous system (CNS), including limbic and cortical areas (Stanic et al., 2006; Walther et al., 2011). In neuronal tissue, the Y2 receptor is localized mainly presynaptically, inhibiting neurotransmitter release upon activation (Smith-White et al., 2001; Stanic et al., 2011). Such autoreceptor functions of the Y2 receptor are well documented, for example with regard to NPY release in hypothalamic

areas, where Y2 receptor agonists including PYY₃₋₃₆ inhibit NPY synthesis and secretion (King et al, 1999; Smith-White et al, 2001; Batterham et al, 2002; Challis et al, 2003).

PYY is secreted by mainly distal intestinal enteroendocrine L-cells in response to eating, and plasma levels of PYY₃₋₃₆ remain elevated for several hours after meals (Adrian et al., 1985; Stanley et al., 2004). The best known functions of PYY₃₋₃₆ are in the gastrointestinal system where it regulates secretions (Yang, 2002) and motility (Imamura, 2002). Many of its actions contribute to the 'ileal brake,' whereby secretions of the distal small intestine slow gastric emptying when nutrients reach the ileum.

More recently, PYY₃₋₃₆ has attained broad recognition with respect to its involvement in energy homeostasis and the control of food intake (see Manning and Batterham, 2014). A landmark study by Batterham et al. (2002) directly implicated PYY₃₋₃₆ in the physiological inhibition of food intake. This effect is mediated through the Y2 receptor (Batterham et al., 2002) and has been documented in diverse conditions and several species, including rodents and humans (**Table 1**). Basal levels are lower and the meal-induced release of PYY₃₋₃₆ is blunted in obese individuals (Alvarez Bartolomé et al., 2002; Batterham et al., 2003; le Roux et al., 2006; Guo et al., Sodowski et al., 2007). Also, PYY overexpression protects against diet-induced obesity (Boey et al., 2008). Importantly, PYY₃₋₃₆ administration reduces food intake similarly in obese and non-obese subjects (Batterham et al., 2003; Sloth et al., 2007), implying that obesity does not decrease PYY₃₋₃₆ sensitivity. Collectively, these observations have attracted considerable interest in PYY₃₋₃₆ as a potential pharmacotherapy for obesity (Karra et al., 2009).

While the precise physiological mechanisms whereby PYY₃₋₃₆ inhibits eating remain unclear, it effectively crosses the blood-brain-barrier from the plasma (Nonaka et al., 2003) and acts centrally as a relatively selective Y2 receptor agonist (Grandt et al.,

1994). Y2 receptor expression is abundant on hypothalamic arcuate neurons that co-express NPY and agouti-related peptide (Agrp) (Broberger et al., 1997; Hahn et al., 1998), and administration of PYY₃₋₃₆ directly into the Arc reduces food intake (Batterham, 2002). Consistent with its action as an inhibitory presynaptic receptor, one prevalent hypothesis suggests that activation of the Y2 receptor inhibits arcuate NPY neurons and reduces the NPY-mediated inhibition of neighboring anorexigenic neurons co-expressing pro-opiomelanocortin (POMC) and cocaine- and amphetamine-regulated transcript (CART) (Broberger et al., 1997; Morton et al., 2014).

In addition to hypothalamic sites of action, there are also alternative (but not mutually exclusive) mechanisms by which PYY₃₋₃₆ could inhibit food intake, particularly in light of the widespread expression of Y2 receptors in cortical and subcortical brain areas (Stanic et al., 2006, 2011). Hence, in addition to NPY neurons, some γ -aminobutyric acid (GABA) or glutamate neurons also express the Y2 receptor (Stanic et al., 2006, 2011). Y2 agonists such as PYY₃₋₃₆ may thus readily influence neural circuits in diverse brain regions. Consistent with this, using functional magnetic resonance imaging (fMRI), Batterham and colleagues found that peripheral administration of PYY₃₋₃₆ induces neuronal activation in several brain regions, including target areas of the mesolimbic and nigrostriatal dopaminergic pathways, brainstem areas including the nucleus tractus solitarii (NTS), and cortical areas including the orbitofrontal cortex (Batterham et al., 2007). Consistent with these findings, our research group also observed widespread neuronal activation following peripheral PYY₃₋₃₆ administration in rats (Stadlbauer et al., 2013a; for further details, see Section 5).

In view of these neuronal effects, it is reasonable to hypothesize that PYY₃₋₃₆ has functional significance in the brain beyond its role in controlling food intake, and experimental research in rodents has recently begun to explore the effects of PYY₃₋₃₆ on

other behaviors. Here, we summarize some of those findings and provide a conceptual framework suggesting that several apparently unrelated behavioral actions of PYY₃₋₃₆ actually reflect different manifestations of modulating the mesocorticolimbic dopamine system.

2. PYY₃₋₃₆ and sensitivity to psychostimulant drugs

Studies in both humans and rats indicate that the peripheral administration of PYY₃₋₃₆ leads to activation of central dopaminergic pathways (Batterham et al., 2007; Stadlbauer et al., 2013a). The largest populations of dopamine cells are localized in two neighboring midbrain nuclei, namely the ventral tegmental area (VTA; A10 cell group) and the substantia nigra (SN; A9 cell group) (Tzschentke, 2001; Björklund and Dunnett, 2007; Van den Heuvel and Pasterkamp, 2008). The majority of VTA dopamine cells projects to limbic and cortical areas along the mesolimbic and mesocortical dopamine pathways, respectively, whereas a large part of the nigral A9 dopamine cells innervate the dorsal striatum forming the nigrostriatal dopaminergic pathway (**Figure 2**). Midbrain dopamine cells are also found in the A8 cell group, which forms a dorsal and caudal extension of the A9 cell group and contains cells that project to both striatal, limbic and cortical areas (**Figure 2**). A8 cells are thus an integral part of the mesolimbic, mesocortical, and nigrostriatal dopamine pathways (Björklund and Dunnett, 2007; Roeper, 2013).

Among other functions, the mesolimbic dopaminergic pathway is important in mediating the behavioral and locomotor responses to drugs of abuse (Soderpalm and Ericson, 2013), whereas the nigrostriatal pathway is critically involved in the control of voluntary movement and motor stereotypies (Groenewegen, 2003). Recent neuropharmacological investigations in mice demonstrate that peripheral

administration of PYY₃₋₃₆ markedly modulates these dopamine-related behavioral functions (Stadlbauer et al., 2014). Specifically, pretreatment with PYY₃₋₃₆ potentiates the locomotor responses to subsequent amphetamine (Amph) exposure and increases stereotypical behavioral reactions to systemic apomorphine (Apo) (Stadlbauer et al., 2014). Amph is an indirect dopamine receptor agonist that efficiently stimulates presynaptic dopamine release (Salahpour et al., 2008), and its administration elicits rigorous locomotor activity (Robinson and Becker, 1986). Apo is a preferential dopamine D1/D2 receptor agonist that dose-dependently increases locomotor activity and other stereotyped behaviors in rodents, including repetitive climbing and leaning (Cabib and Puglisi-Allegra, 1985; Bitanhirwe et al., 2010). The mesolimbic and nigrostriatal dopamine pathways are key neuronal components mediating the behavioral responses to Amph and Apo. Early studies concluded that the locomotor-enhancing effects of low doses of systemic Amph result from increased dopamine transmission in the NAc (Creese and Iversen, 1975; Pijnenburg et al., 1976), particularly in its shell sub-region (Heidbreder and Feldon, 1998). More recent studies suggest that enhanced dopamine release more dorsally in the striatum contributes to Amph-induced locomotor hyperactivity as well (Matthews et al., 2013). The expression of stereotyped behaviors has also often been functionally linked to enhanced activation of striatal dopamine receptors, especially in dorsal parts of the striatum (Arnt et al., 1988; Vasse and Protais, 1989; Charntikov et al., 2011). It has recently been found that PYY₃₋₃₆ potentiates the behavioral responses to both Amph and Apo and that it likely involves increased dopaminergic activity in the mesolimbic and/or nigrostriatal pathways (Stadlbauer et al., 2014). Even though this hypothesis lacks direct confirmation, it is consistent with previous ex-vivo studies reporting that exogenous PYY₃₋₃₆ increases the synthesis and release of dopamine in rat striatal slices (Adewale et al., 2005, 2007).

Work in genetically modified mice lacking the Y2 receptor has provided additional support for the hypothesis that signaling through Y2 receptors can exert a direct influence on striatal dopamine release (Zambello et al., 2011). Thus, accumulating evidence suggests that PYY₃₋₃₆ administration induces neuronal (Batterham et al., 2007; Stadlbauer et al., 2013a), behavioral (Stadlbauer et al., 2014), and neurochemical (Adewale et al., 2005, 2007; Zambello et al., 2011) changes reminiscent of a (transient) hyperdopaminergic state.

3. PYY₃₋₃₆ and behavioral responses to novelty

Responding to a novel environment engages the mesolimbic dopamine system (Bardo et al., 1996; Blanchard et al., 2009), and the magnitude of the response predicts the behavioral responses to dopaminergic psychostimulant drugs (Marinelli and White, 2000). Given these associations, it has been hypothesized that PYY₃₋₃₆, in addition to its effects on potentiating psychostimulant drug sensitivity, would also enhance novelty seeking. In support of this, we observed that peripheral administration of PYY₃₋₃₆ in mice increases novelty seeking in a novel-object exploration task in which mice were allowed to freely explore an unfamiliar object following habituation to the surrounding context (Stadlbauer et al., 2014). These effects were unlikely to be mediated by possible changes in anxiety-like behavior because identical PYY₃₋₃₆ treatment did not affect behavioral indices of innate anxiety (Stadlbauer et al., 2013b, 2014).

More likely is that the enhancement of novel object exploration displayed by PYY₃₋₃₆-treated animals is related to changes in incentive salience. Incentive salience is a motivational attribute that increases the attractiveness of a given stimulus and promotes approach behavior towards it (Berridge and Robinson, 1998). Research in rats has found a positive correlation between the amount of novelty seeking and incentive

salience attribution to reward-associated cues (Beckmann et al., 2011). Dopaminergic mechanisms in general, and increased accumbal dopaminergic activity in particular, are critical in regulating the perception and processing of salient stimuli (Berridge and Robinson, 1998; Wise, 2004). For example, manipulations increasing and decreasing dopaminergic activity in the NAc, respectively, enhance and reduce exploratory activity toward novel stimuli (Rebec et al., 1997; Peters et al., 2007; Fukushiro and Frussa-Filho, 2011; Laricchiuta et al., 2014). Furthermore, rats with increased novelty-seeking have a greater behavioral sensitivity to the indirect dopamine receptor agonist cocaine (Beckmann et al., 2011). Similar parallels exist following peripheral PYY₃₋₃₆ administration in mice, where PYY₃₋₃₆ elicits a concomitant increase in the behavioral response to novelty and to Amph (Stadlbauer et al., 2014). Thus, the positive correlations among mesolimbic dopamine activity, novelty seeking, and incentive salience (Bardo et al., 1996; Berridge and Robinson, 1998; Blanchard et al., 2009; Beckmann et al., 2011) all suggest that PYY₃₋₃₆-induced potentiation of novelty seeking likely involves increased incentive salience attribution to the novel stimuli.

The same processes may also explain the recently reported decreases in social approach behavior following peripheral PYY₃₋₃₆ administration in mice (Stadlbauer et al., 2013b). When given the choice between exploring an unfamiliar mouse and a novel inanimate object, mice (like most other rodents) typically prefer spending more time with the live mouse relative to the inanimate object (Moy et al. 2008; Vuillermot et al., 2011). Following PYY₃₋₃₆ treatment, however, the preference is no longer seen, and PYY₃₋₃₆-treated mice spend more time with the novel object at the expense of reduced time spent with the live mouse (Stadlbauer et al., 2013b). Consistent with this, genetic ablation or pharmacological inhibition of the Y2 receptor causes an opposite pattern of effects, including increased social approach behavior (Karl et al., 2010; Morales-Medina

et al, 2012). Hence, stimulation or attenuation of Y2 receptor signaling reduces or increases social approach behavior, respectively, and these effects may at least partially involve altered incentive salience attribution to unfamiliar congenic species and novel inanimate objects.

4. PYY₃₋₃₆ and central information processing

Aberrant salience processing is also involved in the disruption of central information processing, especially when the brain is required to discriminate between relevant and irrelevant stimuli (Smith et al., 2006; Winton-Brown et al., 2014). Under such conditions, increased mesolimbic dopamine activity enhances the salience of irrelevant stimuli, and as a consequence the organism often fails to differentiate between relevant and irrelevant information (Kapur, 2003; Smith et al., 2006). The essence of this phenomenon can be captured by a behavioral paradigm known as latent inhibition (LI), a model of associative learning in which non-reinforced pre-exposures to a to-be-conditioned stimulus (CS) retard subsequent conditioning between the same CS and the unconditioned stimulus (US) (Lubow and Moore, 1959; Lubow, 2005). Prevalent neuropsychological theories posit that LI is caused by the development of selective attention away from the pre-exposed stimulus, so that non-reinforced CS pre-exposure diminishes the perceived salience of the CS during conditioning (Mackintosh, 1975; Lubow et al., 1981; for other neuropsychological theories, see Weiner, 2003 and Lubow, 2005). LI is often referred to as a form of “salience learning” (Young et al., 2005; Nelson et al., 2011), and its expression is taken as index of the tendency of an organism to successfully ignore stimuli that historically predict no significant consequences (Weiner, 2003). Aberrant salience attribution to inconsequential stimuli weakens LI, and is indicative of a susceptibility to distraction by irrelevant information.

Similar types of central information processing can also be assessed using behavioral paradigms that do not involve explicit associative learning processes. One widely used example is prepulse inhibition (PPI) of the acoustic startle reflex, which is the reduction of a startle reaction to a startle-eliciting stimulus (pulse) when it is shortly preceded by a weak stimulus (prepulse) (Hoffman and Searle, 1965; Braff et al., 2001). PPI provides an operational measure of sensorimotor gating, in which central gating mechanisms protect the processing of the information contained in the initial prepulse from distraction by the subsequent pulse stimulus (Graham, 1975; Braff et al., 2001). PPI thus serves to filter or gate intrusive sensorimotor information. Disruption of such gating mechanisms can lead to central stimulus overload and associated dysfunctions in allocating the limited neuronal resources to only the most important stimuli encountered in the environment (Swerdlow et al., 2000; Braff et al., 2001).

As extensively reviewed elsewhere (Swerdlow et al, 2000; Braff et al., 2001; Weiner, 2003; Lubow, 2005; Young et al., 2005), experimental manipulations or pathological conditions that result in increased mesolimbic dopamine activity disrupt both LI and PPI. Weakening of PPI and LI can arise from manipulations that directly target the central dopamine system, such as administering Amph or Apo (Swerdlow et al, 2000; Braff et al., 2001; Weiner, 2003; Lubow, 2005; Young et al., 2005). Alternatively, PPI and LI deficiency can also be induced by manipulations that do not primarily target the central dopamine system, but instead lead to down-stream increases in mesolimbic dopamine signaling (Meyer and Feldon, 2009; Peleg-Raibstein et al., 2012). Hence, increased mesolimbic dopamine signaling is a common neurochemical mechanism for the disruption of PPI and LI, regardless of whether the experimental manipulation or pathological condition directly or indirectly affects the mesolimbic dopamine pathways.

We have recently found that that acute peripheral PYY₃₋₃₆ treatment markedly reduces sensorimotor gating in the form of PPI and salience learning in the form of LI (Stadlbauer et al., 2013b). Intriguingly, the dopamine receptor antagonist haloperidol is effective in blocking PYY₃₋₃₆-induced PPI disruption (Stadlbauer et al., 2013b), implying that the weakening of central information processing by PYY₃₋₃₆ is mediated by increased dopamine signaling. This is consistent with the report that ablation of the Y2 receptor leads to enhanced PPI (Karl et al., 2010). Thus, increased Y2 activity reduces sensorimotor gating, likely via increasing dopaminergic activity in mesolimbic pathways.

Whether or not PYY₃₋₃₆-induced attenuation of LI is similarly dependent on dopaminergic mechanisms awaits examination. PYY₃₋₃₆-induced loss of LI, however, was found to arise from selective effects in the subgroup of animals that had been pre-exposed to the CS before conditioning (Stadlbauer et al., 2013b), suggesting that exogenous PYY₃₋₃₆ is able to abolish the efficacy of repeated CS pre-exposures to reduce the expression of the conditioned response. As a consequence, pre-exposed animals treated with PYY₃₋₃₆ no longer display the typical reduction in the conditioned response as seen in non-treated pre-exposed animals (Stadlbauer et al., 2013b). An important implication is that the PYY₃₋₃₆-induced disruption of LI does not simply reflect a general deficit in classical conditioning per se, but rather readily mirrors deficits in salience learning that normally regulate the expression of LI (Weiner, 2003; Lubow, 2005; Young et al., 2005; Nelson et al., 2011). While still hypothetical, these findings are compatible with a neuropsychological model, in which PYY₃₋₃₆ can enhance the salience of irrelevant stimuli through neurochemical processes involving increased mesolimbic dopaminergic activity (Kapur, 2003; Smith et al., 2006).

5. PYY₃₋₃₆ and cognition

Signaling at the Y2 receptor has been further implicated in certain types of learning and memory (Borbély et al., 2013), with most data suggesting that activation of the Y2 receptor has beneficial effects on long-term memory. For example, Redrobe et al. (2004) found that mice lacking the Y2 receptor have a selective impairment in long-term retention of spatial memory and long-term memory for objects. Similar effects were observed following acute pharmacological blockade of the Y2 receptor in mice (dos Santos et al., 2013). Y2 receptor signaling has also been implicated in short-term memory, but in contrast to its detrimental effects on long-term memory, attenuation of Y2 signaling exerts beneficial effects on short-term memory (Gonçalves et al., 2012). This “double-edged sword” effect of facilitating long-term memory but impeding short-term memory is consistent with a dual-process model of memory, in which short-term and long-term memory are separate and sometimes competing processes (Sanderson et al., 2009; Sanderson and Bannerman, 2012).

When we directly examined the effects of PYY₃₋₃₆ on learning and memory, we found that intraperitoneal PYY₃₋₃₆ administration markedly impaired working memory in mice (Stadlbauer et al., 2013b). Working memory refers to a short-term memory buffer used to hold relevant information temporarily active in order to guide on-going behavior (Baddeley, 2003). Hence, the negative influence of PYY₃₋₃₆ on working memory is consistent with the concept that activation of Y2 receptor signaling impedes short-term forms of memory (Gonçalves et al., 2012).

Successful performance in working memory tests depends on several factors. First, the test subject must allocate appropriate attention to the relevant stimuli, both during the initial acquisition (learning) trial and subsequent expression (memory) trials. Second, the subject must retrieve the relevant short-term information based on its

previous action during the acquisition pause in order to effectively complete the task on a subsequent memory trial. This cognitive demand is further dependent on the amount of experienced proactive interference, which occurs when cognitive processing during (multiple) acquisition trials negatively affects performance on subsequent test trials (Hartshorne, 2008). Hence, there are several potential neurocognitive mechanisms by which PYY₃₋₃₆ could disrupt working memory. In view of the marked effects of PYY₃₋₃₆ on salience processing and selective attention (see Section 4), it seems feasible that attentional deficits are involved. This interpretation would also be consistent with recent reports that working memory performance positively correlates with central information processing capacity (Singer et al., 2013), both of which are reduced by peripheral PYY₃₋₃₆ administration in mice (Stadlbauer et al., 2013b).

6. Modulation of GABA-dopamine interactions by PYY₃₋₃₆: A common pathway for diverse behavioral changes?

As detailed above, studies from PYY₃₋₃₆-treated mice (Stadlbauer et al., 2013b, 2014), complemented by studies using Y2 receptor-deficient mice or preferential Y2 receptor antagonists (Redrobe et al., 2004; Karl et al., 2010; Zambello et al., 2011; Gonçalves et al., 2012; Morales-Medina et al., 2012), document that PYY₃₋₃₆ modulates behavioral and cognitive activities in addition to simply reducing food intake. An important question is whether the diverse repertoire of neurobehavioral and neurocognitive changes involves different neuronal and neurochemical processes, or whether it can be explained by a common neuronal mechanism.

In support of the latter, many of the behavioral functions influenced by PYY₃₋₃₆ are critically regulated by subcortical dopamine activity. These include Amph-induced locomotor hyperactivity (Robinson and Becker, 1986; Heidbreder and Feldon, 1998),

Apo-induced behavioral stereotypies (Arnt et al., 1988; Vasse and Protais, 1989; Charntikov et al., 2011), novelty seeking (Bardo et al., 1996; Berridge and Robinson, 1998; Blanchard et al., 2009), sensorimotor gating (Swerdlow et al., 2000, 2001; Braff et al., 2001), and selective attention and salience learning (Weiner, 2003; Young et al., 2005), all of which are changed by peripheral PYY₃₋₃₆ administration (Stadlbauer et al., 2013b; 2014). Hence, the central pro-dopaminergic effects of PYY₃₋₃₆ may provide a common mechanism underlying the induction of different behavioral alterations. This interpretation fits with the general proposition that a core disruption in a key neurotransmitter system can give rise to diverse behavioral and cognitive alterations (Meyer and Feldon, 2009, 2010). Consistent with this, many clinical behavioral disorders involve abnormally enhanced dopamine activity (Kapur, 2003; Winton-Brown et al., 2014).

Given the importance of dopamine for so many behaviors, and the observation that PYY₃₋₃₆ influences dopamine functioning, the determination of how PYY₃₋₃₆ influences dopamine signaling is an important goal. Available data suggest that PYY₃₋₃₆ increases striatal dopamine release via a presynaptic modulation of dopamine release in target areas rather than from a direct action on midbrain dopamine cell activity per se (**Figure 3**). Double immunoenzyme staining of the neuronal early gene product c-Fos and the dopaminergic marker tyrosine hydroxylase (TH) revealed that peripheral PYY₃₋₃₆ treatment does not activate TH-positive dopamine cells in the VTA or SNc (Stadlbauer et al., 2014). It does, however, induce neuronal activation in ventral (NAc) and dorsal (CPu) parts of the striatum (Stadlbauer et al., 2014), which are the two primary areas innervated by VTA and SNc dopaminergic neurons (Van den Heuvel and Pasterkamp, 2008). Hence, striatal neuronal activation following PYY₃₋₃₆ treatment can emerge in the absence of direct activation of the mesoaccumbal (VTA to NAc) or nigrostriatal (SNc to

CPu) dopaminergic pathways. Consistent with this, ex-vivo pharmacological studies in rat brain striatal slices demonstrated that PYY₃₋₃₆ increases dopamine release even though the dopaminergic axon terminals were disconnected from their cell bodies (Adewale et al., 2007).

Y2 receptors are localized presynaptically where they inhibit neurotransmitter release (Smith-White et al., 2001; Stanic et al., 2006). This has been well documented for hypothalamic NPY release, where Y2 agonists including PYY₃₋₃₆ inhibit NPY synthesis and secretion (King et al, 1999; Smith-White et al, 2001; Batterham et al, 2002; Challis et al, 2003). The Y2 receptor is also abundantly expressed in the striatum and many other subcortical structures (Stanic et al., 2006). Interestingly, however, this expression seems to be restricted to non-dopaminergic cells and fibers, as presynaptic dopaminergic terminals lack a clear expression of Y2 receptors (Stanic et al., 2011). The implication is that any modulation of striatal dopamine release by PYY₃₋₃₆ is unlikely to involve direct Y2 signaling at dopaminergic fibers. Instead, it may be largely driven by other neurotransmitter systems that are functionally connected to presynaptic dopamine terminals. In particular, PYY₃₋₃₆ may activate Y2 receptors expressed on striatal GABAergic interneurons, which in turn can robustly attenuate striatal dopamine release by providing inhibitory inputs to presynaptic dopamine terminals (Smith and Kieval, 2000; David et al., 2005). Since activation of Y2 receptors induces neuronal inhibition, it can be expected that PYY₃₋₃₆-induced activation of these receptors inhibits neuronal activity of striatal GABAergic interneurons (Acuna-Goycolea et al., 2005; see also **Figure 3**). Such inhibition would, in turn, weaken the inhibitory inputs of striatal GABAergic interneurons onto presynaptic dopamine terminals, thereby facilitating the release of dopamine (**Figure 3**). Hence, PYY₃₋₃₆ may induce its pro-dopaminergic effects by

weakening the fast-forward inhibition of presynaptic dopaminergic fibers by striatal GABAergic interneurons.

Consistent with this, both human imaging studies (Batterham et al., 2007) and immunohistochemical findings in mice (Stadlbauer et al., 2014) indicate that PYY₃₋₃₆ induces neuronal activation in striatal areas, and leads to increased neuronal activity in down-stream brain areas that are directly innervated by striatal neurons, including the ventral palladium (VP) (Stadlbauer et al., 2014). The VP is a primary projection site of the ventral striatum (Groenewegen et al., 1996; Groenewegen, 2003) and has been implicated in behavioral functions that are affected by exogenous PYY₃₋₃₆ treatment, including sensorimotor gating (Kodsi and Swerdlow, 1995; Kretschmer and Koch, 1998), behavioral sensitivity to dopamine-stimulating drugs such as Amph (Swerdlow and Koob, 1987; Mele et al., 1998), and incentive salience attribution (Tindell et al., 2005). The general consensus is that these behavioral and neuropsychological functions are markedly affected by increased VP activity similarly to what has been observed following peripheral PYY₃₋₃₆ administration (Stadlbauer et al., 2013b, 2014).

It is likely that PYY₃₋₃₆-induced suppression of GABAergic activity is not restricted to striatal areas (Acuna-Goycolea et al., 2005), which in turn may have functional relevance as well. For example, working memory is dependent on the integrity of GABAergic signaling, especially in cortical structures such as the PFC (Lewis et al., 2005; Lewis and Moghaddam, 2006). GABA-mediated inhibition is an essential component in the synchronization of neuronal rhythms and oscillatory activity (Lewis et al., 2005; Kohl and Paulsen, 2010), and these in turn are important for working memory (Lewis et al., 2005; Lewis and Moghaddam, 2006). According to the prevailing view, reduced activity of cortical GABAergic interneurons leads to reduced peri-somatic inhibition of excitatory pyramidal cells, and consequently impairs the synchronized excitatory neural response

that is required for optimal working memory functions (Lewis et al., 2005; Lewis and Moghaddam, 2006; Kohl and Paulsen, 2010). It is unknown whether and/or to what extent PYY₃₋₃₆ could interfere with such neuronal synchronization processes. Given that PYY₃₋₃₆ can efficiently reduce GABAergic activity (Acuna-Goycolea et al., 2005), however, interference with GABA-mediated neuronal synchronization may offer a plausible mechanism by which PYY₃₋₃₆ disrupts working memory (Stadlbauer et al., 2013b).

7. Effects of PYY₃₋₃₆ on food intake and other behaviors: Separate functional entities or pieces of the same puzzle?

Activation of the Y2 receptor by PYY₃₋₃₆ has thus far mostly been studied with respect to the control of food intake and regulation of energy homeostasis (Chandarana and Batterham, 2008; Neary and Batterham, 2009). Although some of these studies also looked at brain areas involved in the hedonic control of eating (e.g., Batterham et al., 2007), most of them focused on PYY₃₋₃₆'s effect on homeostatic brain regions such as the hypothalamus (Broberger et al., 1997; Hahn et al., 1998). As summarized in the preceding sections, however, there is increasing evidence that PYY₃₋₃₆ modulates numerous other behavioral and cognitive functions beyond eating and activates a broad spectrum of brain regions and neurotransmitter systems. This raises the intriguing question as to whether the effects of PYY₃₋₃₆ on food intake and other behaviors represent distinct and independent behavioral processes, or whether they may be somehow interrelated.

Current knowledge does not readily allow for an evidence-based answer to this question. There are, however, a number of potential neural and neuropsychological processes that could provide a link between the PYY₃₋₃₆-induced inhibition of food intake and other functional changes in seemingly distinct behavioral domains. One

possible link relates to the role of dopamine in reward and incentive values on the one hand, and to the associations between reward and eating behavior on the other hand (Hnaskoa et al., 2004). These functional associations are highly complex and likely involve intricate interactions among homeostatic, hedonic, motivational, and associative processes (Berthoud et al., 2011; Glimcher, 2011; Kenny, 2011; Salamone and Correa, 2012; Richard et al., 2013; Morton et al., 2014). As part of these interactions, it is becoming increasingly evident that dopamine signaling cannot simply be equated with hedonic experience, i.e., the feeling of pleasure. Indeed, many studies cast doubt on the “common dopamine hypothesis of reward” concept, which in essence suggests that the experience of pleasure positively correlates with mesolimbic dopaminergic activity (for a detailed discussion, see Salamone and Correa, 2012; Richard et al., 2013). It may therefore also be questioned whether excessive food intake necessarily reflects an attempt to generate more reward in compensation for reduced mesolimbic dopamine signaling (Pothos et al., 1998; Blum et al., 2000; Volkow and Wise, 2005; Volkow et al., 2008). The mirror image of this proposition implies that increases in mesolimbic dopamine activity would lead to an inhibition of food intake because sufficient dopamine signaling suppresses the further need of more hedonic value associated with food intake. Whether or not such hedonic processes offer a possible link between the PYY₃₋₃₆-induced enhancement of striatal dopamine activity and inhibition of food intake is currently unknown. In view of the emerging limitations of the “common dopamine hypothesis of reward” (Salamone and Correa, 2012), however, we believe that this link cannot simply be explained by a dopamine-mediated modification of the hedonic value of food. Rather, we agree with the rich literature suggesting that immediate and unpredicted hedonic experiences (“liking”) are linked only minimally to mesolimbic

dopamine signaling, and instead are more directly associated with and precipitated by opioidergic signals (Berridge et al., 2009; Richard et al., 2013).

In contrast to its limited influence on “liking,” mesolimbic dopamine signaling likely plays a crucial role in “wanting,” which in relation to food intake is typically conceptualized as incentive salience (Berridge et al., 2009). Incentive salience in this case is a type of motivation that promotes approach toward and consumption of rewards, and is largely mediated by subcortical neural systems that include mesolimbic dopamine projections (Berridge et al., 2009; Richard et al., 2013). Notably, “wanting” can apply to innate (unconditioned) incentive stimuli or to conditioned stimuli that were originally neutral but now predict the availability of rewarding stimuli following prior conditioning with an innate incentive stimulus (Berridge, 2007). Depending on the context, “wanting” can thus be precipitated by various neuropsychological processes, including appetitive motivation, approach behavior, reward prediction, and exertion of effort (Berridge, 2007; Berridge et al., 2009; Salamone and Correa, 2012; Richard et al., 2013). Considering these multiple possibilities, it seems obvious that the role of dopamine in “wanting” is multifaceted. As extensively reviewed elsewhere (Berridge, 2007; Berridge et al., 2009; Salamone and Correa, 2012; Richard et al., 2013), however, it appears that striatal dopamine activity generally promotes many of the neuropsychological mechanisms underlying “wanting” and thus facilitates appetitive motivation, approach behavior, reward prediction, and exertion of effort. One prediction from these findings is that the PYY₃₋₃₆-induced elevation of striatal dopamine activity would be associated with increased “wanting” for food, and consequently, would lead to increased food intake. But this prediction is clearly at odds with the numerous findings demonstrating reduced food intake following PYY₃₋₃₆ treatment (**Table 1**), even if the peptide is administered before subjects have access to food (Batterham et al., 2002; Cox

and Randich, 2004; Koegler et al., 2005). Consequently, dopamine-mediated changes in “wanting” are unlikely to offer a plausible link between the PYY₃₋₃₆-induced enhancement of striatal dopamine activity and inhibition of food intake.

Based on the robust effects of PYY₃₋₃₆ on central information processing and salience learning discussed above, it is tempting to hypothesize that dopamine-mediated changes in salience attribution to neutral stimuli could contribute to the inhibition of food intake by PYY₃₋₃₆. Indeed, increased striatal dopaminergic activity can markedly enhance the salience of stimuli, even if they are neutral and/or have previously been associated with inconsequential experiences (Berridge and Robinson, 1998; Wise, 2004). A good example is the abolition of the LI effect by dopamine-stimulating drugs. Under conditions of low dopaminergic activity, subjects who are pre-exposed to a neutral stimulus (the CS) display slower conditioning between the CS and a consequential stimulus (the US) because they learn that the CS is a weak predictor of the US. Under such conditions, non-reinforced CS pre-exposure thus diminishes the perceived salience of the CS during conditioning (Mackintosh, 1975; Lubow et al., 1981; Weiner, 2003; Lubow, 2005). Under conditions of high dopaminergic activity, however, the inhibitory influence of CS pre-exposure on CS salience is weakened, so that subjects continue to attribute high levels of salience to the CS. As a consequence, CS-pre-exposed subjects with high dopaminergic activity behave as if they have not been pre-exposed and go on to treat the CS as a novel stimulus that attracts much of their attention. As discussed above, PYY₃₋₃₆ has a marked impact on such salience learning, with PYY₃₋₃₆-treated animals attributing high levels of salience to previously pre-exposed neutral stimuli. One may therefore predict that PYY₃₋₃₆ administration before or even during access to food could alter salience or “attractiveness” of food and shift the subject’s attentive resources away from food to other stimuli that are present at the time of food intake. Such a shift

may direct attention to internal perceptive processes or to extraneous external stimuli such as visual, auditory, or social cues that are present in the context in which food consumption occurs. While this hypothesis is novel in the context of PYY₃₋₃₆, similar concepts have been forwarded by others in other contexts. For example, it has been suggested that AMPH-induced hypophagia is not caused primarily by loss of appetite, but rather by an altered brain state in which animals cannot respond selectively (Heffner et al., 1977; Cannon et al., 2004). These postulated dopamine-mediated processes await verification. Moreover, by no means do we speculate that the PYY₃₋₃₆-induced effects on food intake are primarily or solely driven by the peptide's pro-dopaminergic effects as gastrointestinal peptides typically engage multiple processes to control food intake (Schwartz et al., 2000; Rüttimann et al., 2009; Berthoud, 2011; Woods and Ramsay, 2011; Woods and Langhans, 2012; Begg et al., 2013). Rather, our view is that the pro-dopaminergic effects of PYY₃₋₃₆ are a likely contributing factor to the inhibition of food intake and may provide an intriguing link between the peptides' effects on food intake and other behaviors.

8. Physiological versus pharmacological effects of PYY₃₋₃₆

One important question that remains to be answered by future investigations relates to the physiological relevance of the effects of PYY₃₋₃₆ on behavioral and cognitive functions. It remains currently unknown whether the aforementioned behavioral and cognitive changes induced by peripheral PYY₃₋₃₆ administration may primarily represent pharmacological effects, or alternatively, whether they also have physiological relevance. The current knowledge does not allow an evidence-based answer to this question with respect to behavior and cognition. However, numerous findings in both

humans and rodents strongly support a physiological role of PYY₃₋₃₆ in the control of food intake.

For example, genetically modified mice that lack PYY develop an obesity phenotype (Batterham et al., 2006; Boey et al., 2006), indicating that endogenous PYY signaling contributes to energy homeostasis and related metabolic processes. This hypothesis is further strengthened by the observation that obese individuals display attenuated circulating levels of PYY (Batterham et al., 2003; Chandarana et al., 2011). Moreover, various human and animal studies in which exogenous PYY₃₋₃₆ was administered in different regimens and in which post-prandial physiological levels were mimicked, efficiently reduced food intake and attenuated body weight gain (reviewed in Chandarana and Batterham, 2008; Kirchner et al., 2010).

Another important piece of evidence supporting a physiological role of PYY₃₋₃₆ in the control of food intake stems from recent functional neuroimaging studies demonstrating that physiological levels of PYY₃₋₃₆, besides activating homeostatic brain areas such as the hypothalamus, also activate numerous other cortical and subcortical brain areas, some of which play crucial roles in central reward processing (Batterham et al., 2007; De Silva et al., 2011; Weise et al., 2012). For example, Batterham et al. (2007) observed that exogenous PYY₃₋₃₆ infusion in humans, which resulted in circulating PYY₃₋₃₆ concentrations that were similar to those observed post-prandially, modulated neuronal activity within corticolimbic and higher cortical brain areas, including hypothalamus, striatum, and orbitofrontal cortex. While highlighting extra-hypothalamic effects of PYY₃₋₃₆ at physiologically relevant concentrations, the data also highlight the possibility that physiological concentrations of PYY₃₋₃₆ modulate behavioral functions beyond food intake. As mentioned above, however, the latter hypothesis awaits direct exploration by

future investigations ascertaining possible behavioral and cognitive effects of exogenous PYY₃₋₃₆ treatment at physiologically relevant concentrations.

Related to this, it remains essentially unknown whether (physiological) variations in plasma PYY₃₋₃₆ levels, be it after short-term food restriction or in the post-prandial state, could influence behavioral and cognitive functions such as incentive salience, short-term memory, and/or sensorimotor gating. Most studies that explored the behavioral effects of dietary modulations such as food restriction or binge-eating were based on experimental designs in which the dietary manipulation was chronic (Inoue et al., 2004; Carlini et al., 2008; Khabour et al., 2010; Labouesse et al., 2013). Under such conditions, the behavioral changes could readily be attributable to a broad spectrum of factors, including long-term neuronal and neurochemical adaptations. Moreover, among the few studies that investigated possible behavioral modifications following short-term food restriction (Inoue et al., 2004; McLaughlin et al., 2011; Rajab et al., 2014), none directly correlated the behavioral outcomes with plasma PYY₃₋₃₆ levels. Therefore, additional studies are clearly warranted in order to explore whether (physiological) variations in plasma PYY₃₋₃₆ levels can exert a significant influence of multiple behavioral and cognitive functions akin to the effects induced by peripheral PYY₃₋₃₆ administration. The inclusion of genetically modified animals such as mice deficient for PYY (Batterham et al., 2002) and the Y2 receptor (Baldock et al., 2002; Karl et al., 2010) may help provide answers for these open questions.

Such attempts would also help discern the Y receptor subtypes that mediate the behavioral and cognitive effects of exogenous PYY₃₋₃₆ treatment (Stadlbauer et al., 2013b, 2014). Since PYY₃₋₃₆ is a high-affinity Y2 receptor ligand (Walther et al., 2011), it is believed that the effects of peripheral PYY₃₋₃₆ administration on behavioral and cognitive functions primarily involve signaling at the Y2 receptor (Stadlbauer et al.,

2013b, 2014). This hypothesis would indeed be in agreement with findings obtained in the context of food intake: Mice deficient of the Y2 receptor are resistant to the anorectic effect of exogenous PYY₃₋₃₆ (Batterham et al., 2002), and pharmacological blockade of the Y2 receptor using a selective Y2 receptor antagonist abolishes the anorectic actions of PYY₃₋₃₆ in rats (Abbott et al., 2005). At high concentrations, however, PYY₃₋₃₆ may also bind to other Y receptor subtypes that are expressed in the CNS, including the Y1 receptor (Stanic et al., 2011). It thus remains to be explored whether the effects of exogenous PYY₃₋₃₆ treatment on incentive salience, short-term memory, and sensorimotor gating (Stadlbauer et al., 2013b, 2014) may be mediated by signaling at multiple Y receptor subclasses, or whether these may represent selective Y2 receptor-mediated effects.

9. Concluding remarks

Examining the effects of exogenous PYY₃₋₃₆ in animal models has revealed that this gut-derived peptide influences a wide spectrum of behavioral and cognitive functions. Hence, the behavioral effects of PYY₃₋₃₆ are not restricted to the control of food intake and regulation of energy homeostasis. Rather, they extend to numerous other functional domains such as central information processing, salience learning, working memory, and behavioral responding to novelty and dopamine-stimulating drugs. Whether PYY₃₋₃₆'s effects on food intake and other behaviors are somehow interrelated remains unanswered and warrants further investigation. One intriguing possibility is that PYY₃₋₃₆-induced changes in dopaminergic activity may bridge diverse behavioral manifestations to elicit inhibitory effects on food intake. The continuous integration of behavioral and cognitive neuroscience with research on food intake and metabolism may therefore be a particularly fruitful approach to address these open questions as it

may offer a heuristic appreciation of the interactions between gut-derived signals, energy homeostasis, reward, and behavioral adaptations.

Acknowledgements

We thank Marie A. Labouesse for the stimulating discussions and critical reading of the manuscript. Related work by the authors has been supported by grants from the Swiss National Science Foundation (310030_146217, U.M.) and the ETH Zurich (47 12-2, W.L.).

Tables and figures

Species	Route of administration	Food intake	References
Rat	Intraperitoneal (acute)	↓	Batterham et al., 2002; Cox et al., 2004; Nordheim et al., 2004.
	Intraperitoneal (chronic)	↓	Batterham et al., 2002; Chelikani et al., 2007;
	Intravenous (acute)	↓	Chelikani et al., 2005; Stadlbauer et al., 2013.
	Subcutaneous (chronic)	↓	Pittner et al., 2004.
Mouse	Intraperitoneal (acute)	↓	Challis et al., 2003; Halatchev et al., 2005; Martin et al., 2004; Pittner et al., 2004.
	Subcutaneous (chronic)	↓	Pittner et al., 2004.
Non-human primates	Intramuscular (acute)	↓	Moran et al., 2005.
	Intravenous (acute)	↓	Koegler et al., 2005.
Human	Intravenous (acute)	↓	Batterham et al., 2002; Degen et al., 2005; le Roux et al., 2008.

Table 1. A summary of the inhibitory effects of peripheral PYY₃₋₃₆ administration on food intake in various species. As summarized and discussed in detail elsewhere (Manning and Batterham, 2014), there are also studies reporting no significant effects of peripheral PYY₃₋₃₆ administration on food intake.

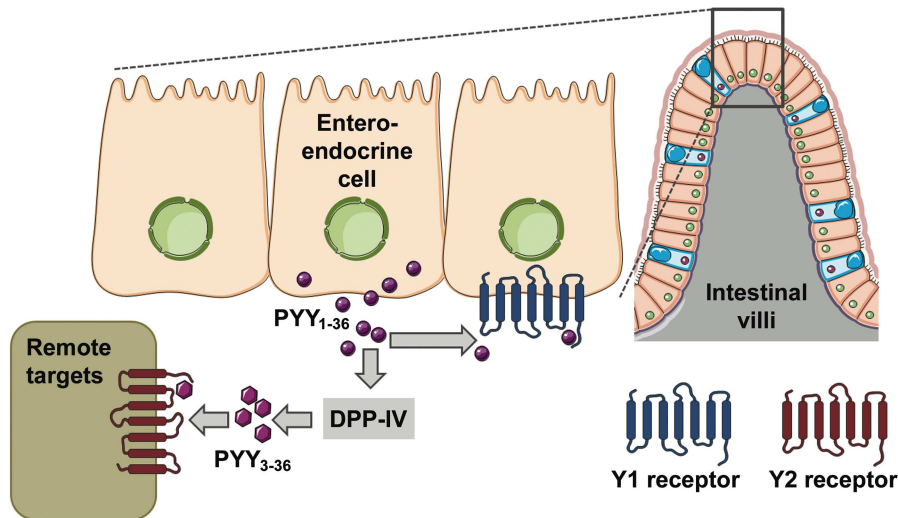


Figure 1. Simplified schematic illustration of the sites of production and action of PYY. PYY₁₋₃₆ is mainly released from distal intestinal enteroendocrine L-cells in response to luminal nutrient stimulation. PYY₁₋₃₆ can exert paracrine actions on neighboring cells by activating Y1 receptors. In the blood, PYY₁₋₃₆ is rapidly converted to PYY₃₋₃₆ by the ubiquitously expressed enzyme, dipeptidyl-peptidase IV (DPP-IV), which cleaves the two N-terminal amino acids. Circulating PYY₃₋₃₆ exerts endocrine functions and can influence remote targets such as the central nervous system by activating the Y2 receptor. Modified from Schwartz and Holst, 2010.

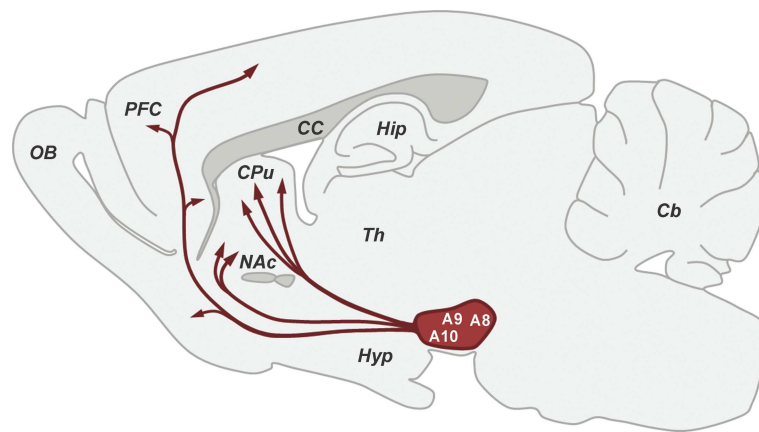


Figure 2. Schematic illustration of midbrain dopamine cell groups and their projections in rodents. A large population of dopamine cells are localized in discrete cell groups (A8-A10) of the ventral midbrain. The majority of A10 dopamine cells form the ventral tegmental area (VTA) and project to cortical areas such as the prefrontal cortex (PFC) and to limbic areas such as the nucleus accumbens (NAc), hypothalamus (Hyp), and amygdala (not shown). These projections form the mesocortical and mesolimbic dopamine pathways, respectively. A9 dopamine cells form the substantia nigra pars compacta (SNc) and project to dorsal parts of the striatum (= caudate putamen, CPu), giving rise to the nigrostriatal dopamine pathway. The A8 cell group forms a dorsal and caudal extension of the A9 cell group and contains cells that project to both striatal, limbic and cortical areas. Other dopamine cell groups such as those found in hypothalamic (A11 and A12), preoptic (A14), and olfactory (A16) areas are not shown. Cb, cerebellum; CC, corpus callosum; Hip, hippocampus; OB, olfactory bulb; Th, thalamus.

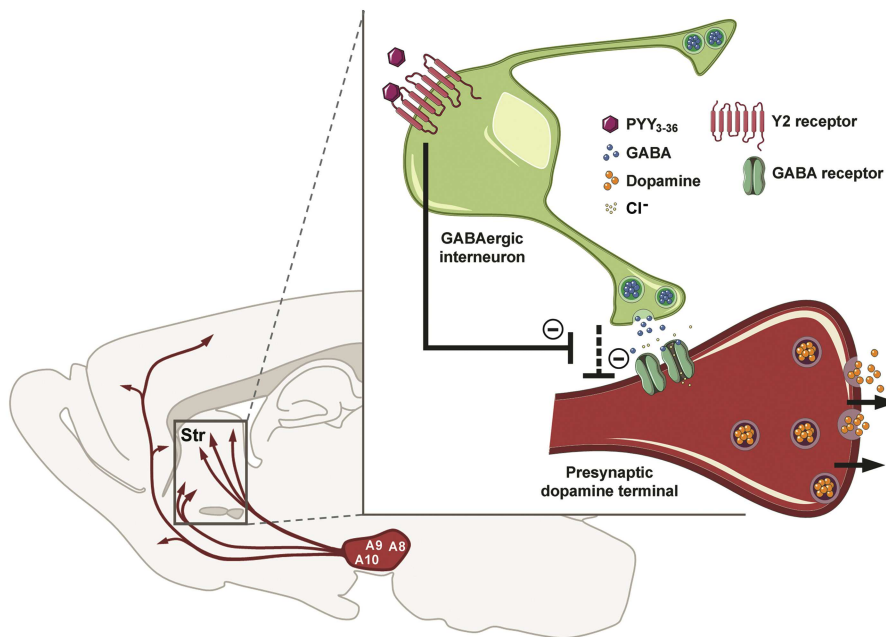


Figure 3. Proposed model by which PYY₃₋₃₆ induces hyperdopaminergic states in striatal areas. GABAergic interneurons (green) located in the striatum (Str) tonically inhibit striatal dopamine terminals (red). Activation of GABA receptors on dopamine fibers causes chloride ion (Cl⁻) influx and consequently results in hyperpolarization of presynaptic dopamine terminals. PYY₃₋₃₆-induced activation of Y2 receptors located on striatal GABAergic interneurons reduces the neural activity of GABAergic cells, which in turn weakens their inhibitory inputs onto presynaptic dopamine terminals. The PYY₃₋₃₆-induced attenuation of this fast-forward inhibitory mechanism facilitates the release of dopamine (as indicated by the black arrows).

References

- Abbott CR, Small CJ, Kennedy AR, Neary NM, Sajedi A, Ghatei MA, Bloom SR (2005). Blockade of the neuropeptide Y Y2 receptor with the specific antagonist BIIE0246 attenuates the effect of endogenous and exogenous peptide YY(3-36) on food intake. *Brain Res.* 1043(1-2):139-44.
- Acuna-Goycolea C, van den Pol AN (2005). Peptide YY(3-36) inhibits both anorexigenic proopiomelanocortin and orexigenic neuropeptide Y neurons: implications for hypothalamic regulation of energy homeostasis. *J Neurosci* 25(45): 10510-10519.
- Adewale AS, Macarthur H, Westfall TC (2005). Neuropeptide Y induced modulation of dopamine synthesis in the striatum. *Regul. Pept.* 129(1-3): 73-78.
- Adewale AS, Macarthur H, Westfall TC (2007). Neuropeptide Y-induced enhancement of the evoked release of newly synthesized dopamine in rat striatum: mediation by Y2 receptors. *Neuropharmacology* 52(6): 1396-1402.
- Adrian TE, Ferri GL, Bacarese-Hamilton AJ, Fuessl HS, Polak JM, Bloom SR (1985). Human distribution and release of a putative new gut hormone, peptide YY. *Gastroenterology* 89(5): 1070-1077.
- Alvarez Bartolomé M, Borque M, Martinez-Sarmiento J, Aparicio E, Hernández C, Cabrerizo L, Fernández-Represa JA (2002). Peptide YY secretion in morbidly obese patients before and after vertical banded gastroplasty. *Obes Surg.* 12(3):324-7.
- Arnt J, Bogeso KP, Hyttel J, Meier E (1988). Relative dopamine D1 and D2 receptor affinity and efficacy determine whether dopamine agonists induce hyperactivity or oral stereotypy in rats. *Pharmacol. Toxicol.* 62(3): 121-130.
- Baddeley A (2003). Working memory: looking back and looking forward. *Nature reviews Neuroscience.* 4(10): 829-839.
- Bardo MT, Donohew RL, Harrington NG (1996). Psychobiology of novelty seeking and drug seeking behavior. *Behav Brain Res.* 77(1-2):23-43.
- Batterham RL, Cohen MA, Ellis SM, Le Roux CW, Withers DJ, Frost GS, Ghatei MA, Bloom SR (2003). Inhibition of food intake in obese subjects by peptide YY3-36. *N Engl J Med.* 349(10):941-8.
- Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL, Wren AM, Brynes AE, Low MJ, Ghatei MA, Cone RD, Bloom SR (2002). Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature* 418(6898): 650-654.
- Batterham RL, ffytche DH, Rosenthal JM, Zelaya FO, Barker GJ, Withers DJ, Williams SC (2007). PYY modulation of cortical and hypothalamic brain areas predicts feeding behaviour in humans. *Nature* 450(7166): 106-109.

- Batterham RL, Heffron H, Kapoor S, Chivers JE, Chandarana K, Herzog H, Le Roux CW, Thomas EL, Bell JD, Withers DJ (2006). Critical role for peptide YY in protein-mediated satiation and body-weight regulation. *Cell Metab* 4(3): 223-233.
- Beckmann JS, Marusich JA, Gipson CD, Bardo MT (2011). Novelty seeking, incentive salience and acquisition of cocaine self-administration in the rat. *Behav. Brain Res.* 216(1): 159-165.
- Begg DP, Woods SC (2013). The endocrinology of food intake. *Nat Rev Endocrinol.* 9(10):584-97.
- Berglund MM, Hipskind PA, Gehlert DR (2003). Recent developments in our understanding of the physiological role of PP-fold peptide receptor subtypes. *Exp Biol Med (Maywood)* 228(3): 217-244.
- Berridge KC (2007). The debate over dopamine's role in reward: the case for incentive salience. *Psychopharmacology (Berl).* 191(3):391-431.
- Berridge KC, Robinson TE (1998). What is the role of dopamine in reward: hedonic impact, reward learning, or incentive salience? *Brain Res. Brain Res. Rev.* 28(3): 309-369.
- Berridge KC, Robinson TE, Aldridge JW (2009). Dissecting components of reward: 'liking', 'wanting', and learning. *Curr Opin Pharmacol.* 9(1):65-73.
- Berthoud HR, Lenard NR, Shin AC (2011). Food reward, hyperphagia, and obesity. *Am J Physiol Regul Integr Comp Physiol.* 300(6):R1266-77.
- Berthoud HR. Metabolic and hedonic drives in the neural control of appetite: who is the boss? *Curr Opin Neurobiol.* 21(6):888-96.
- Bitanirwe BK, Peleg-Raibstein D, Mouttet F, Feldon J, Meyer U (2010). Late prenatal immune activation in mice leads to behavioral and neurochemical abnormalities relevant to the negative symptoms of schizophrenia. *Neuropsychopharmacology* 35(12):2462-2478.
- Björklund A, Dunnett SB (2007). Dopamine neuron systems in the brain: an update. *Trends Neurosci.* 30(5):194-202.
- Blanchard MM1, Mendelsohn D, Stamp JA (2009). The HR/LR model: Further evidence as an animal model of sensation seeking. *Neurosci Biobehav Rev.* 33(7):1145-54.
- Blum K, Braverman ER, Holder JM, Lubar JF, Monastra VJ, Miller D, Lubar JO, Chen TJ, Comings DE (2000). Reward deficiency syndrome: a biogenetic model for the diagnosis and treatment of impulsive, addictive, and compulsive behaviors. *J Psychoactive Drugs.* 32 Suppl:i-iv, 1-112.
- Boey D, Lin S, Enriquez RF, Lee NJ, Slack K, Couzens M, Baldock PA, Herzog H, Sainsbury A (2008). PYY transgenic mice are protected against diet-induced and genetic obesity. *Neuropeptides* 42(1): 19-30.
- Boey D, Lin S, Karl T, Baldock P, Lee N, Enriquez R, Couzens M, Slack K, Dallmann R, Sainsbury A, Herzog H (2006). Peptide YY ablation in mice leads to the development of hyperinsulinaemia and obesity. *Diabetologia* 49:1360-70.

- Borbély E, Scheich B, Helyes Z. Neuropeptides in learning and memory. *Neuropeptides*. 47(6):439-50.
- Braff DL, Geyer MA, Swerdlow NR (2001). Human studies of prepulse inhibition of startle: normal subjects, patient groups, and pharmacological studies. *Psychopharmacology* 156(2-3): 234-258.
- Broberger C, Landry M, Wong H, Walsh JN, Hokfelt T (1997). Subtypes Y1 and Y2 of the neuropeptide Y receptor are respectively expressed in pro-opiomelanocortin- and neuropeptide-Y-containing neurons of the rat hypothalamic arcuate nucleus. *Neuroendocrinology* 66(6):393-408.
- Cabib S, Puglisi-Allegra S (1985). Different effects of apomorphine on climbing behavior and locomotor activity in three strains of mice. *Pharmacol. Biochem. Behav.* 23(4): 555-557.
- Cabrele C, Beck-Sickinger AG (2000). Molecular characterization of the ligand-receptor interaction of the neuropeptide Y family. *J Pept Sci.* 6(3):97-122.
- Cannon CM, Abdallah L, Tecott LH, Durning MJ, Palmiter RD (2004). Dysregulation of striatal dopamine signaling by amphetamine inhibits feeding by hungry mice. *Neuron*. 44(3):509-20.
- Carlini VP, Martini AC, Schiöth HB, Ruiz RD, Fiol de Cuneo M, de Barioglio SR (2008). Decreased memory for novel object recognition in chronically food-restricted mice is reversed by acute ghrelin administration. *Neuroscience* 153(4):929-34.
- Challis BG, Pinnock SB, Coll AP, Carter RN, Dickson SL, O'Rahilly S. Acute effects of PYY3-36 on food intake and hypothalamic neuropeptide expression in the mouse. *Biochem Biophys Res Commun.* 311(4):915-9.
- Chandarana K, Batterham R (2008). Peptide YY. *Curr Opin Endocrinol Diabetes Obes.* 15(1):65-72.
- Chandarana K, Gelegen C, Karra E, Choudhury AI, Drew ME, Fauveau V, Viollet B, Andreelli F, Withers DJ, Batterham RL (2011). Diet and gastrointestinal bypass-induced weight loss: the roles of ghrelin and peptide YY. *Diabetes* 60:810-18
- Charntikov S, Der-Ghazarian T, Herbert MS, Horn LR, Widarma CB, Gutierrez A, Varela FA, McDougall SA (2011). Importance of D1 and D2 receptors in the dorsal caudate-putamen for the locomotor activity and stereotyped behaviors of preweanling rats. *Neuroscience*. 183:121-33.
- Chelikani PK, Haver AC, Reidelberger RD (2005). Intravenous infusion of peptide YY(3-36) potently inhibits food intake in rats. *Endocrinology* 146(2): 879-888.
- Chelikani PK, Haver AC, Reidelberger RD (2007). Intermittent intraperitoneal infusion of peptide YY(3-36) reduces daily food intake and adiposity in obese rats. *Am J Physiol Regul Integr Comp Physiol.* 293(1):R39-46.

- Cox JE, Randich A (2004). Enhancement of feeding suppression by PYY(3-36) in rats with area postrema ablations. *Peptides*. 25(6):985-9.
- Creese I, Iversen SD (1975). The pharmacological and anatomical substrates of the amphetamine response in the rat. *Brain Res*. 83(3): 419-436.
- David HN, Anseau M, Abirini JH (2005). Dopamine-glutamate reciprocal modulation of release and motor responses in the rat caudate-putamen and nucleus accumbens of "intact" animals. *Brain Res Brain Res Rev*. 50(2):336-60.
- Degen L, Oesch S, Casanova M, Graf S, Ketterer S, Drewe J, Beglinger C (2005). Effect of peptide YY3-36 on food intake in humans. *Gastroenterology* 129(5): 1430-1436.
- De Silva A, Salem V, Long CJ, Makwana A, Newbould RD, Rabiner EA, Ghatei MA, Bloom SR, Matthews PM, Beaver JD, Dhillo WS (2011). The gut hormones PYY3-36 and GLP-1 7-36 amide reduce food intake and modulate brain activity in appetite centers in humans. *Cell Metab*. 14:700-6.
- dos Santos VV, Santos DB, Lach G, Rodrigues AL, Farina M, De Lima TC, Prediger RD (2013). Neuropeptide Y (NPY) prevents depressive-like behavior, spatial memory deficits and oxidative stress following amyloid- β (A β (1-40)) administration in mice.
- Fukushiro DF, Frussa-Filho R (2011). Chronic amphetamine transforms the emotional significance of a novel but not a familiar environment: implications for addiction. *Int. J. Neuropsychopharmacol*. 14(7): 955-965.
- Gehlert DR (1994). Subtypes of receptors for neuropeptide Y: implications for the targeting of therapeutics. *Life Sci*. 55: 551-562.
- Glimcher PW (2011). Understanding dopamine and reinforcement learning: the dopamine reward prediction error hypothesis. *Proc Natl Acad Sci USA*. 108 Suppl 3:15647-54.
- Gonçalves J, Baptista S, Olesen MV, Fontes-Ribeiro C, Malva JO, Woldbye DP, Silva AP (2012). Methamphetamine-induced changes in the mice hippocampal neuropeptide Y system: implications for memory impairment. *J Neurochem*. 123(6):1041-53.
- Graham FK (1975) The more or less startling effects of weak prestimulation. *Psychophysiology*. 12 238-248
- Grandt D, Schmiczek M, Beglinger C, Layer P, Goebell H, Eysselein VE, et al (1994). Two molecular forms of peptide YY (PYY) are abundant in human blood: characterization of a radioimmunoassay recognizing PYY 1-36 and PYY 3-36. *Regul Pept* 51(2): 151-159.
- Groenewegen HJ (2003). The basal ganglia and motor control. *Neural plasticity* 10(1-2): 107-120.
- Groenewegen HJ, Wright CI, Beijer A (1996). The nucleus accumbens: gateway for limbic structures to reach the motor system? *Prog Brain Res*. 107:485-511.

- Guo Y, Ma L, Enriori PJ, Koska J, Franks PW, Brookshire T, Cowley MA, Salbe AD, Delparigi A, Tataranni PA (2006). Physiological evidence for the involvement of peptide YY in the regulation of energy homeostasis in humans. *Obesity (Silver Spring)* 14(9):1562-70.
- Hahn TM, Breininger JF, Baskin DG, Schwartz MW (1998). Coexpression of Agrp and NPY in fasting-activated hypothalamic neurons. *Nat Neurosci* 1(4):271-272.
- Halatchev IG, Cone RD (2005). Peripheral administration of PYY(3-36) produces conditioned taste aversion in mice. *Cell Metab* 1(3): 159-168.
- Hartshorne JK (2008). Visual working memory capacity and proactive interference. *PLoS One*. 3(7):e2716.
- Heffner TG, Zigmond MJ, Stricker EM (1977). Effects of dopaminergic agonists and antagonists of feeding in intact and 6-hydroxydopamine-treated rats. *J Pharmacol Exp Ther*. 201(2):386-99.
- Heidbreder C, Feldon J (1998). Amphetamine-induced neurochemical and locomotor responses are expressed differentially across the anteroposterior axis of the core and shell subterritories of the nucleus accumbens. *Synapse* 29(4): 310-322.
- Baldock PA, Sainsbury A, Couzens M, Enriquez RF, Thomas GP, Gardiner EM, Herzog H (2002). Hypothalamic Y2 receptors regulate bone formation. *J Clin Invest*. 109(7):915-21.
- Hnasko TS, Szczycka MS, Alaynick WA, During MJ, Palmiter RD (2004). A role for dopamine in feeding responses produced by orexigenic agents. *Brain Res*. 1023(2):309-18.
- Hoffman HS, Searle JL (1965). Acoustic Variables in the Modification of Startle Reaction in the Rat. *J Comp Physiol Psychol*. 60: 53-58.
- Imamura M. (2002). Effects of surgical manipulation of the intestine on peptide YY and its physiology. *Peptides* 23:403-7.
- Inoue K, Zorrilla EP, Tabarin A, Valdez GR, Iwasaki S, Kiriike N, Koob GF (2004). Reduction of anxiety after restricted feeding in the rat: implication for eating disorders. *Biol Psychiatry*. 55(11):1075-81.
- Kapur S (2003). Psychosis as a state of aberrant salience: a framework linking biology, phenomenology, and pharmacology in schizophrenia. *Am J Psychiatry* 160(1):13-23.
- Karl T, Chesworth R, Duffy L, Herzog H (2010). Schizophrenia-relevant behaviours in a genetic mouse model for Y2 deficiency. *Behav Brain Research* 207(2): 434-440.
- Karra E, Batterham RL (2010). The role of gut hormones in the regulation of body weight and energy homeostasis. *Mol Cell Endocrinol*. 25;316(2):120-8.
- Karra E, Chandarana K, Batterham RL (2009). The role of peptide YY in appetite regulation and obesity. *J Physiol*. 15;587:19-25.
- Kenny PJ (2011). Common cellular and molecular mechanisms in obesity and drug addiction. *Nat Rev Neurosci*. 12(11):638-51.

- Khabour OF, Alzoubi KH, Alomari MA, Alzubi MA (2010). Changes in spatial memory and BDNF expression to concurrent dietary restriction and voluntary exercise. *Hippocampus*. 20(5):637-45.
- King PJ, Widdowson PS, Doods HN, Williams G (1999). Regulation of neuropeptide Y release by neuropeptide Y receptor ligands and calcium channel antagonists in hypothalamic slices. *J Neurochem* 73(2): 641-646.
- Kirchner H, Tong J, Tschöp MH, Pfluger PT (2010). Ghrelin and PYY in the regulation of energy balance and metabolism: lessons from mouse mutants. *Am. J. Physiol. Endocrinol. Metab.* 298: 909–19.
- Kodsi MH, Swerdlow NR (1995). Prepulse inhibition in the rat is regulated by ventral and caudodorsal striato-pallidal circuitry. *Behav Neurosci.* 109(5):912-28.
- Koegler FH, Enriori PJ, Billes SK, Takahashi DL, Martin MS, Clark RL, Evans AE, Grove KL, Cameron JL, Cowley MA (2005). Peptide YY(3-36) inhibits morning, but not evening, food intake and decreases body weight in rhesus macaques. *Diabetes*. 54(11):3198-204.
- Kohl MM, Paulsen O (2010). The roles of GABA_B receptors in cortical network activity. *Adv Pharmacol.* 58:205-29.
- Kretschmer BD, Koch M (1998). The ventral pallidum mediates disruption of prepulse inhibition of the acoustic startle response induced by dopamine agonists, but not by NMDA antagonists. *Brain Res.* 798(1-2):204-10.
- Labouesse MA, Stadlbauer U, Langhans W, Meyer U (2013). Chronic high fat diet consumption impairs sensorimotor gating in mice. *Psychoneuroendocrinology*. 38(11):2562-74.
- Laricchiuta D, Musella A, Rossi S, Centonze D (2014). Behavioral and electrophysiological effects of endocannabinoid and dopaminergic systems on salient stimuli. *Front Behav Neurosci.* 19;8:183.
- le Roux CW, Batterham RL, Aylwin SJ, Patterson M, Borg CM, Wynne KJ, Kent A, Vincent RP, Gardiner J, Ghatei MA, Bloom SR (2006). Attenuated peptide YY release in obese subjects is associated with reduced satiety. *Endocrinology* 147(1):3-8.
- le Roux CW, Borg CM, Murphy KG, Vincent RP, Ghatei MA, Bloom SR (2008). Supraphysiological doses of intravenous PYY3-36 cause nausea, but no additional reduction in food intake. *Ann Clin Biochem* 45(Pt 1): 93-95.
- Lewis DA, Hashimoto T, Volk DW (2005). Cortical inhibitory neurons and schizophrenia. *Nat Rev Neurosci.* 6(4):312-24.
- Lewis DA, Moghaddam B (2006). Cognitive dysfunction in schizophrenia: convergence of gamma-aminobutyric acid and glutamate alterations. *Arch Neurol.* 63(10):1372-6.
- Lubow RE (2005). Construct validity of the animal latent inhibition model of selective attention deficits in schizophrenia. *Schizophr Bull* 31(1): 139-153.

- Lubow RE, Moore AU (1959). Latent inhibition: the effect of nonreinforced pre-exposure to the conditional stimulus. *J Comp Physiol Psychol.* 52:415-9.
- Lubow RE, Weiner I, Schnur P (1981) Conditioned attention theory. In: Bower GH (ed) *The psychology of learning and motivation.* Academic, New York
- Mackintosh NJ (1975). A theory of attention: variations in the associability of stimuli with reinforcement. *Psychol Rev* 82:276–298.
- Manning S, Batterham RL (2014). The role of gut hormone peptide YY in energy and glucose homeostasis: twelve years on. *Annu Rev Physiol.* 76:585-608.
- Marinelli M1, White FJ (2000). Enhanced vulnerability to cocaine self-administration is associated with elevated impulse activity of midbrain dopamine neurons. *J Neurosci.* 20(23):8876-85.
- Martin NM, Small CJ, Sajedi A, Patterson M, Ghatei MA, Bloom SR (2004). Pre-obese and obese agouti mice are sensitive to the anorectic effects of peptide YY(3-36) but resistant to ghrelin. *Int J Obes Relat Metab Disord.* 28(7):886-93.
- Matthews M, Bondi C, Torres G, Moghaddam B (2013). Reduced presynaptic dopamine activity in adolescent dorsal striatum. *Neuropsychopharmacology.* 38(7):1344-51.
- McLaughlin IB, Dess NK, Chapman CD (2011). Modulation of methylphenidate effects on wheel running and acoustic startle by acute food deprivation in commercially and selectively bred rats. *Pharmacol Biochem Behav.* 97(3):500-8.
- Medeiros MD1, Turner AJ (1994). Processing and metabolism of peptide-YY: pivotal roles of dipeptidylpeptidase-IV, aminopeptidase-P, and endopeptidase-24.11. *Endocrinology* 134(5):2088-94.
- Mentlein R, Dahms P, Grandt D, Kruger R (1993). Proteolytic processing of neuropeptide Y and peptide YY by dipeptidyl peptidase IV. *Regul. Pept.* 49:133–44.
- Meyer U, Feldon J (2009). Neural basis of psychosis-related behaviour in the infection model of schizophrenia. *Behav Brain Res.* 204(2):322-34.
- Meyer U, Feldon J (2010). Epidemiology-driven neurodevelopmental animal models of schizophrenia. *Prog Neurobiol.*; 90(3):285-326.
- Michel MC, Beck-Sickinger A, Cox H, Doods HN, Herzog H, Larhammar D, Quirion R, Schwartz T, Westfall T (1998). XVI. International Union of Pharmacology recommendations for the nomenclature of neuropeptide Y, peptide YY, and pancreatic polypeptide receptors. *Pharmacol Rev.* 50(1):143-50.
- Morales-Medina JC, Dumont Y, Bonaventure P, Quirion R (2012). Chronic administration of the Y2 receptor antagonist, JNJ-31020028, induced anti-depressant like-behaviors in olfactory bulbectomized rat. *Neuropeptides* 46(6): 329-334.

- Moran TH, Smedh U, Kinzig KP, Scott KA, Knipp S, Ladenheim EE (2005). Peptide YY(3-36) inhibits gastric emptying and produces acute reductions in food intake in rhesus monkeys. *Am J Physiol Regul Integr Comp Physiol.* 288(2):384-8.
- Morton GJ, Meek TH, Schwartz MW (2014). Neurobiology of food intake in health and disease. *Nat Rev Neurosci.* 15(6):367-78.
- Moy SS, Nadler JJ, Young NB, Nonneman RJ, Segall SK, Andrade GM, Crawley JN, Magnuson TR (2008). Social approach and repetitive behavior in eleven inbred mouse strains. *Behav Brain Res.* 191(1):118-29.
- Neary MT, Batterham RL (2013). Peptide YY: food for thought. *Physiol Behav.* 97(5):616-9.
- Nelson AJ, Thur KE, Marsden CA, Cassaday HJ (2011). Dopamine in nucleus accumbens: salience modulation in latent inhibition and overshadowing. *J Psychopharmacol.* 25(12):1649-60.
- Nonaka N, Shioda S, Niehoff ML, Banks WA (2003). Characterization of blood-brain barrier permeability to PYY3-36 in the mouse. *J Pharmacol Exp Ther* 306(3):948-53.
- Nordheim U, Hofbauer KG (2004). Stimulation of NPY Y2 receptors by PYY3-36 reveals divergent cardiovascular effects of endogenous NPY in rats on different dietary regimens. *Am J Physiol Regul Integr Comp Physiol.* 286(1):138-42.
- Peleg-Raibstein D, Feldon J, Meyer U (2012). Behavioral animal models of antipsychotic drug actions. *Handb Exp Pharmacol.* (212):361-406.
- Peters JR, Vallie B, Difronzo M, Donaldson ST (2007). Role of dopamine D1 receptors in novelty seeking in adult female Long-Evans rats. *Brain Res. Bull.* 74(4): 232-236.
- Pijnenburg AJ, Honig WM, Van der Heyden JA, Van Rossum JM (1976). Effects of chemical stimulation of the mesolimbic dopamine system upon locomotor activity. *Eur. J. Pharmacol.* 35(1): 45-58.
- Pittner RA, Moore CX, Bhavsar SP, Gedulin BR, Smith PA, Jodka CM, Parkes DG, Paterniti JR, Srivastava VP, Young AA (2004). Effects of PYY[3-36] in rodent models of diabetes and obesity. *Int J Obes Relat Metab Disord.* 28(8):963-71.
- Pothos EN, Sulzer D, Hoebel BG (1998). Plasticity of quantal size in ventral midbrain dopamine neurons: possible implications for the neurochemistry of feeding and reward. *Appetite.* 31(3):405.
- Rajab E, Alqanbar B, Naiser MJ, Abdulla HA, Al-Momen MM, Kamal A (2014). Sex differences in learning and memory following short-term dietary restriction in the rat. *Int J Dev Neurosci.* 36:74-80.
- Rebec GV, Christensen JR, Guerra C, Bardo MT (1997). Regional and temporal differences in real-time dopamine efflux in the nucleus accumbens during free-choice novelty. *Brain Res.* 776(1-2): 61-67.

- Redrobe JP, Dumont Y, Herzog H, Quirion R (2004). Characterization of neuropeptide Y, Y(2) receptor knockout mice in two animal models of learning and memory processing. *J Mol Neurosci* 22(3): 159-166.
- Richard JM, Castro DC, Difeliceantonio AG, Robinson MJ, Berridge KC (2013). Mapping brain circuits of reward and motivation: in the footsteps of Ann Kelley. *Neurosci Biobehav Rev.* 37(9):1919-31.
- Robinson TE, Becker JB (1986). Enduring changes in brain and behavior produced by chronic amphetamine administration: a review and evaluation of animal models of amphetamine psychosis. *Brain Res.* 396(2): 157-198.
- Roeper J (2013). Dissecting the diversity of midbrain dopamine neurons. *Trends Neurosci.* 36(6):336-42.
- Rüttimann EB, Arnold M, Hillebrand JJ, Geary N, Langhans W (2009). Intrameal hepatic portal and intraperitoneal infusions of glucagon-like peptide-1 reduce spontaneous meal size in the rat via different mechanisms. *Endocrinology* 150(3):1174-81.
- Salahpour A, Ramsey AJ, Medvedev IO, Kile B, Sotnikova TD, Holmstrand E, et al (2008). Increased amphetamine-induced hyperactivity and reward in mice overexpressing the dopamine transporter. *Proc. Natl. Acad. Sci. USA* 105(11): 4405-4410.
- Salamone JD, Correa M (2012). The mysterious motivational functions of mesolimbic dopamine. *Neuron.* 2012 76(3):470-85.
- Salamone JD, Correa M (2013). Dopamine and food addiction: lexicon badly needed. *Biol Psychiatry.* 2013 73(9):e15-24.
- Sanderson DJ, Bannerman DM (2012). The role of habituation in hippocampus-dependent spatial working memory tasks: evidence from GluA1 AMPA receptor subunit knockout mice. *Hippocampus.* 22(5):981-94.
- Sanderson DJ, Good MA, Skelton K, Sprengel R, Seeburg PH, Rawlins JN, Bannerman DM (2009). Enhanced long-term and impaired short-term spatial memory in GluA1 AMPA receptor subunit knockout mice: evidence for a dual-process memory model. *Learn Mem.* 16(6):379-86.
- Schwartz MW, Woods SC, Porte D Jr, Seeley RJ, Baskin DG (2000). Central nervous system control of food intake. *Nature.* 404(6778):661-71.
- Schwartz TW, Holst B (2010). An enteroendocrine full package solution. *Cell Metab.* 11(6):445-7.
- Sloth B1, Davidsen L, Holst JJ, Flint A, Astrup A (2007). Effect of subcutaneous injections of PYY1-36 and PYY3-36 on appetite, ad libitum energy intake, and plasma free fatty acid concentration in obese males. *Am J Physiol Endocrinol Metab.* 293(2):E604-9.
- Smith A, Li M, Becker S, Kapur S (2006). Dopamine, prediction error and associative learning: a model-based account. *Network.* 17(1):61-84.

- Smith Y, Kieval JZ (2000). Anatomy of the dopamine system in the basal ganglia. *Trends Neurosci.* 23(10):S28-33.
- Smith-White MA, Hardy TA, Brock JA, Potter EK (2001). Effects of a selective neuropeptide Y Y2 receptor antagonist, BIIE0246, on Y2 receptors at peripheral neuroeffector junctions. *Br J Pharmacol* 132(4): 861-868.
- Soderpalm B, Ericson M (2013). Neurocircuitry involved in the development of alcohol addiction: the dopamine system and its access points. *Curr Top Behav Neurosci* 13: 127-161.
- Sodowski K, Zwirska-Korczala K, Kuka D, Kukła M, Budziszewska P, Czuba B, Włoch A, Cnota W, Bielański W, Brzozowski T, Rehfeld JF, Zdun R, Konturek PC (2007). Basal and postprandial gut peptides affecting food intake in lean and obese pregnant women. *J Physiol Pharmacol.* 58 Suppl 1:37-52.
- Stadlbauer U, Arnold M, Weber E, Langhans W (2013a). Possible mechanisms of circulating PYY-induced satiation in male rats. *Endocrinology* 154(1): 193-204.
- Stadlbauer U, Langhans W, Meyer U (2013b). Administration of the Y2 receptor agonist PYY3-36 in mice induces multiple behavioral changes relevant to schizophrenia. *Neuropsychopharmacology* 38(12):2446-55.
- Stadlbauer U, Weber E, Langhans W, Meyer U (2014). The Y2 receptor agonist PYY(3-36) increases the behavioural response to novelty and acute dopaminergic drug challenge in mice. *Int J Neuropsychopharmacol.* 17(3):407-19.
- Stanic D, Brumovsky P, Fetissov S, Shuster S, Herzog H, Hokfelt T (2006). Characterization of neuropeptide Y2 receptor protein expression in the mouse brain. I. Distribution in cell bodies and nerve terminals. *J Comp Neurol* 499(3): 357-390.
- Stanic D, Mulder J, Watanabe M, Hokfelt T (2011). Characterization of NPY Y2 receptor protein expression in the mouse brain. II. Coexistence with NPY, the Y1 receptor, and other neurotransmitter-related molecules. *J Comp Neurol* 519(7): 1219-1257.
- Stanley S, Wynne K, Bloom S (2004). Gastrointestinal satiety signals III. Glucagon-like peptide 1, oxyntomodulin, peptide YY, and pancreatic polypeptide. *Am J Physiol Gastrointest Liver Physiol.* 286(5):G693-7.
- Swordlow NR, Braff DL, Geyer MA (2000). Animal models of deficient sensorimotor gating: what we know, what we think we know, and what we hope to know soon. *Behav Pharmacol.* 11(3-4): 185-204.
- Swordlow NR, Geyer MA, Braff DL (2001). Neural circuit regulation of prepulse inhibition of startle in the rat: current knowledge and future challenges. *Psychopharmacology;* 156(2-3):194-215.

- Swerdlow NR, Koob GF (1987). Lesions of the dorsomedial nucleus of the thalamus, medial prefrontal cortex and pedunclopontine nucleus: effects on locomotor activity mediated by nucleus accumbens-ventral pallidal circuitry. *Brain Res.* 412(2):233-43.
- Tindell AJ, Berridge KC, Zhang J, Peciña S, Aldridge JW (2005). Ventral pallidal neurons code incentive motivation: amplification by mesolimbic sensitization and amphetamine. *Eur J Neurosci.* 22(10):2617-34.
- Tzschentke TM (2001). Pharmacology and behavioral pharmacology of the mesocortical dopamine system. *Prog Neurobiol* 63(3): 241-320.
- Van den Heuvel DM, Pasterkamp RJ (2008). Getting connected in the dopamine system. *Prog Neurobiol* 85(1): 75-93.
- Vasse M, Protais P (1989). Potentiation of apomorphine-induced stereotyped behaviour by acute treatment with dopamine depleting agents: a potential role for an increased stimulation of D1 dopamine receptors. *Neuropharmacology.* 28(9): 931-939.
- Volkow ND, Wang GJ, Fowler JS, Telang F (2008). Overlapping neuronal circuits in addiction and obesity: evidence of systems pathology. *Philos Trans R Soc Lond B Biol Sci.* 363(1507):3191-200.
- Volkow ND, Wise RA (2005). How can drug addiction help us understand obesity? *Nat Neurosci.* 8(5):555-60.
- Vuillermot S, Joodmardi E, Perlmann T, Ove Ogren S, Feldon J, Meyer U (2011). Schizophrenia-relevant behaviors in a genetic mouse model of constitutive Nurr1 deficiency. *Genes Brain Behav.* 10(5): 589-603.
- Walther C, Mörl K, Beck-Sickinger AG (2011). Neuropeptide Y receptors: ligand binding and trafficking suggest novel approaches in drug development. *J Pept Sci.* 17(4):233-46.
- Weiner I (2003). The “two-headed” latent inhibition model of schizophrenia: modeling positive and negative symptoms and their treatment. *Psychopharmacology.* 169:257–297.
- Weise CM, Thiyyagura P, Reiman EM, Chen K, Krakoff J (2012). Postprandial plasma PYY concentrations are associated with increased regional gray matter volume and rCBF declines in caudate nuclei—a combined MRI and H2150 PET study. *Neuroimage* 60:592–600.
- Widdowson PS (1993). Quantitative receptor autoradiography demonstrates a differential distribution of neuropeptide-Y Y1 and Y2 receptor subtypes in human and rat brain. *Brain Res.* 631: 27–38.
- Winton-Brown TT, Fusar-Poli P, Ungless MA, Howes OD (2014). Dopaminergic basis of salience dysregulation in psychosis. *Trends Neurosci.* 37(2):85-94.
- Wise RA (2004). Dopamine, learning and motivation. *Nat. Rev. Neurosci.* 5(6): 483-494.

- Woods SC, Langhans W (2012). Inconsistencies in the assessment of food intake. *Am J Physiol Endocrinol Metab.* 303(12):E1408-18.
- Woods SC, Ramsay DS (2011). Food intake, metabolism and homeostasis. *Physiol Behav.* 104(1):4-7
- Yang H. 2002. Central and peripheral regulation of gastric acid secretion by peptide YY. *Peptides* 23:349-58.
- Young AM, Moran PM, Joseph MH (2005). The role of dopamine in conditioning and latent inhibition: what, when, where and how? *Neurosci Biobehav Rev.* 29(6):963-76.
- Zambello E, Zanetti L, Hédou GF, Angelici O, Arban R, Tasan RO, Sperk G, Caberlotto L (2011). Neuropeptide Y-Y2 receptor knockout mice: influence of genetic background on anxiety-related behaviors. *Neuroscience.* 10;176:420-30.