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## **Premise and promise of mesenchymal stem cell-based therapies in clinical vascularized composite allotransplantation**

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**Abstract:** **PURPOSE OF REVIEW** Over the past decade, clinical vascularized composite allotransplantation (VCA) has enabled functional and quality of life restoration in a wide range of indications secondary to devastating tissue loss. However, the spectre of toxicity and long-term complications of chronic immunosuppression has curtailed the momentum of VCA. This study summarizes the literature evidence behind successful mesenchymal stem cell (MSC)-based cell therapies highlighting their multi-pronged immunomodulatory, restorative and regenerative characteristics with special emphasis towards VCA applications. **RECENT FINDINGS** Experimental and clinical studies in solid organs and VCA have confirmed that MSCs facilitate immunosuppression-free allograft survival or tolerance, stimulate peripheral nerve regeneration, attenuate ischaemia-reperfusion injury, and improve tissue healing after surgery. It has been hypothesized that MSC-induced long-term operational tolerance in experimental VCA is mediated by induction of mixed donor-specific chimerism and regulatory T-cell mechanisms. All these characteristics of MSCs could thus help expand the scope and clinical feasibility of VCA. **SUMMARY** Cellular therapies, especially those focusing on MSCs, are emerging in solid organ transplantation including VCA. Although some clinical trials have begun to assess the effects of MSCs in solid organ transplantation, much scientific domain remains uncharted, especially for VCA.

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# Premise and promise of mesenchymal stem cell-based therapies in clinical vascularized composite allotransplantation

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## Purpose of review

Over the past decade, clinical vascularized composite allotransplantation (VCA) has enabled functional and quality of life restoration in a wide range of indications secondary to devastating tissue loss. However, the spectre of toxicity and long-term complications of chronic immunosuppression has curtailed the momentum of VCA. This study summarizes the literature evidence behind successful mesenchymal stem cell (MSC)-based cell therapies highlighting their multipronged immunomodulatory, restorative and regenerative characteristics with special emphasis towards VCA applications.

## Recent findings

Experimental and clinical studies in solid organs and VCA have confirmed that MSCs facilitate immunosuppression-free allograft survival or tolerance, stimulate peripheral nerve regeneration, attenuate ischaemia-reperfusion injury, and improve tissue healing after surgery. It has been hypothesized that MSC-induced long-term operational tolerance in experimental VCA is mediated by induction of mixed donor-specific chimerism and regulatory T-cell mechanisms. All these characteristics of MSCs could thus help expand the scope and clinical feasibility of VCA.

## Summary

Cellular therapies, especially those focusing on MSCs, are emerging in solid organ transplantation including VCA. Although some clinical trials have begun to assess the effects of MSCs in solid organ transplantation, much scientific domain remains uncharted, especially for VCA.

## Keywords

hand and face transplantation, immunomodulation, mixed chimerism, regulatory T cells, tolerance

## INTRODUCTION

Until recently, few reconstructive options were available for patients with disfiguring and functional devastating tissue defects secondary to trauma, oncological resection or congenital malformation. Over the past 2 decades, the technical, immunologic and functional feasibility of vascularized composite allotransplantation (VCA) as an alternative restorative option has been established in such indications. Overall, intermediate and long-term graft and patient outcomes have been encouraging for extremity and facial VCA with improved quality of life [1,2]. The prospect of allograft dependency on chronic, lifelong drug immunosuppression, with the risk of infectious, metabolic or neoplastic complications remains a significant hurdle for clinical advancement of VCA [3]. Development of safe and effective protocols consistent with immunosuppression-free graft survival is an immediate priority in nonlife saving transplants such as VCA.

Unlike other solid organs, optimal functional recovery is a prerequisite in VCA, which relies on timely and proper sensory and motor nerve regeneration and reintegration for overall success. Without it, a VCA would be deemed a failure, increasing the risk/benefit ratio of long-term immunosuppression and questioning the ethical equipoise of VCA. Also unlike solid organs, VCA are extraneous grafts requiring matching for size, skin colour, tone and anatomical congruity. These requirements limit the

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## KEY POINTS

- Clinical vascularized composite allotransplantation is an expanding therapeutic option for functional restoration after devastating tissue loss, however, hampered by the need for long-term immunosuppression.
- Cellular therapies including mesenchymal stem cells show promise in establishing long-term operational allograft tolerance and obviate to the hurdles of drug-immunosuppression.
- Mesenchymal stem cells show additional beneficial effects (peripheral nerve regeneration, attenuation of I/R injury, facilitation of tissue healing after surgery) that additionally expand clinical feasibility of VCA.

available donor-pool and may require donor grafts to be shared over long distances, with increased cold ischaemia times and possible risks of worse ischaemia-reperfusion (I/R) injury. All these constraints jeopardize the life-enhancing benefits of VCA procedures. Research efforts striving to overcome these obstacles have included the use of cellular therapies such as those incorporating mesenchymal stem cells (MSCs), given their multifaceted effects ranging from immunomodulation and tissue healing to neuroregeneration and mitigation of I/R injury [4,5<sup>6</sup>,6].

Herein we provide a brief overview of the premise and potential of MSC therapy in addressing different therapeutic goals inherent to VCA, focusing on their broad-based beneficial effects and recently emerging outcomes of experimental studies and clinical trials in solid organ transplantation (SOT) and VCA.

## MESENCHYMAL STEM CELLS

A broad selection of different cell types could play a relevant role in cell-based therapy for VCA. Regulatory T cells (T<sub>regs</sub>) [7], dendritic cells [8,9], MSCs, whole bone marrow (WBM) and other cells have been evaluated in SOT and VCA. MSCs are undifferentiated, multipotent, self-renewing cells, widespread throughout the body, possessing angiogenic, immunomodulatory, pro-neurogenic and antiapoptotic functionality and capable of mesenchymal tissue differentiation, for example bone, muscle, cartilage, endothelium and fat [10,11]. Together, these characteristics have relevance and impact in VCA. MSCs have been isolated from various tissues including, bone marrow mesenchymal stromal cells (BM-MSCs) and subcutaneous fat tissue (adipose-derived stem cells; ASCs). The latter source is appealing due to ease of procurement, low

morbidity and high cell yields. ASCs may also have superior immunomodulatory potency [5<sup>6</sup>], as compared with bone marrow or other sources of MSCs [5<sup>6</sup>,6,12,13].

## POTENTIAL OF MESENCHYMAL STEM CELL THERAPIES IN CLINICAL VASCULARIZED COMPOSITE ALLOTRANSPLANTATION

### Donor-cell chimerism

A multitude of experimental reports in small and large animals have demonstrated prolonged allograft survival in VCA either after single or repetitive MSC administration [14<sup>15</sup>,15,16,17<sup>18</sup>,19–24]. MSCs seem to favour the establishment of mixed donor-cell chimerism in VCA recipients [25–27]. Cetrulo *et al.* found that hematopoietic cell infusions induced donor-specific tolerance to myocutaneous flap VCA in a large animal model with persistent donor-cell chimerism over weeks in a manner similar to VCA transplantation in established chimeras [28]. This is of important translational value for reconstructive transplantation, as VCA donors are usually brain dead and all tissues have to be procured simultaneously. Although some authors successfully achieved long-term tolerance after single infusion of WBM even without persistent peripheral donor-cell chimerism [25], other reports suggest a positive effect of preestablished and stable donor-specific chimerism on composite allograft survival across major histocompatibility mismatches [16]. Thus, although donor-cell chimerism is regarded as key step for long-term tolerance, it is not yet clear if sustained chimerism is actually required for graft survival. Interestingly, in a recent report allografts survived after loss of donor-specific chimerism and were not rejected after reconstitution with host hematopoietic cells [29].

### Modulation of immune response

The immunomodulatory and anti-inflammatory properties of MSCs hold promise as a therapeutic approach for induction of allograft tolerance, even though it is not clear yet if MSCs alone are able to induce a solid long-term transplant tolerance or if cellular therapies have to be supported at least in part by other cell types and/or drug-based immunosuppressive regimens. As far as it concerns VCA, there is only a small body of evidence for the modulatory effects of MSCs. Recent reports point out to a beneficial immunomodulatory effect of both BM-MSCs and ASCs on the survival rate of rat hind limbs

while coupled to a short initial course of calcineurin inhibitors [17<sup>•</sup>,18<sup>•</sup>].

There are several putative mechanisms by which MSCs can exert their effect to dampen the immune response to alloantigens, all summarized recently by Kim *et al.* (reviewed in [30]).

One mechanism under intensive investigation is the MSC-mediated T<sub>reg</sub> recruitment and expansion in both peripheral blood and allograft [17<sup>•</sup>,22,31]. In contrast, however, Jiang *et al.* found that T<sub>reg</sub> depletion in transplant recipients did not compromise BM-MSCs ability to suppress allograft rejection in murine heart transplant recipients [32].

Most of MSC's immunosuppressive activity is mediated through paracrine signalling, with the MSC secretome playing a pivotal role and acting on a variety of pathways including interaction with T and B lymphocytes, as well as inhibition of macrophages, monocytes and natural killer cells among others.

Soluble factors involved are for example indoleamine 2,3-dioxygenase, transforming growth factor- $\beta$ , nitric oxide, tumour necrosis factor-inducible gene 6 protein and prostaglandins. Moreover, cells of the immune system can shift from a pro-inflammatory to an anti-inflammatory phenotype under MSC influence.

Other than dampening the recipient's immune reaction against allografts, through their suppressive function MSCs offer the opportunity to attenuate graft-versus-host disease (GvHD) as well (reviewed in [33]), especially when given in conjunction with hematopoietic stem cell (HSC) or WBM, where the risk of GvHD is increased.

### Syngeneic, allogeneic and xenogeneic mesenchymal stromal cells

It is important to distinguish between allogeneic, syngeneic or even xenogeneic MSC sources. Donor-derived (allogeneic) cells need to be procured at the time of transplantation along with the graft and either frozen down, further processed or placed in culture and expanded for later administration. Autologous (syngeneic) cells are readily available from the host, can be freshly procured any time and repetitively administered. Chen *et al.* reported that a single injection of syngeneic ASCs along with antilymphocyte serum and a short course of cyclosporine A achieved rodent limb VCA survival in 66% recipients [18<sup>•</sup>]. In comparison, studies using allogeneic MSCs reported a lower long-term survivor rate [17<sup>•</sup>,20], suggesting that even if MSCs are immunoprivileged (due to absence of MHC Class II markers), autologous cells could have a higher survival rate and better impact on the outcome. One

group recently used (xenogeneic) human ASCs (hASC) to induce skin allograft tolerance in mice [24]. hASC monotherapy accomplished only slight improvements in graft survival. In contrast, combination of hASCs with murine WBM achieved 100% long-term survival, suggesting rejection of the xenogeneic ASCs. We recently reported that BM-MSCs and ASCs are not substantially different in efficacy in a rat osteomyocutaneous VCA model with a short course of tacrolimus under an antilymphocyte serum preconditioning regimen [17<sup>•</sup>]. Both regimens successfully promoted immunosuppression-free long-term survival in around 50% of the recipients with transient peripheral chimerism and increased T<sub>reg</sub> levels. The in-vitro immunomodulatory function of ASCs was superior to that of BM-MSCs.

### Drug-mesenchymal stem cell interactions

Recent literature evidence reaffirms that an induction regimen or early short-course immunosuppression is probably essential for the success of tolerance strategies [34,35]. Eggenhofer *et al.* reported accelerate rejection and worse outcomes after MSC therapy without concomitant immunosuppression [36]. Allogeneic mouse islets were prolonged after MSCs in combination with CTLA-4-Ig co-stimulatory blockade but not with isolated MSC therapy [37<sup>•</sup>]. Lee *et al.* reported that in the absence of preconditioning, hASCs achieved only a discrete prolongation of murine allogeneic skin grafts survival injection of conditioned medium prolonged graft survival and reduced inflammatory tissue cytokine levels, suggesting a paracrine mechanism [34]. In another study, repetitive administration of omental rat ASCs over 3 days without adjunct immunosuppression delayed skin graft rejection without long-term tolerance, despite increased T<sub>reg</sub> levels in skin specimens [35]. Similarly, Larocca *et al.* found that allogeneic ASCs can prolong skin graft survival and recruit T<sub>regs</sub> into draining lymph nodes. Of interest, in that study, allogeneic donor-matched ASCs were more efficient than host-matched syngeneic or third-party ASCs [38]. Nevertheless, the toxic effect of concomitant conditioning and maintenance drugs on MSC viability and function should not be underestimated as supported by our recent in-vitro findings [39]. Singh *et al.* reported that sirolimus (rapamycin) is superior to tacrolimus in preserving the T<sub>reg</sub> phenotype and FoxP3 expression [40]. Given the in-vivo impact and relevance of such toxicity, the timing, dosing and frequency of cellular therapies may need to be optimized and tailored in the context of such induction regimens in order to avoid collateral effects.

## Supportive use of different cell types

The presence of active vascularized bone marrow in certain VCA may benefit allograft survival [41]. It remains unclear if systemic administration of a similar amount of WBM would achieve similar effects or if the donor-specific bone marrow niche is a requirement that fosters the development of robust and stable donor-cell chimerism.  $T_{reg}$  induction and recruitment may be important for maintenance of peripheral tolerance and avoidance of rejection. A recent report by Obermajer *et al.* suggests direct T-cell conversion from Th1-T cells to  $T_{regs}$  by MSC influence as a mechanism of  $T_{reg}$  induction [42]. A 2013 study revealed that  $T_{reg}$  injection (at lower doses than with WBM therapy) along with vascularized bone marrow transplantation achieved 90% graft acceptance and sustained peripheral donor-cell chimerism without need for cyto-reduction [7]. In a canine VCA model, Mathes *et al.* found that intragraft  $T_{reg}$  levels were similar in long-term VCA recipients regardless of bone marrow infusion. This was confirmed *in vitro* by increased suppressor function on alloantigen-stimulated T-cell proliferation by  $T_{regs}$  derived from long-term acceptors [14<sup>\*\*\*</sup>]. An interesting potential approach could be the use of purified MSCs along with HSCs, which have been shown to enhance engraftment and reconstitution [43]. Early clinical protocols combining ASCs and HSCs for tolerance induction in kidney transplantation show promising results [5<sup>\*\*</sup>]. However, further experimental studies are needed to elucidate if this is a valid option for VCA.

## Therapy of acute and chronic rejection

Studies investigating the potential ability of autologous, allogeneic, fresh or frozen MSCs for attenuation and therapy of rejection in allotransplantation are scant. The characteristics of MSC in promoting  $T_{reg}$  recruitment, anti-inflammatory function and paracrine secretion of anti-inflammatory factors may all benefit their role in prevention or management of acute or chronic rejection. However, without substantive research, these purported advantages remain speculative in VCA.

## Routes of administration

The mode of MSC administration in VCA is of relevance as most cells get entrapped in lungs and filtering organs such as liver and spleen after intravenous injection [44]. Albeit the simplicity and ease of intravenous injection, barring potential embolic events, it is still under debate if MSCs need to home and engraft locally into the target tissue to exert their therapeutic effect or if they can exert their

immunomodulatory effects from distance in a paracrine fashion. According to a recent report early, high-dose, intravenous MSC administration should be preferred [32]. Alternative approaches represent the direct intraarterial injection to improve target cell-load [45], local scaffold-assisted delivery to favour cell viability and proliferation at the target site and diminish spreading of cells to have high loco-regional effects [46] or cell encapsulation to allow for systemic administration but with improved cell survival through a protective 'cell coating' [47,48]. Despite all these approaches, the ideal therapeutic dosage of MSCs for a given application remains unknown.

## Nerve regeneration, protection from ischaemia-reperfusion injury, wound and bone healing

The versatility of MSC therapy potentially extends beyond tolerance induction and modulation of immune responses after VCA. There are many other supplementary benefits of MSCs of relevance to VCA outcomes. Most notable is peripheral nerve regeneration, which is critical in VCA, where unlike in SOT, overall functional restoration relies on motor and sensory recovery in the graft. Multiple studies confirm that bone marrow and ASCs facilitate functional recovery after peripheral nerve injury [49–51]. Intravenous MSCs home to injured sciatic nerves and improve functional recovery after transection [51]. ASCs have been shown to exert their beneficial effects through paracrine neurotrophic and angiogenic effects, a mechanism that is still debated [49]. MSC therapy aiming at enhancing nerve regeneration could necessitate repetitive treatments due to the slow nature of the regeneration process. I/R injury is linked with acute and chronic rejection and worse long-term outcomes in transplantation [52]. The ability of MSCs in mitigating reperfusion injury has been attributed to paracrine anti-inflammatory effects [53], with reduction in I/R injury in skin flaps [54,55] and in renal I/R injury [56,57]. In the latter study, MSCs but not conditioned medium were able to ameliorate I/R injured kidneys, which is in line with own findings where BM-MSCs but not conditioned medium were anti-inflammatory on activated endothelium in a critically ischaemic skin flap [53]. Effective mitigation of I/R injury after cold ischaemia could promote VCA allocation over larger geographical distances, decreasing ischaemia-related time constraints and expanding VCA donor-pool. MSCs also enhance bone healing, which is of utmost relevance for (bony) face and extremity transplantation [58]. BM-MSCs have been shown to improve healing of

critical calvarial defects after systemic infusion despite a low local cell engraftment rate [59]. Finally, the antiscarring, antiapoptotic and angiogenic properties of MSCs could all potentially contribute to improved wound healing and overall outcomes.

### Mesenchymal stem cell therapies for clinical solid organ transplantation and bone-marrow transplantation

First clinical trials have begun evaluating the use of MSCs for induction of allograft tolerance in SOT, especially in kidney transplantation [59,60–63]. MSC administration prior and after kidney transplantation seems to be safe and efficacious in reducing the dose of maintenance drugs [60,62]. In a very recent case series, pre- and one post-transplant administration of autologous MSCs expanded peripheral T<sub>regs</sub> and decreased T-cell proliferation retaining graft function in kidney transplant patients [63]. An interesting approach by Vanikar *et al.* included combined ASC and HSC therapy prior to transplantation under a nonmyeloablative conditioning regimen for immunomodulation in kidney transplantation showing a 94.7% 5-year survival compared with 84% in the control group [59]. Other than tolerance induction, cellular therapy of GvHD is another aspect under clinical investigation [64,65]. In cases of steroid-refractory GvHD in HSC-transplanted patients, repetitive third-party BM-MS-C infusions over 3 weeks achieved more than 70% responders with cessation of GvHD symptoms [66], which is in line with a very recent report [64]. Fortunately, human freeze-thawed BM-MS-Cs have recently been found to retain their immunosuppressive activity against GvHD, potentially allowing for third party off-the-shelf cell products for therapy of such conditions [67].

The Pittsburgh Protocol is the first cell therapy protocol used in upper extremity transplantation incorporating WBM that has facilitated monotherapy maintenance of VCA in compliant patients [13]. Other groups have administered WBM in clinical VCA as part of a multidrug immunosuppression protocol [6]. However, combination cellular therapies of WBM, HSCs and MSCs remain unexplored. A combined approach could bring promising benefits in VCA as well, underscored by recent evidence suggesting improved engraftment of HSC co-administered with MSCs [68]. Experimental data for other cell products such as ASCs in VCA are also scarce.

### CONCLUSION

Proof of principle has been established for cellular therapies using MSCs in both SOT and VCA. Clinical

trials have investigated MSC therapy for renal transplantation. Although there are clinical reports of WBM infusion after hand and face transplants [6,69], there are no reported clinical attempts incorporating isolated or enriched MSCs in VCA. Multiple studies in the literature reinforce the promise and potential for MSCs in prolonging allograft survival and other aspects (promoting nerve regeneration, protecting from I/R injury) that could improve overall graft outcomes after VCA. Many questions do persist, such as the mechanisms underlying tolerance or graft acceptance, optimizing the conditioning regimen in the context of induction immunosuppression, the dosing, timing, route and frequency of cell administration and the use of other cell types such as ASCs in conjunction with MSCs to improve synergistic, complementary or additive efficacy after VCA.

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### Conflicts of interest

There are no conflicts of interest.

### REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Petruzzo P, Kanitakis J, Badet L, *et al.* Long-term followup in composite tissue allotransplantation: in-depth study of five (hand and face) recipients. *Am J Transplant* 2011; 11:808–816.
  2. Petruzzo P, Testelin S, Kanitakis J, *et al.* First human face transplantation: 5 years outcomes. *Transplantation* 2012; 93:236–240.
  3. Gorantla VS, Barker JH, Jones JW Jr, *et al.* Immunosuppressive agents in transplantation: mechanisms of action and current antirejection strategies. *Microsurgery* 2000; 20:420–429.
  4. Vanikar AV, Trivedi HL, Kumar A, *et al.* Co-infusion of donor adipose tissue-derived mesenchymal and hematopoietic stem cells helps safe minimization of immunosuppression in renal transplantation – single center experience. *Ren Fail* 2014; 36:1376–1384.
  5. Vanikar AV, Trivedi HL. Stem cell transplantation in living donor renal transplantation for minimization of immunosuppression. *Transplantation* 2012; 94:845–850.
- This study is a first in its genre, achieving long-term tolerance in clinical renal transplantation with dramatically minimized drug-immunosuppression. This is the first clinical study using adipose-derived stem cells for tolerance establishment and suggesting a beneficial synergistic effect of combined administration of adipose-derived and hematopoietic stem cells.
6. Schneeberger S, Gorantla VS, Brandacher G, *et al.* Upperextremity transplantation using a cell-based protocol to minimize immunosuppression. *Ann Surg* 2013; 257:345–351.
  7. Lin JY, Tsai FC, Wallace CG, *et al.* Combined treatment with regulatory T cells and vascularized bone marrow transplantation creates mixed chimerism and induces donor-specific tolerance to vascularized composite allografts without cytoreductive conditioning. *Chimerism* 2013; 4:20–22.
  8. Ikeguchi R, Sacks JM, Unadkat JV, *et al.* Long-term survival of limb allografts induced by pharmacologically conditioned, donor alloantigen-pulsed dendritic cells without maintenance immunosuppression. *Transplantation* 2008; 85:237–246.

9. Wei Y, Zheng D, Li X, *et al.* Infusion of dendritic cells carrying donor lymphocytes treated with 8-methoxypsoralen and ultraviolet A light induces CD19+ IL-10+ regulatory B cells and promotes skin allograft survival. *Transplant Proc* 2014; 46:3641–3646.
  10. Parekkadan B, Milwid JM. Mesenchymal stem cells as therapeutics. *Annu Rev Biomed Eng* 2010; 12:87–117.
  11. Keating A. Mesenchymal stromal cells: new directions. *Cell Stem Cell* 2012; 10:709–716.
  12. Reinders ME, Bank JR, Dreyer GJ, *et al.* Autologous bone marrow derived mesenchymal stromal cell therapy in combination with everolimus to preserve renal structure and function in renal transplant recipients. *J Transl Med* 2014; 12:331.
  13. Gorantla VS, Brandacher G, Schneeberger S, *et al.* Favoring the risk-benefit balance for upper extremity transplantation—the Pittsburgh Protocol. *Hand Clin* 2011; 27:511–520; ix–x.
  14. Mathes DW, Chang J, Hwang B, *et al.* Simultaneous transplantation of hematopoietic stem cells and a vascularized composite allograft leads to tolerance. *Transplantation* 2014; 98:131–138.
- Important paper suggesting that preestablished donor-specific chimerism is not required for long-term operational tolerance in VCA. This is of strong translational value due to concomitant cell- and graft harvesting from brain-dead heart-beating donors in the clinical practice.
15. Mathes DW, Hwang B, Graves SS, *et al.* Tolerance to vascularized composite allografts in canine mixed hematopoietic chimeras. *Transplantation* 2011; 92:1301–1308.
  16. Mathes DW, Solari MG, Gazelle GS, *et al.* Stable mixed hematopoietic chimerism permits tolerance of vascularized composite allografts across a full major histocompatibility mismatch in swine. *Transpl Int* 2014; 27:1086–1096.
  17. Plock J, Schnider J, Zhang W, *et al.* Adipose and bone marrow derived mesenchymal stem cells prolong graft survival in vascularized composite allotransplantation. *Transplantation* 2015; 99:1765–1773.
- In this first head-to-head comparison the authors show increased immunomodulatory potency of adipose-derived stem cells compared to bone marrow-derived mesenchymal cells *in vitro*. This was however not confirmed *in vivo* in rodent VCA where both cell types increased drug-free long-term allograft survival in a similar manner.
18. Cheng HY, Ghetu N, Huang WC, *et al.* Syngeneic adipose-derived stem cells with short-term immunosuppression induce vascularized composite allotransplantation tolerance in rats. *Cytotherapy* 2014; 16:369–380.
- Interesting study revealing increased long-term allograft survival in rodent VCA after application of syngeneic adipose-derived stem cells. Strikingly, the established tolerance was donor-specific even with the use of syngeneic cells.
19. Kuo YR, Chen CC, Goto S, *et al.* Immunomodulatory effects of bone marrow-derived mesenchymal stem cells in a swine hemi-facial allotransplantation model. *PLoS One* 2012; 7:e35459.
  20. Kuo YR, Chen CC, Goto S, *et al.* Modulation of immune response and T-cell regulation by donor adipose-derived stem cells in a rodent hind-limb allotransplant model. *Plast Reconstr Surg* 2011; 128:661e–672e.
  21. Kuo YR, Chen CC, Goto S, *et al.* Mesenchymal stem cells as immunomodulators in a vascularized composite allotransplantation. *Clin Dev Immunol* 2012; 2012:854846.
  22. Kuo YR, Chen CC, Shih HS, *et al.* Prolongation of composite tissue allotransplant survival by treatment with bone marrow mesenchymal stem cells is correlated with T-cell regulation in a swine hind-limb model. *Plast Reconstr Surg* 2011; 127:569–579.
  23. Kuo YR, Goto S, Shih HS, *et al.* Mesenchymal stem cells prolong composite tissue allotransplant survival in a swine model. *Transplantation* 2009; 87:2009; 87:1769–1777.
  24. Davis TA, Anam K, Lazdun Y, *et al.* Adipose-derived stromal cells promote allograft tolerance induction. *Stem Cells Transl Med* 2014; 3:1444–1450.
  25. Xu H, Ramsey DM, Wu S, *et al.* Simultaneous bone marrow and composite tissue transplantation in rats treated with nonmyeloablative conditioning promotes tolerance. *Transplantation* 2013; 95:301–308.
  26. Pilat N, Klaus C, Schwarz C, *et al.* Rapamycin and CTLA4lg synergize to induce stable mixed chimerism without the need for CD40 blockade. *Am J Transplant* 2015; 15:1568–1579.
  27. Hara H, Lin YJ, Tai HC, *et al.* Hematopoietic chimerism following allotransplantation of the spleen, splenocytes or kidney in pigs. *Transpl Immunol* 2014; 31:125–133.
  28. Leonard DA, Kurtz JM, Mallard C, *et al.* Vascularized composite allograft tolerance across MHC barriers in a large animal model. *Am J Transplant* 2014; 14:343–355.
  29. Graves SS, Mathes DW, Georges GE, *et al.* Long-term tolerance to kidney allografts after induced rejection of donor hematopoietic chimerism in a preclinical canine model. *Transplantation* 2012; 94:562–568.
  30. Kim N, Cho SG. New strategies for overcoming limitations of mesenchymal stem cell-based immune modulation. *Int J Stem Cells* 2015; 8:54–68.
  31. Jiang X, Sun W, Guo D, *et al.* Cardiac allograft acceptance induced by blockade of CD40-CD40L costimulation is dependent on CD4+CD25+ regulatory T cells. *Surgery* 2011; 149:336–346.
  32. Jiang X, Liu C, Hao J, *et al.* CD4(+)CD25(+) regulatory T cells are not required for mesenchymal stem cell function in fully MHC-mismatched mouse cardiac transplantation. *Cell Tissue Res* 2014; 358:503–514.
  33. Crop M, Baan C, Weimar W, Hoogduijn M. Potential of mesenchymal stem cells as immune therapy in solid-organ transplantation. *Transpl Int* 2009; 22:365–376.
  34. Lee SM, Lee SC, Kim SJ. Contribution of human adipose tissue-derived stem cells and the secretome to the skin allograft survival in mice. *J Surg Res* 2014; 188:280–289.
  35. Jeong SH, Ji YH, Yoon ES. Immunosuppressive activity of adipose tissue-derived mesenchymal stem cells in a rat model of hind limb allotransplantation. *Transplant Proc* 2014; 46:1606–1614.
  36. Eggenhofer E, Steinmann JF, Renner P, *et al.* Mesenchymal stem cells together with mycophenolate mofetil inhibit antigen presenting cell and T cell infiltration into allogeneic heart grafts. *Transpl Immunol* 2011; 24:157–163.
  37. Takahashi T, Tibell A, Ljung K, *et al.* Multipotent mesenchymal stromal cells synergize with costimulation blockade in the inhibition of immune responses and the induction of foxp3+ regulatory T cells. *Stem Cells Transl Med* 2014; 3:1484–1494.
- Study depicting the importance of a combined cell-based therapy combined with at least a short course of drug-assisted tolerance induction and maintenance for long-term VCA survival.
38. Larocca RA, Moraes-Vieira PM, Bassi EJ, *et al.* Adipose tissue-derived mesenchymal stem cells increase skin allograft survival and inhibit Th-17 immune response. *PLoS One* 2013; 8:e76396.
  39. Tsuji W, Rubin JP, Marra KG. Adipose-derived stem cells: implications in tissue regeneration. *World J Stem Cells* 2014; 6:312–321.
  40. Singh K, Stempora L, Harvey RD, *et al.* Superiority of rapamycin over tacrolimus in preserving nonhuman primate T<sub>reg</sub> half-life and phenotype after adoptive transfer. *Am J Transplant* 2014; 14:2691–2703.
  41. Ramirez AE, Cheng HY, Lao WW, *et al.* A novel rat full thickness hemi-abdominal wall/hindlimb osteomyocutaneous combined flap: influence of allograft mass and vascularized bone marrow content on vascularized composite allograft survival. *Transpl Int* 2014; 27:977–986.
  42. Obermajer N, Popp FC, Soeder Y, *et al.* Conversion of Th17 into IL-17A(neg) regulatory T cells: a novel mechanism in prolonged allograft survival promoted by mesenchymal stem cell-supported minimized immunosuppressive therapy. *J Immunol* 2014; 193:4988–4999.
  43. De Toni F, Poglio S, Youcef AB, *et al.* Human adipose-derived stromal cells efficiently support hematopoiesis in vitro and in vivo: a key step for therapeutic studies. *Stem Cells Dev* 2011; 20:2127–2138.
  44. Fischer UM, Harting MT, Jimenez F, *et al.* Pulmonary passage is a major obstacle for intravenous stem cell delivery: the pulmonary first-pass effect. *Stem Cells Dev* 2009; 18:683–692.
  45. Zonta S, Martino M, Bedino G, *et al.* Which is the most suitable and effective route of administration for mesenchymal stem cell-based immunomodulation therapy in experimental kidney transplantation: endovenous or arterial? *Transplant Proc* 2010; 42:1336–1340.
  46. Guo R, Ward CL, Davidson JM, *et al.* A transient cell-shielding method for viable MSC delivery within hydrophobic scaffolds polymerized in situ. *Biomaterials* 2015; 54:21–33.
  47. Blocki A, Beyer S, Dewavrin JY, *et al.* Microcapsules engineered to support mesenchymal stem cell (MSC) survival and proliferation enable long-term retention of MSCs in infarcted myocardium. *Biomaterials* 2015; 53:12–24.
  48. Oda H, Konno T, Ishihara K. Efficient differentiation of stem cells encapsulated in a cyto-compatible phospholipid polymer hydrogel with tunable physical properties. *Biomaterials* 2015; 56:86–91.
  49. Kingham PJ, Kolar MK, Novikova LN, *et al.* Stimulating the neurotrophic and angiogenic properties of human adipose-derived stem cells enhances nerve repair. *Stem Cells Dev* 2014; 23:741–754.
  50. Oliveira JT, Bittencourt-Navarrete RE, de Almeida FM, *et al.* Enhancement of median nerve regeneration by mesenchymal stem cells engraftment in an absorbable conduit: improvement of peripheral nerve morphology with enlargement of somatosensory cortical representation. *Front Neuroanat* 2014; 8:111.
  51. Matthes SM, Reimers K, Janssen I, *et al.* Intravenous transplantation of mesenchymal stromal cells to enhance peripheral nerve regeneration. *BioMed Res Int* 2013; 2013:573169.
  52. Halloran PF, Homik J, Goes N, *et al.* The 'injury response': a concept linking nonspecific injury, acute rejection, and long-term transplant outcomes. *Transplant Proc* 1997; 29:79–81.
  53. Schweizer R, Kamat P, Schweizer D, *et al.* Bone marrow-derived mesenchymal stromal cells improve vascular regeneration and reduce leukocyte-endothelium activation in critical ischemic murine skin in a dose-dependent manner. *Cytotherapy* 2014; 16:1345–1360.
  54. Reichenberger MA, Heimer S, Schaefer A, *et al.* Adipose derived stem cells protect skin flaps against ischemia-reperfusion injury. *Stem Cell Rev* 2012; 8:854–862.
  55. Kelahmetoglu O, Demir R, Okten G, *et al.* The effect of mesenchymal stem cells and sildenafil on flap viability in perforator-based flaps for ischemia/reperfusion injury: an experimental study. *Microsurgery* 2015. [Epub ahead of print]
  56. Liu H, Liu S, Li Y, *et al.* The role of SDF-1/CXCR4/CXCR7 axis in the therapeutic effects of hypoxia-preconditioned mesenchymal stem cells for renal ischemia/reperfusion injury. *PLoS One* 2012; 7:e34608.
  57. Xing L, Cui R, Peng L, *et al.* Mesenchymal stem cells, not conditioned medium, contribute to kidney repair after ischemia-reperfusion injury. *Stem Cell Res Ther* 2014; 5:101.

58. Yao W, Lane NE. Targeted delivery of mesenchymal stem cells to the bone. *Bone* 2015; 70:62–65.
59. Liu Y, Yang R, Shi S. Systemic infusion of mesenchymal stem cells improves cell-based bone regeneration via upregulation of regulatory T cells. *Tissue Eng Part A* 2015; 21:498–509.
60. Peng Y, Ke M, Xu L, *et al.* Donor-derived mesenchymal stem cells combined with low-dose tacrolimus prevent acute rejection after renal transplantation: a clinical pilot study. *Transplantation* 2013; 95:161–168.
61. Vanikar AV, Trivedi HL, Gopal SC, *et al.* Pretransplant co-infusion of donor-adipose tissue derived mesenchymal stem cells and hematopoietic stem cells may help in achieving tolerance in living donor renal transplantation. *Ren Fail* 2014; 36:457–460.
62. Perico N, Casiraghi F, Gotti E, *et al.* Mesenchymal stromal cells and kidney transplantation: pretransplant infusion protects from graft dysfunction while fostering immunoregulation. *Transpl Int* 2013; 26:867–878.
63. Mudrabettu C, Kumar V, Rakha A, *et al.* Safety and efficacy of autologous mesenchymal stromal cells transplantation in patients undergoing living donor kidney transplantation: a pilot study. *Nephrology* 2015; 20:25–33.
64. Zhao K, Lou R, Huang F, *et al.* Immunomodulation effects of mesenchymal stromal cells on acute graft-versus-host disease after hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant* 2015; 21:97–104.
65. Introna M, Lucchini G, Dander E, *et al.* Treatment of graft versus host disease with mesenchymal stromal cells: a phase I study on 40 adult and pediatric patients. *Biol Blood Marrow Transpl* 2014; 20:375–381.
66. Sanchez-Guijo F, Caballero-Velazquez T, Lopez-Villar O, *et al.* Sequential third-party mesenchymal stromal cell therapy for refractory acute graft-versus-host disease. *Biol Blood Marrow Transpl* 2014; 20:1580–1585.
67. Holubova M, Lysak D, Vlas T, *et al.* Expanded cryopreserved mesenchymal stromal cells as an optimal source for graft-versus-host disease treatment. *Biologicals* 2014; 42:139–144.
68. Wu Y, Cao Y, Li X, *et al.* Cotransplantation of haploidentical hematopoietic and umbilical cord mesenchymal stem cells for severe aplastic anemia: successful engraftment and mild GVHD. *Stem Cell Res* 2014; 12:132–138.
69. Hequet O, Morelon E, Bourgeot JP, *et al.* Allogeneic donor bone marrow cells recovery and infusion after allogeneic face transplantation from the same donor. *Bone Marrow Transplant* 2008; 41:1059–1061.