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## **Drugs and hepatic transporters: A review**

Jetter, Alexander ; Kullak-Ublick, Gerd A

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1 **Drugs and hepatic transporters: A review**

2 Alexander Jetter, Gerd A. Kullak-Ublick

3 Department of Clinical Pharmacology and Toxicology, University Hospital Zurich, University of Zurich,

4 Rämistrasse 100, CH-8091 Zürich, Switzerland.

5 Email for correspondence: alexander.jetter@usz.ch

6

7

8 **Abstract**

9 The liver is the primary organ for the metabolic degradation of xenobiotics. Transmembrane transport  
10 proteins from the ABC and the SLC families mediate the uptake of endogenous compounds and  
11 xenobiotics into the hepatocyte as well as their elimination from the cells. Multiple processes are  
12 involved. The uptake of xenobiotics in hepatocytes is mediated by organic anion transporting  
13 polypeptides (OATPs) and by organic anion and cation transporters (OATs and OCTs). The  
14 elimination of drugs and metabolites from the liver cell back to the bloodstream is accomplished mainly  
15 by multidrug resistance-associated protein 3 (MRP3) and MRP4, while the elimination towards the  
16 biliary canaliculi is mediated by several different transporters (MRP2, BCRP, MDR1 and MATE1).  
17 Since bile acids and their salts are toxic detergents for hepatocytes, they have to be eliminated  
18 efficiently. This task is accomplished by the bile salt export pump BSEP. Two further transporters,  
19 MDR3 and ATP8B1 are involved in the proper constitution of bile. All these transporters can be  
20 influenced, mainly inhibited by a number of drugs, but also by metabolites from endogenous  
21 compounds such as estrogens. Additionally, rare monogenetic diseases exist which can be explained  
22 by absence of function or dysfunction of specific hepatic transporters, such as progressive familial  
23 intrahepatic cholestasis type 2 by genetic modifications in BSEP that lead to a loss of transporter  
24 function. Functional impairment of other transporters by genetics or by drugs also leads to liver injury,  
25 a potentially life-threatening disease that is still not fully understood. Hence, the interplay between  
26 drugs and hepatic transporters is multiple, and the knowledge of this interplay helps in understanding  
27 the etiology and molecular mechanisms behind some forms of (drug-induced) liver injury.

28

29

## 30 **Introduction**

31 Besides drug metabolizing enzymes, transmembrane drug transporters exert a relevant influence on  
32 the pharmacokinetics, and consequently the pharmacodynamics, of several drugs. These transporters  
33 are responsible for either the uptake of drugs into a cell, or for their extrusion from the cell. Mainly two  
34 families of transmembrane transporters, which are involved in drug transport, exist: ATP-binding  
35 cassette transporters (ABC) and solute carrier transporters (SLC). Transporters from other classes  
36 infrequently also play a role in transmembrane drug transport. While SLC-transporters are either  
37 uptake or bidirectional transporters, transporters from the ABC family mediate efflux of drugs and  
38 metabolites from the cells into the bloodstream or into the bile. Additionally, hepatic transporters may  
39 be inhibited by drugs or their metabolites, while they are not transporting these drugs themselves.

40

## 41 **Drug transporters in the liver**

42 Since the liver is the main organ responsible for drug metabolism, both the uptake and the efflux of  
43 drugs and respective metabolites are in most cases transporter-mediated. The concept of transporter-  
44 mediated uptake of compounds into the hepatocytes was first developed when it was shown that the  
45 uptake of bile salts at the basolateral side was not a mere passive diffusion, but a carrier-mediated,  
46 sodium-dependent process which followed Michaelis-Menten kinetics [1]. Molecular cloning later  
47 identified the sodium-taurocholate cotransporting polypeptide NTCP, gene symbol *SLC10A1*, as the  
48 responsible transporter [2, 3], which not only transports bile salts across the basolateral membrane of  
49 hepatocytes, but also statins [4], and even hepatitis B and D viruses [5, 6].

50

## 51 **Basolateral or sinusoidal uptake transporters**

52 Organic anion transporting polypeptides (OATPs)

53 A number of other transport proteins in the basolateral membrane have been detected, cloned, and  
54 functionally characterized since the discovery of transporter-mediated uptake. Namely different  
55 members of the family of organic anion transporting polypeptides (OATPs) are present in hepatocytes.  
56 Although the first OATP cloned from human liver, OATP1A2 [7], later turned out not to be present in

57 hepatocytes, but to be mainly expressed in the neurons of the hippocampus and the frontal cortex [8],  
58 other members of this family of transporters are strongly expressed in human hepatocytes. The three  
59 OATPs most abundantly expressed in the liver are OATP1B1 (gene symbol *SLCO1B1*), OATP1B3  
60 (gene symbol *SLCO1B3*), and OATP2B1 (gene symbol *SLCO2B1*) [9]. These transporters act  
61 bidirectionally, and can mediate the uptake of amphipathic and anionic substances in exchange with  
62 reduced glutathione or bicarbonate. They have overlapping substrate specificities, which are caused  
63 by the high degree of amino acid homology between the three transporters [10]. Numerous  
64 endogenous compounds are transported into the hepatocyte by OATPs, but also xenobiotics act as  
65 substrates [11]. One of the most important class of drugs which are taken up into the liver cells by  
66 OATPs are statins, HMG-CoA-reductase inhibitors [12]. Inhibition of the OATP-mediated uptake of  
67 statins into the liver cell leads to increased statin concentrations in the bloodstream and may translate  
68 into concentration-dependent adverse effects of statins, such as myopathy. This has been shown e.g.  
69 for ciclosporin [13], or gemfibrozil, although the extent of effect may vary according to the  
70 pharmacokinetic properties of the statin [14]. Besides statins, methotrexate, fexofenadine, some  
71 angiotensin-II-receptor antagonists and angiotensin converting enzyme inhibitors are described as  
72 substrates for OATPs [11, 13, 15]. For OATP1B1, it has been shown that the presence of genetic  
73 variants which decrease transporter function lead to an increase in drug exposure in the blood  
74 (reviewed in [13]). Particularly the c.521T>C SNP (rs4149056) has been shown to increase the AUC  
75 of virtually all statins and hence leads to an increased rate of concentration-dependent adverse effects  
76 like myopathy. In a genome wide association study on markers for simvastatin toxicity which included  
77 more than 300'000 markers in 85 patients with simvastatin-induced myopathy and 90 controls, this  
78 SNP in *OATP1B1* was the only functionally active SNP which was strongly associated with statin-  
79 induced myopathy [16]. Clinically relevant drug-drug interactions due to inhibition of OATPs may also  
80 be expected for several further drugs such as rifampicin [17], octreotide [18] and tyrosine kinase  
81 inhibitors, which have been shown in vitro to inhibit hepatic OATP activity. While most tyrosine kinase  
82 inhibitors are substrates for OATP1B1 and OATP1B3 [19], pazopanib and nilotinib inhibit OATP1B1  
83 with IC<sub>50</sub> values of 3.89±1.21 and 2.78±1.13 µM, respectively [20]. It has to be mentioned, however,  
84 that tyrosine kinase inhibitors are not selective inhibitors of OATPs, but that also OCT1, OAT3 and  
85 other hepatic transporters are inhibited by these drugs, depending on the individual tyrosine kinase  
86 inhibitor [21].

87 There are also inherited diseases linked to OATPs. The human Rotor syndrome, an autosomal  
88 recessive disorder characterized by conjugated hyperbilirubinemia, coproporphyrinuria, and practically  
89 absent hepatic uptake of anionic diagnostic agents, is caused by genetic variants in *OATP1B1* and  
90 *OATP1B3* [22]. Physiologically, bilirubin is conjugated in the hepatocytes mainly by UGT1A1 before a  
91 substantial fraction is excreted back into the bloodstream by the multidrug resistance protein 3 (MRP3,  
92 *ABCC3*), which is also responsible for the canalicular excretion of bilirubin glucuronides. Thereafter,  
93 *OATP1B1* and *OATP1B3* mediate the reabsorption of conjugated bilirubin into the hepatocytes (so-  
94 called “hepatocyte hopping”) [22]. Given the role of human *OATP1B1/1B3* as bilirubin (glucuronide)  
95 uptake transporters, drug-drug interactions at the basolateral entry site of hepatocytes may lead to a  
96 reduced clearance of such endogenous substrates.

97

#### 98 Organic anion transporters (OATs)

99 In addition to the role of OATPs, some members of the organic anion transporters are present in the  
100 sinusoidal membrane of hepatocytes. Besides transporting various endogenous compounds such as  
101 estrone-3-sulfate, cGMP and others, *OAT2 (SLC22A7)* mediates the uptake of different xenobiotics  
102 [23], e.g. entecavir [24] and tolbutamide [25], a clinical marker substrate for CYP2C9 activity [26].  
103 Another OAT, which is present more exclusively in the liver, is *OAT7 (SLC22A9)*, while *OAT2* is also  
104 present in the kidneys. For *OAT7*, one of the few exogenous substrates, which are known to date, is  
105 pravastatin [27].

106

#### 107 Organic cation transporters (OCTs)

108 Like OATs, OCTs are widely distributed throughout tissues and are present mainly in the kidneys, the  
109 liver and the intestines [28]. In humans, *OCT1 (gene symbol SLC22A1)* is present mainly in the liver,  
110 where it mediates the uptake of positively charged hydrophilic compounds. Metformin is a broadly  
111 used antidiabetic which is transported by OCTs, and it has been reported that the *OCT1*-mediated  
112 uptake of metformin can be inhibited by rosiglitazone and repaglinide, two other orally administered  
113 antidiabetics [28].

114

115 **Basolateral efflux transporters**

116 Multidrug resistance-associated proteins (MRP3, MRP4)

117 These basolateral efflux transporters belong to the class of ABC-transporters. While the  
118 aforementioned proteins are located in the basolateral membrane of hepatocytes, MRP2 is located at  
119 the canalicular side (see below). Besides expression in the liver, MRPs are present in many other  
120 tissues with a barrier function such as the lung, the intestinal cells or the blood-brain-barrier [29].  
121 MRP3 (gene symbol *ABCC3*) and MRP4 (gene symbol *ABCC4*) seem to be present in higher  
122 concentrations in the liver. A large range of both endogenous and xenobiotic organic anions is  
123 transported by the MRPs. Since they were first discovered in the research elucidating mechanisms of  
124 resistance of tumor cells against antineoplastic agents, it is not surprising that among the xenobiotics  
125 extruded from the cells by MRPs are vinca alkaloids, methotrexate, alkylating agents, and nucleoside  
126 and nucleotide analogs [30]. Glutathione (GSH) plays an important role in the transport mechanism of  
127 MRP2, which is not yet fully understood. While some xenobiotics are extruded from the cell as GSH-  
128 conjugates, the MRP-mediated transport of others is dependent on, or stimulated by, the presence of  
129 GSH at the transporter site [30]. This stimulation of transport of unmodified drugs by GSH has been  
130 shown e.g. vinblastine via MRP2 [31] in vitro. In contrast to this, MRP3 does not transport GSH or  
131 glutathione-conjugated substances, but shows a preference for glucuronidated compounds [30].  
132 Methotrexate is also transported by MRP3 [29]. In mice, it has been shown that both *Mrp3* and *Mrp4*  
133 are important for the elimination of acetaminophen metabolites from the hepatocytes towards the  
134 bloodstream [32, 33], while *Mrp2* and *Bcrp* eliminate acetaminophen conjugates towards the bile [33].  
135 Additionally, in mice, the efflux of morphine glucuronides to the bloodstream was mediated by *Mrp3*,  
136 while *Mrp2* was the responsible transport mechanism for biliary elimination of morphine 3-glucuronide,  
137 the predominant morphine metabolite [34]. Additionally, MRP3 and MRP4 can also transport bile salts  
138 and their glucuronides to the bloodstream, which constitutes a salvage mechanism in the case that the  
139 main elimination pathway for bile salts via BSEP is impaired [35]. These findings underline the role of  
140 MRPs in the elimination of potentially toxic compounds from the hepatocytes. The expression of MRP4  
141 in the liver seems to be low, as is the case for MRP1 [30]. MRP4 transports a broad range of  
142 xenobiotics and contributes to the elimination of bile acids, uric acid, steroid hormones and cyclic  
143 nucleotides from the liver cell to the sinusoidal blood [36]. Since its expression is upregulated in  
144 cholestasis and other diseases with impaired biliary elimination of organic anions, and since this

145 upregulation is even more pronounced than the upregulation of MRP3, MRP4 may be an important  
146 way of detoxification of bile salts in cholestasis [30].

147

#### 148 **Canalicular efflux transporters**

149 Bile salt export pump (BSEP)

150 The bile salt export pump BSEP (gene symbol *ABCB11*) is the primary transporter for the extrusion of  
151 bile salts into the bile canaliculi [35], which acts against a steep concentration gradient [37]. Bile salts  
152 have detergent properties [38] and may damage mitochondria [39], which leads to cytotoxicity and  
153 liver injury [40]. There appears to be no backup transporter for the canalicular export of bile salts, so  
154 that the inactivation of BSEP leads to intracellular accumulation of bile salts and hence liver damage.  
155 Evidence for this is available both from the bench and from the clinics. An inherited inactivation of  
156 BSEP leads to progressive familial intrahepatic cholestasis (PFIC) type 2 [41]. Mutations in *ABCB11*  
157 causing less severe reductions in BSEP function have been identified as being causative for benign  
158 recurrent intrahepatic cholestasis (BRIC) type 2 [42]. Drugs and / or drug metabolites do not appear to  
159 be transported by BSEP, but may act as BSEP-inhibitors. Since the transporter can also be  
160 competitively inhibited by ciclosporin, rifampicin, glibenclamide, bosentan and a number of other drugs  
161 [35], acquired forms of intrahepatic cholestasis also exist which clinically manifest with elevated  
162 transaminases or even as drug-induced liver injury (DILI). However, besides intrahepatic cholestasis  
163 caused by transporter inhibition, there are other reasons why DILI may develop, and only some of  
164 them are fully understood [43]. A clinically great challenge is the early and specific diagnosis of DILI.  
165 Therefore, current research activities focus on the identification and validation of DILI-specific  
166 biomarkers [44]. The antidiabetic drug troglitazone was withdrawn from the market because of an  
167 elevated incidence of hepatotoxicity. This hepatotoxic potential of troglitazone may be explained by  
168 the BSEP-inhibiting properties of the main metabolite troglitazone sulfate [45]. The troglitazone  
169 example shows nicely that metabolites which are produced intrahepatically may be the causative  
170 agents for inhibition of a transporter function and hence hepatotoxicity. In vitro investigations may  
171 hence fail to show hepatotoxic potential of a drug, if only the parent compound and not the metabolites  
172 are tested. These findings underline the importance of BSEP for the elimination of bile salts. Bile salts  
173 are ligands for the nuclear receptor farnesoid-x-receptor (FXR) and regulate thereby their synthesis,  
174 conjugation and transport [46]. FXR ligand-activators may hence be useful in disease states like

175 cholestatic liver diseases where a FXR-mediated activation of transporter function may be beneficial  
176 [47]. Since bile salts play such an important role in some forms of DILI, they could also be useful as  
177 biomarkers for DILI caused by BSEP-inhibition [48].

178

179 Multidrug resistance-associated protein 2 (MRP2)

180 The multidrug resistance-associated protein 2 (gene symbol *ABCC2*) is another transporter of the  
181 MRP family, which is, in contrast to MRP1, 3, and 4, directed towards the canalicular system.  
182 Substrates for this transporter are drug metabolites, including methotrexate [49], acetaminophen-  
183 glucuronide [33], ezetimibe and etoposide [50]. MRP2 is also expressed in the kidneys. Particularly in  
184 cholestatic conditions, MRP2 is also capable to extrude bile salts from the hepatocytes and thereby  
185 helps in mitigating the toxic effect of high intracellular bile salt concentrations [35]. The function of  
186 MRP2 is relevant for the biliary elimination of bilirubin-glucuronide. This becomes clear e.g. in patients  
187 who have genetic polymorphisms in *ABCC2* which lead, by different mechanisms, to a loss of MRP2  
188 function, and which present with Dubin-Johnson syndrome, a rare, autosomal-recessive, hereditary  
189 disease which presents with conjugated hyperbilirubinemia [51]. Estrogen metabolites are involved in  
190 the development of cholestasis in women taking oral contraceptives, but also in cholestasis of  
191 pregnancy [35]. Estradiol-17 $\beta$ -glucuronide, which is a strongly cholestatic estradiol metabolite in  
192 animal experiments [52], has to be excreted into the bile via MRP2 in order to inhibit BSEP from the  
193 luminal side and thereby blocking the elimination of bile salts [35].

194

195 Breast cancer resistance protein (BCRP)

196 The expression of BCRP (gene symbol *ABCG2*) is not limited to the liver, as is the case for many  
197 other transporters. BCRP shows a broad substrate specificity. As this transporter was first identified in  
198 the context of chemotherapy, the irinotecan metabolite SN-38, topotecan, and doxorubicin are  
199 examples for BCRP substrates [53]. Also newer drugs like the tyrosine-kinase inhibitor sunitinib are  
200 both substrates and inhibitors of BCRP, as it has been shown in humans [54], in rat experiments using  
201 pantoprazole as an inhibitor of BCRP [55] and in cells overexpressing BCRP [56]. However, it has to  
202 be acknowledged that the impact of hepatic BCRP on the overall pharmacokinetics of BCRP  
203 substrates cannot readily be estimated, because BCRP is also present in the intestines and the



204 kidneys, where the effect of BCRP on the pharmacokinetics of drugs is probably much larger than at  
205 the canalicular side of liver cells.

206

207 P-glycoprotein (Multidrug resistance gene product 1;MDR1)

208 P-glycoprotein (gene symbol *ABCB1*) is the most prominent xenobiotic transporter present in virtually  
209 all tissues with barrier function. The transporter, which is also called multidrug resistance protein 1  
210 (MDR1), was first discovered and extensively investigated in the context of resistance of tumor cells  
211 against antineoplastic agents. The transporter mediates the elimination of a broad variety of  
212 xenobiotics from cells, and it shows wide overlap in substrate specificity with other outward-directed  
213 drug transporters such as the MRP-transporters MRP3 (at the basolateral hepatocyte membrane) and  
214 MRP2 (at the canalicular side of hepatocytes) [57]. Additionally, most drugs transported by MDR1 are  
215 also substrates for the most important drug-metabolizing cytochrome P450 enzyme, CYP3A4. The  
216 expression of P-glycoprotein at the canalicular membrane of hepatocytes is sevenfold lower than the  
217 expression in enterocytes of the small intestines, and a considerable interindividual variation has also  
218 been noted [57]. Although more than 100 genetic variants in the *ABCB1* gene are known, clinical  
219 consequences are at best controversial [58]. The most frequent genetic variants C3435T (rs1045642)  
220 or C1236T (rs1128503) do not lead to amino acid exchanges, while the variant G2677T/A (rs2032582)  
221 is responsible for an amino acid exchange which appears to cause relatively small changes in P-  
222 glycoprotein function [58, 59]. Since MDR1 is expressed in many tissues and cells and appears to be  
223 functionally more relevant in the intestines and the blood-brain-barrier in comparison to the  
224 hepatocyte, inhibition of MDR1 function by drugs such as verapamil or ritonavir [57, 60] usually leads  
225 to pharmacokinetic changes of the victim drug, which cannot be attributed to a single expression site.  
226 In summary, it may be possible that MDR1 is not as important in the liver as other drug transporters,  
227 and as MDR1 is important in other tissues such as the enterocytes or the endothelial cells at the  
228 blood-brain-barrier.

229

230 Multidrug resistance gene product 3 (MDR3)

231 The MDR3 transporter (gene symbol *ABCB4*) is a phosphatidylcholine transporter expressed also at  
232 the canalicular membrane of hepatocytes. It translocates phospholipids to the outer leaflet of the

233 membrane lipid bilayer. Since phospholipids are essential in bile in order to solubilize cholesterol and  
234 bile salts in mixed micelles, genetic deficiencies of this transporter lead to a spectrum of cholestatic  
235 liver diseases ranging from transient neonatal cholestasis to biliary cirrhosis in adults, some forms of  
236 cholelithiasis, but also progressive familial intrahepatic cholestasis type 3, intrahepatic cholestasis of  
237 pregnancy, and drug-induced cholestasis [61, 62]. Although the severe loss-of-function variants are  
238 rare, up to 60% of a phenotypically healthy European population presents with genetic variants in the  
239 *ABCB4* gene [63]. Unlike its close relative MDR1, MDR3 appears to be a phospholipid transporter with  
240 no drugs as substrates. However, azole antifungals such as itraconazole, posaconazole and  
241 ketoconazole can act as inhibitors of MDR3 and thereby cause cholestatic liver injury, as it has been  
242 shown using a functional assay to measure MDR3 activity [64]. When the phospholipid excretion into  
243 the bile is inhibited, the bile becomes more toxic because of a reduction in formation of mixed micelles,  
244 which mitigate the toxic effect of bile salts towards the biliary epithelium. A number of other  
245 compounds has been identified as being inhibitors of MDR3. Most of these compounds are known as  
246 causes of drug-induced liver injury and have either been withdrawn from the market or have warnings  
247 in the drug information leaflets [65]. Interestingly, azole antifungals like many of these potentially  
248 hepatotoxic drugs are also inhibitors of BSEP, thereby leading to a dual mechanism by which they can  
249 cause drug-induced liver injury in susceptible patients.

250

#### 251 Type 4 P-type ATPase ATP8B1

252 A third transport mechanism is necessary for appropriate bile formation and avoiding bile salt toxicity:  
253 ATP8B1. This canalicular transporter is a phosphatidylserine translocase or flippase.  
254 Phosphatidylserines serve to make the outer layer of the membrane more resistant against the  
255 detergent properties of bile acids [66]. Rare genetic variants in the ATP8B1 gene lead to a loss of  
256 function of this transporter, which manifests clinically as progressive familial intrahepatic cholestasis  
257 type 1 (PFIC1), also called Byler's disease, or in less severe cases to benign recurrent intrahepatic  
258 cholestasis type 1 (BRIC1) [67, 68].

259

#### 260 Multidrug and toxin extrusion protein 1 (MATE1)

261 MATE transporters are abundantly expressed in the kidneys and play an important role in the tubular  
262 elimination of mainly cationic drugs and endogenous compounds [69]. In the canalicular membrane of  
263 hepatocytes, MATE1 (gene symbol *SLC47A1*) is expressed, but other MATE proteins have not been  
264 found [70]. It has been postulated that MATE1 presents a scavenger transport mechanism to P-  
265 glycoprotein, because it shares various neutral and cationic organic substrates with this transporter  
266 like fexofenadine, levofloxacin and quinidine, and that substrates which are taken up into the liver cell  
267 by OCTs are (at least partially) extruded into the bile by MATE1 [69, 71]. Example substrates for this  
268 latter mechanism are metformin and cimetidine. However, MATE1 plays a more important role in the  
269 kidneys than in the liver, and data on the specific role of MATE1 in the liver is scarce.

270

#### 271 **Further hepatic transmembrane transporters**

272 Besides these transmembrane transport proteins which have been relatively well characterized in the  
273 liver, other transporters such as the equilibrative nucleoside transporters ENT1 and ENT2 or the  
274 organic solute and steroid transporter, OST alpha-OST beta, exist in hepatocytes, but also in intestinal  
275 epithelial cells. The latter transporter is an unusual heterodimer, which is important in bile acid and  
276 steroid homeostasis. This transport mechanism is mainly expressed in enterocytes, but also on the  
277 basolateral membrane of hepatocytes [72]. In hypoxic states, the transport activity of OST alpha-OST  
278 beta is induced [73], and it has been shown that this transporter heterodimer can be transactivated by  
279 FXR [74], the nuclear receptor that mediates bile acid homeostasis.

280

#### 281 **Conclusion**

282 The uptake of endogenous and foreign compounds into the liver cell is closely regulated by a number  
283 of specific transporters, as is the elimination of such compounds and their metabolites, which are  
284 formed intrahepatically towards the bile and back to the bloodstream. A particular function of  
285 hepatocytes is the formation of bile, which is both necessary and toxic because of its contents in bile  
286 salts. If BSEP, the transporter necessary for proper elimination of bile salts from hepatocytes, is  
287 impaired by genetic factors or inhibited by drugs and metabolites, liver injury by intrahepatic  
288 cholestasis develops. Functional impairment of other transporters by genetics or by drugs also leads  
289 to liver injury, a potentially life-threatening disease, which is still not fully understood. Hence, the

290 interplay between drugs and hepatic transporters is multiple, and the knowledge of this interplay helps  
291 in understanding the etiology and molecular mechanisms behind some forms of (drug-induced) liver  
292 injury.

293

294 **Conflict of interest**

295 None of the authors reports a conflict of interest with regard to this publication.

296

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299

- 301 [1] J. Reichen, G. Paumgartner, Uptake of bile acids by perfused rat liver, *Am J Physiol* 231(3) (1976)  
302 734-42.
- 303 [2] B. Hagenbuch, B. Stieger, M. Foguet, H. Lubbert, P.J. Meier, Functional expression cloning and  
304 characterization of the hepatocyte Na<sup>+</sup>/bile acid cotransport system, *Proc Natl Acad Sci U S A* 88(23)  
305 (1991) 10629-33.
- 306 [3] B. Hagenbuch, P.J. Meier, Molecular cloning, chromosomal localization, and functional  
307 characterization of a human liver Na<sup>+</sup>/bile acid cotransporter, *J Clin Invest* 93(3) (1994) 1326-31.
- 308 [4] Y.A. Bi, X. Qiu, C.J. Rotter, E. Kimoto, M. Piotrowski, M.V. Varma, A.F. Ei-Kattan, Y. Lai,  
309 Quantitative assessment of the contribution of sodium-dependent taurocholate co-transporting  
310 polypeptide (NTCP) to the hepatic uptake of rosuvastatin, pitavastatin and fluvastatin, *Biopharm Drug*  
311 *Dispos* 34(8) (2013) 452-61.
- 312 [5] Y. Ni, F.A. Lempp, S. Mehrle, S. Nkongolo, C. Kaufman, M. Falth, J. Stindt, C. Koniger, M. Nassal,  
313 R. Kubitz, H. Sultmann, S. Urban, Hepatitis B and D viruses exploit sodium taurocholate co-  
314 transporting polypeptide for species-specific entry into hepatocytes, *Gastroenterology* 146(4) (2014)  
315 1070-83.
- 316 [6] H. Yan, G. Zhong, G. Xu, W. He, Z. Jing, Z. Gao, Y. Huang, Y. Qi, B. Peng, H. Wang, L. Fu, M.  
317 Song, P. Chen, W. Gao, B. Ren, Y. Sun, T. Cai, X. Feng, J. Sui, W. Li, Sodium taurocholate  
318 cotransporting polypeptide is a functional receptor for human hepatitis B and D virus, *Elife* 1 (2012)  
319 e00049.
- 320 [7] G.A. Kullak-Ublick, B. Hagenbuch, B. Stieger, C.D. Scheuingart, A.F. Hofmann, A.W. Wolkoff, P.J.  
321 Meier, Molecular and functional characterization of an organic anion transporting polypeptide cloned  
322 from human liver, *Gastroenterology* 109(4) (1995) 1274-82.
- 323 [8] B. Gao, S.R. Vavricka, P.J. Meier, B. Stieger, Differential cellular expression of organic anion  
324 transporting peptides OATP1A2 and OATP2B1 in the human retina and brain: implications for carrier-  
325 mediated transport of neuropeptides and neurosteroids in the CNS, *Pflugers Arch* 467(7) (2015) 1481-  
326 1493.
- 327 [9] Z.M. Zair, J.J. Eloranta, B. Stieger, G.A. Kullak-Ublick, Pharmacogenetics of OATP (SLC21/SLCO),  
328 OAT and OCT (SLC22) and PEPT (SLC15) transporters in the intestine, liver and kidney,  
329 *Pharmacogenomics* 9(5) (2008) 597-624.
- 330 [10] G.A. Kullak-Ublick, M.G. Ismail, B. Stieger, L. Landmann, R. Huber, F. Pizzagalli, K. Fattinger,  
331 P.J. Meier, B. Hagenbuch, Organic anion-transporting polypeptide B (OATP-B) and its functional  
332 comparison with three other OATPs of human liver, *Gastroenterology* 120(2) (2001) 525-33.
- 333 [11] A. Kallikowski, M. Niemi, Impact of OATP transporters on pharmacokinetics, *Br J Pharmacol*  
334 158(3) (2009) 693-705.
- 335 [12] P.J. Neuvonen, M. Niemi, J.T. Backman, Drug interactions with lipid-lowering drugs: mechanisms  
336 and clinical relevance, *Clin Pharmacol Ther* 80(6) (2006) 565-81.
- 337 [13] M. Niemi, M.K. Pasanen, P.J. Neuvonen, Organic anion transporting polypeptide 1B1: a  
338 genetically polymorphic transporter of major importance for hepatic drug uptake, *Pharmacol Rev* 63(1)  
339 (2011) 157-81.
- 340 [14] Y. Shitara, M. Hirano, H. Sato, Y. Sugiyama, Gemfibrozil and its glucuronide inhibit the organic  
341 anion transporting polypeptide 2 (OATP2/OATP1B1:SLC21A6)-mediated hepatic uptake and  
342 CYP2C8-mediated metabolism of cerivastatin: analysis of the mechanism of the clinically relevant  
343 drug-drug interaction between cerivastatin and gemfibrozil, *J Pharmacol Exp Ther* 311(1) (2004) 228-  
344 36.
- 345 [15] L. Liu, Y. Cui, A.Y. Chung, Y. Shitara, Y. Sugiyama, D. Keppler, K.S. Pang, Vectorial transport of  
346 enalapril by Oatp1a1/Mrp2 and OATP1B1 and OATP1B3/MRP2 in rat and human livers, *J Pharmacol*  
347 *Exp Ther* 318(1) (2006) 395-402.
- 348 [16] Search Collaborative Group, E. Link, S. Parish, J. Armitage, L. Bowman, S. Heath, F. Matsuda, I.  
349 Gut, M. Lathrop, R. Collins, SLCO1B1 variants and statin-induced myopathy--a genomewide study, *N*  
350 *Engl J Med* 359(8) (2008) 789-99.
- 351 [17] S.R. Vavricka, J. Van Montfoort, H.R. Ha, P.J. Meier, K. Fattinger, Interactions of rifamycin SV  
352 and rifampicin with organic anion uptake systems of human liver, *Hepatology* 36(1) (2002) 164-72.
- 353 [18] M. Visentin, B. Stieger, M. Merz, G.A. Kullak-Ublick, Octreotide inhibits the bilirubin carriers  
354 organic anion transporting polypeptides 1B1 and 1B3 and the multidrug resistance-associated protein  
355 2, *J Pharmacol Exp Ther* 355(2) (2015) 145-51.
- 356 [19] V. Khurana, M. Minocha, D. Pal, A.K. Mitra, Role of OATP-1B1 and/or OATP-1B3 in hepatic  
357 disposition of tyrosine kinase inhibitors, *Drug Metabol Drug Interact* 29(3) (2014) 179-90.
- 358 [20] V. Khurana, M. Minocha, D. Pal, A.K. Mitra, Inhibition of OATP-1B1 and OATP-1B3 by tyrosine  
359 kinase inhibitors, *Drug Metabol Drug Interact* 29(4) (2014) 249-59.

360 [21] R.A. Johnston, T. Rawling, T. Chan, F. Zhou, M. Murray, Selective inhibition of human solute  
361 carrier transporters by multikinase inhibitors, *Drug Metab Dispos* 42(11) (2014) 1851-7.

362 [22] E. van de Steeg, V. Stranecky, H. Hartmannova, L. Noskova, M. Hrebicek, E. Wagenaar, A. van  
363 Esch, D.R. de Waart, R.P. Oude Elferink, K.E. Kenworthy, E. Sticova, M. al-Edreesi, A.S. Knisely, S.  
364 Knoch, M. Jirsa, A.H. Schinkel, Complete OATP1B1 and OATP1B3 deficiency causes human Rotor  
365 syndrome by interrupting conjugated bilirubin reuptake into the liver, *J Clin Invest* 122(2) (2012) 519-  
366 28.

367 [23] H. Shen, Y. Lai, A.D. Rodrigues, Organic Anion Transporter 2: An Enigmatic Human Solute  
368 Carrier, *Drug Metab Dispos* 45(2) (2017) 228-236.

369 [24] T. Furihata, H. Morio, M. Zhu, Y. Suzuki, H. Ide, A. Tsubota, Z. Fu, N. Anzai, K. Chiba, Human  
370 organic anion transporter 2 is an entecavir, but not tenofovir, transporter, *Drug Metab Pharmacokin*  
371 32(1) (2017) 116-119.

372 [25] Y.A. Bi, S. Mathialagan, L. Tylaska, M. Fu, J. Keefer, A. Vildhede, C. Costales, A.D. Rodrigues,  
373 M.V.S. Varma, Organic Anion Transporter 2 Mediates Hepatic Uptake of Tolbutamide, a CYP2C9  
374 Probe Drug, *J Pharmacol Exp Ther* 364(3) (2018) 390-398.

375 [26] A. Jetter, M. Kinzig-Schippers, A. Skott, A. Lazar, D. Tomalik-Scharte, J. Kirchheiner, M.  
376 Walchner-Bonjean, U. Hering, V. Jakob, M. Rodamer, W. Jabrane, D. Kasel, J. Brockmoller, U. Fuhr,  
377 F. Sorgel, Cytochrome P450 2C9 phenotyping using low-dose tolbutamide, *Eur J Clin Pharmacol*  
378 60(3) (2004) 165-71.

379 [27] A. Emami Riedmaier, O. Burk, B.A. van Eijck, E. Schaeffeler, K. Klein, S. Fehr, S. Biskup, S.  
380 Muller, S. Winter, U.M. Zanger, M. Schwab, A.T. Nies, Variability in hepatic expression of organic  
381 anion transporter 7/SLC22A9, a novel pravastatin uptake transporter: impact of genetic and regulatory  
382 factors, *Pharmacogenomics J* 16(4) (2016) 341-51.

383 [28] A.T. Nies, H. Koepsell, K. Damme, M. Schwab, Organic cation transporters (OCTs, MATEs), in  
384 vitro and in vivo evidence for the importance in drug therapy, *Handb Exp Pharmacol* (201) (2011) 105-  
385 67.

386 [29] D. Keppler, Multidrug resistance proteins (MRPs, ABCs): importance for pathophysiology and  
387 drug therapy, *Handb Exp Pharmacol* (201) (2011) 299-323.

388 [30] R.G. Deeley, C. Westlake, S.P. Cole, Transmembrane transport of endo- and xenobiotics by  
389 mammalian ATP-binding cassette multidrug resistance proteins, *Physiol Rev* 86(3) (2006) 849-99.

390 [31] R. Evers, M. de Haas, R. Sparidans, J. Beijnen, P.R. Wielinga, J. Lankelma, P. Borst, Vinblastine  
391 and sulfinpyrazone export by the multidrug resistance protein MRP2 is associated with glutathione  
392 export, *Br J Cancer* 83(3) (2000) 375-83.

393 [32] M.J. Zamek-Gliszczynski, K. Nezasa, X. Tian, A.S. Bridges, K. Lee, M.G. Belinsky, G.D. Kruh,  
394 K.L. Brouwer, Evaluation of the role of multidrug resistance-associated protein (Mrp) 3 and Mrp4 in  
395 hepatic basolateral excretion of sulfate and glucuronide metabolites of acetaminophen, 4-  
396 methylumbelliferone, and harmol in *Abcc3*<sup>-/-</sup> and *Abcc4*<sup>-/-</sup> mice, *J Pharmacol Exp Ther* 319(3) (2006)  
397 1485-91.

398 [33] M.R. McGill, H. Jaeschke, Metabolism and disposition of acetaminophen: recent advances in  
399 relation to hepatotoxicity and diagnosis, *Pharm Res* 30(9) (2013) 2174-87.

400 [34] K. van de Wetering, N. Zelcer, A. Kuil, W. Feddema, M. Hillebrand, M.L. Vlaming, A.H. Schinkel,  
401 J.H. Beijnen, P. Borst, Multidrug resistance proteins 2 and 3 provide alternative routes for hepatic  
402 excretion of morphine-glucuronides, *Mol Pharmacol* 72(2) (2007) 387-94.

403 [35] B. Stieger, Role of the bile salt export pump, BSEP, in acquired forms of cholestasis, *Drug Metab*  
404 *Rev* 42(3) (2010) 437-45.

405 [36] N. Thakkar, J.R. Slizgi, K.L.R. Brouwer, Effect of Liver Disease on Hepatic Transporter  
406 Expression and Function, *J Pharm Sci* 106(9) (2017) 2282-2294.

407 [37] B. Stieger, The role of the sodium-taurocholate cotransporting polypeptide (NTCP) and of the bile  
408 salt export pump (BSEP) in physiology and pathophysiology of bile formation, *Handb Exp Pharmacol*  
409 (201) (2011) 205-59.

410 [38] A.F. Hofmann, L.R. Hagey, Key discoveries in bile acid chemistry and biology and their clinical  
411 applications: history of the last eight decades, *J Lipid Res* 55(8) (2014) 1553-95.

412 [39] S. Krähenbühl, C. Talos, S. Fischer, J. Reichen, Toxicity of bile acids on the electron transport  
413 chain of isolated rat liver mitochondria, *Hepatology* 19(2) (1994) 471-9.

414 [40] S. Tujios, R.J. Fontana, Mechanisms of drug-induced liver injury: from bedside to bench, *Nat Rev*  
415 *Gastroenterol Hepatol* 8(4) (2011) 202-11.

416 [41] E. Jacquemin, Progressive familial intrahepatic cholestasis, *Clin Res Hepatol Gastroenterol* 36  
417 *Suppl* 1 (2012) S26-35.

418 [42] S.W. van Mil, W.L. van der Woerd, G. van der Brugge, E. Sturm, P.L. Jansen, L.N. Bull, I.E. van  
419 den Berg, R. Berger, R.H. Houwen, L.W. Klomp, Benign recurrent intrahepatic cholestasis type 2 is  
420 caused by mutations in ABCB11, *Gastroenterology* 127(2) (2004) 379-84.

421 [43] G.A. Kullak-Ublick, R.J. Andrade, M. Merz, P. End, A. Benesic, A.L. Gerbes, G.P. Aithal, Drug-  
422 induced liver injury: recent advances in diagnosis and risk assessment, *Gut* 66(6) (2017) 1154-1164.

423 [44] R.J. Church, G.A. Kullak-Ublick, J. Aubrecht, H.L. Bonkovsky, N. Chalasani, R.J. Fontana, J.C.  
424 Goepfert, F. Hackman, N.M.P. King, S. Kirby, P. Kirby, J. Marcinak, S. Ormarsdottir, S.J. Schomaker,  
425 I. Schuppe-Koistinen, F. Wolenski, N. Arber, M. Merz, J.M. Sauer, R.J. Andrade, F. van Bommel, T.  
426 Poynard, P.B. Watkins, Candidate biomarkers for the diagnosis and prognosis of drug-induced liver  
427 injury: An international collaborative effort, *Hepatology* (2018).

428 [45] C. Funk, C. Ponelle, G. Scheuermann, M. Pantze, Cholestatic potential of troglitazone as a  
429 possible factor contributing to troglitazone-induced hepatotoxicity: in vivo and in vitro interaction at the  
430 canalicular bile salt export pump (Bsep) in the rat, *Mol Pharmacol* 59(3) (2001) 627-35.

431 [46] J.J. Eloranta, G.A. Kullak-Ublick, The role of FXR in disorders of bile acid homeostasis,  
432 *Physiology (Bethesda)* 23 (2008) 286-95.

433 [47] Z. Gai, M. Visentin, T. Gui, L. Zhao, W.E. Thasler, S. Hausler, I. Hartling, A. Cremonesi, C. Hiller,  
434 G.A. Kullak-Ublick, Effects of Farnesoid X Receptor Activation on Arachidonic Acid Metabolism, NF-kB  
435 Signaling, and Hepatic Inflammation, *Mol Pharmacol* 94(2) (2018) 802-811.

436 [48] H.S. Schadt, A. Wolf, F. Pognan, S.D. Chibout, M. Merz, G.A. Kullak-Ublick, Bile acids in drug  
437 induced liver injury: Key players and surrogate markers, *Clin Res Hepatol Gastroenterol* 40(3) (2016)  
438 257-266.

439 [49] M. Masuda, Y. Iizuka, M. Yamazaki, R. Nishigaki, Y. Kato, K. Ni'inuma, H. Suzuki, Y. Sugiyama,  
440 Methotrexate is excreted into the bile by canalicular multispecific organic anion transporter in rats,  
441 *Cancer Res* 57(16) (1997) 3506-10.

442 [50] C. Fahrmayr, J. Konig, D. Auge, M. Mieth, M.F. Fromm, Identification of drugs and drug  
443 metabolites as substrates of multidrug resistance protein 2 (MRP2) using triple-transfected MDCK-  
444 OATP1B1-UGT1A1-MRP2 cells, *Br J Pharmacol* 165(6) (2012) 1836-1847.

445 [51] A.T. Nies, D. Keppler, The apical conjugate efflux pump ABCC2 (MRP2), *Pflugers Arch* 453(5)  
446 (2007) 643-59.

447 [52] L. Huang, J.W. Smit, D.K. Meijer, M. Vore, Mrp2 is essential for estradiol-17beta(beta-D-  
448 glucuronide)-induced cholestasis in rats, *Hepatology* 32(1) (2000) 66-72.

449 [53] Q. Mao, J.D. Unadkat, Role of the breast cancer resistance protein (BCRP/ABCG2) in drug  
450 transport--an update, *AAPS J* 17(1) (2015) 65-82.

451 [54] T. Mizuno, M. Fukudo, T. Terada, T. Kamba, E. Nakamura, O. Ogawa, K. Inui, T. Katsura, Impact  
452 of genetic variation in breast cancer resistance protein (BCRP/ABCG2) on sunitinib pharmacokinetics,  
453 *Drug Metab Pharmacokinet* 27(6) (2012) 631-9.

454 [55] S. Kunimatsu, T. Mizuno, M. Fukudo, T. Katsura, Effect of P-glycoprotein and breast cancer  
455 resistance protein inhibition on the pharmacokinetics of sunitinib in rats, *Drug Metab Dispos* 41(8)  
456 (2013) 1592-7.

457 [56] C.L. Dai, Y.J. Liang, Y.S. Wang, A.K. Tiwari, Y.Y. Yan, F. Wang, Z.S. Chen, X.Z. Tong, L.W. Fu,  
458 Sensitization of ABCG2-overexpressing cells to conventional chemotherapeutic agent by sunitinib was  
459 associated with inhibiting the function of ABCG2, *Cancer Lett* 279(1) (2009) 74-83.

460 [57] I. Cascorbi, P-glycoprotein: tissue distribution, substrates, and functional consequences of genetic  
461 variations, *Handb Exp Pharmacol* (201) (2011) 261-83.

462 [58] S. Wolking, E. Schaeffeler, H. Lerche, M. Schwab, A.T. Nies, Impact of Genetic Polymorphisms of  
463 ABCB1 (MDR1, P-Glycoprotein) on Drug Disposition and Potential Clinical Implications: Update of the  
464 Literature, *Clin Pharmacokinet* 54(7) (2015) 709-35.

465 [59] M. Schaefer, I. Roots, T. Gerloff, In-vitro transport characteristics discriminate wild-type ABCB1  
466 (MDR1) from ALA893SER and ALA893THR polymorphisms, *Pharmacogenet Genomics* 16(12) (2006)  
467 855-61.

468 [60] C. Wyen, U. Fuhr, D. Frank, R.E. Aarnoutse, T. Klaassen, A. Lazar, A. Seeringer, O.  
469 Doroshenko, J.C. Kirchheiner, F. Abdulrazik, N. Schmeisser, C. Lehmann, W. Hein, E. Schomig,  
470 D.M. Burger, G. Fatkenheuer, A. Jetter, Effect of an antiretroviral regimen containing ritonavir boosted  
471 lopinavir on intestinal and hepatic CYP3A, CYP2D6 and P-glycoprotein in HIV-infected patients, *Clin*  
472 *Pharmacol Ther* 84(1) (2008) 75-82.

473 [61] C. Lang, Y. Meier, B. Stieger, U. Beuers, T. Lang, R. Kerb, G.A. Kullak-Ublick, P.J. Meier, C.  
474 Pauli-Magnus, Mutations and polymorphisms in the bile salt export pump and the multidrug resistance  
475 protein 3 associated with drug-induced liver injury, *Pharmacogenet Genomics* 17(1) (2007) 47-60.

476 [62] E. Gonzales, A. Davit-Spraul, C. Baussan, C. Buffet, M. Maurice, E. Jacquemin, Liver diseases  
477 related to MDR3 (ABCB4) gene deficiency, *Front Biosci (Landmark Ed)* 14 (2009) 4242-56.

478 [63] C. Pauli-Magnus, R. Kerb, K. Fattinger, T. Lang, B. Anwald, G.A. Kullak-Ublick, U. Beuers, P.J.  
479 Meier, BSEP and MDR3 haplotype structure in healthy Caucasians, primary biliary cirrhosis and  
480 primary sclerosing cholangitis, *Hepatology* 39(3) (2004) 779-91.

481 [64] Z.M. Mahdi, U. Sinal-Hermanns, A. Yoker, K.P. Locher, B. Stieger, Role of Multidrug Resistance  
482 Protein 3 in Antifungal-Induced Cholestasis, *Mol Pharmacol* 90(1) (2016) 23-34.  
483 [65] M.D. Aleo, F. Shah, K. He, P.D. Bonin, A.D. Rodrigues, Evaluating the Role of Multidrug  
484 Resistance Protein 3 (MDR3) Inhibition in Predicting Drug-Induced Liver Injury Using 125  
485 Pharmaceuticals, *Chem Res Toxicol* 30(5) (2017) 1219-1229.  
486 [66] D.E. Folmer, R.P. Elferink, C.C. Paulusma, P4 ATPases - lipid flippases and their role in disease,  
487 *Biochim Biophys Acta* 1791(7) (2009) 628-35.  
488 [67] L.N. Bull, R.J. Thompson, Progressive Familial Intrahepatic Cholestasis, *Clin Liver Dis* 22(4)  
489 (2018) 657-669.  
490 [68] G.A. Kullak-Ublick, P.J. Meier, Mechanisms of cholestasis, *Clin Liver Dis* 4(2) (2000) 357-85.  
491 [69] K. Damme, A.T. Nies, E. Schaeffeler, M. Schwab, Mammalian MATE (SLC47A) transport  
492 proteins: impact on efflux of endogenous substrates and xenobiotics, *Drug Metab Rev* 43(4) (2011)  
493 499-523.  
494 [70] M. Otsuka, T. Matsumoto, R. Morimoto, S. Arioka, H. Omote, Y. Moriyama, A human transporter  
495 protein that mediates the final excretion step for toxic organic cations, *Proc Natl Acad Sci U S A*  
496 102(50) (2005) 17923-8.  
497 [71] K.M. Hillgren, D. Keppler, A.A. Zur, K.M. Giacomini, B. Stieger, C.E. Cass, L. Zhang, C.  
498 International Transporter, Emerging transporters of clinical importance: an update from the  
499 International Transporter Consortium, *Clin Pharmacol Ther* 94(1) (2013) 52-63.  
500 [72] N. Ballatori, N. Li, F. Fang, J.L. Boyer, W.V. Christian, C.L. Hammond, OST alpha-OST beta: a  
501 key membrane transporter of bile acids and conjugated steroids, *Front Biosci (Landmark Ed)* 14  
502 (2009) 2829-44.  
503 [73] C.A. Schaffner, J. Mwinyi, Z. Gai, W.E. Thasler, J.J. Eloranta, G.A. Kullak-Ublick, The organic  
504 solute transporters alpha and beta are induced by hypoxia in human hepatocytes, *Liver Int* 35(4)  
505 (2015) 1152-61.  
506 [74] J.F. Landrier, J.J. Eloranta, S.R. Vavricka, G.A. Kullak-Ublick, The nuclear receptor for bile acids,  
507 FXR, transactivates human organic solute transporter-alpha and -beta genes, *Am J Physiol*  
508 *Gastrointest Liver Physiol* 290(3) (2006) G476-85.

509