



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2019

Regulation of Glucose Metabolism by NAD⁺ and ADP-Ribosylation

Hopp, Ann-Katrin ; Grüter, Patrick ; Hottiger, Michael O

DOI: <https://doi.org/10.3390/cells8080890>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-186286>

Journal Article

Published Version



The following work is licensed under a Creative Commons: Attribution 4.0 International (CC BY 4.0) License.

Originally published at:

Hopp, Ann-Katrin; Grüter, Patrick; Hottiger, Michael O (2019). Regulation of Glucose Metabolism by NAD⁺ and ADP-Ribosylation. *Cells*, 8(8):pii: E890.

DOI: <https://doi.org/10.3390/cells8080890>

Review

Regulation of Glucose Metabolism by NAD⁺ and ADP-Ribosylation

Ann-Katrin Hopp ^{1,2}, Patrick Grüter ¹ and Michael O. Hottiger ^{1,*} 

¹ Department of Molecular Mechanisms of Disease (DMMD), University of Zurich, CH-8057 Zurich, Switzerland

² Molecular Life Science Ph.D. Program, Life Science Zurich Graduate School, CH-8057 Zurich, Switzerland

* Correspondence: michael.hottiger@dmmd.uzh.ch

Received: 18 July 2019; Accepted: 11 August 2019; Published: 13 August 2019



Abstract: Cells constantly adapt their metabolic pathways to meet their energy needs and respond to nutrient availability. During the last two decades, it has become increasingly clear that NAD⁺, a coenzyme in redox reactions, also mediates several ubiquitous cell signaling processes. Protein ADP-ribosylation is a post-translational modification that uses NAD⁺ as a substrate and is best known as part of the genotoxic stress response. However, there is increasing evidence that NAD⁺-dependent ADP-ribosylation regulates other cellular processes, including metabolic pathways. In this review, we will describe the compartmentalized regulation of NAD⁺ biosynthesis, consumption, and regeneration with a particular focus on the role of ADP-ribosylation in the regulation of glucose metabolism in different cellular compartments.

Keywords: carbohydrate metabolism; ADP-ribosylation; ARTD; PARP; NAD⁺; NR; NMN; NAM

1. Introduction

1.1. NAD⁺ in Redox Reactions

Metabolic pathways are tightly regulated on various levels, which include co-enzyme availability, allosteric regulation with various metabolic intermediates, and post-translational modification of metabolic enzymes. Nicotinamide adenine dinucleotide (NAD⁺, in its oxidized state) is a vital small molecule, best known as a cofactor that regulates metabolism through its electron transfer function in redox reactions that regulate glycolysis, tricarboxylic acid (TCA) cycle, and oxidative phosphorylation driven energy metabolism [1–3]. In redox reactions, NAD⁺ is reversibly reduced to NADH [4]. NAD⁺ can be regenerated in the cytoplasm from NADH through the conversion of pyruvate to lactate by the lactate dehydrogenase (LDH), thereby maintaining active aerobic glycolysis [5]. Alternatively, electrons from NADH can enter the mitochondrial electron transport chain via the glycerol-3-phosphate or the malate-aspartate shuttle [6,7]. Within the mitochondria, NAD⁺ plays a central role by carrying high-energy electrons and driving oxidative phosphorylation [8]. NAD⁺ accepts electrons from different sources and transfers them to complex I of the electron transport chain, which results in the generation of ATP. The maintenance of NAD⁺ levels, as well as the NAD⁺/NADH ratio, is crucial for mitochondrial homeostasis and ATP production.

1.2. Cellular NAD⁺ Pools

The in-vivo half-life of NAD⁺ varies between 15 min and 15 h depending on the tissue [9]. Decreased NAD⁺ levels dampen the activities of NAD(H)-dependent enzymes involved in oxidative phosphorylation, TCA cycle, and glycolysis, which lowers the ATP production [10]. Several approaches have been developed to try to quantify NAD⁺ availability in different subcellular compartments.

Compartment-targeted expression of the catalytic domain of ADP-ribosyltransferase diphtheria toxin-like 1 (ARTD1, formerly called PARP1) has been used as a molecular detector of NAD⁺ because formation of poly-ADP-ribose, which can be detected by immunofluorescence, functions as a proxy for NAD⁺ availability [11,12]. In addition, NAD⁺ metabolite tracing methods have been developed that quantify NAD⁺ and NADH levels, and define how their ratios in different subcellular compartments under several stress conditions change [11,13–15]. Free NAD⁺ has been measured in almost all organelles, including the mitochondria, peroxisomes, the endoplasmic reticulum (ER), and the Golgi complex [11–13]. These measurements have revealed that steady state NAD⁺ concentrations, as well as changes in NAD⁺ concentrations in response to different stimuli, vary considerably between different subcellular compartments. Hence, NAD⁺ dynamics seem to be compartment specific. Furthermore, these dynamics also vary depending on the cell type, stress, and redox status as well as metabolic fitness [16]. The NAD⁺ concentrations are highest in the mitochondria (approx. 400 μM, 40%–70% of the total cellular NAD⁺ pool [17,18]), intermediate in the nucleus and cytoplasm (approx. 100 μM) and low (<1 μM) in the extracellular space [13,15,19]. Cytoplasmic and nuclear NAD⁺/NADH ratios are typically between 60 and 700 in eukaryotes depending on the cell type, while the mitochondrial ratio is much lower at around 7–8 [20,21]. Nuclear and cytoplasmic NAD⁺ pools are interconnected as NAD⁺ levels in both compartments are generally comparable and NAD⁺ precursors freely diffuse through the nuclear pore complex. In contrast, NAD⁺, NADH and NAD⁺ precursors cannot freely diffuse across mitochondrial membranes, and thus mitochondrial NAD⁺ pools are more isolated [22]. This compartmentalization of NAD⁺-levels provides a special level of regulation and sophistication that requires tightly regulated expression of enzymes involved in NAD⁺ biogenesis.

1.3. NAD⁺ Biogenesis/Synthesis

NAD⁺ is constantly synthesized, degraded, and recycled, not only in the cytoplasm where most research is focused, but also within major organelles including the nucleus, Golgi, and peroxisomes [23,24]. In mammals, NAD⁺ is generated either de-novo via the kynurenine pathway from tryptophan, via the Preiss-Handler pathway from nicotinic acid (NA), via the salvage pathway from nicotinamide (NAM), or via the nicotinamide ribose kinase pathway from nicotinamide riboside (NR) [25–28]. Many enzymes involved in the biosynthesis of NAD⁺ are highly compartmentalized on the subcellular level and tightly regulated on the transcriptional level, which provides the rationale as to why some tissues and/or cell types are able to synthesize NAD⁺ from certain sources and others are not. A detailed overview of the NAD⁺ metabolic pathways has been extensively described in other reviews [1,25,29,30].

In-vivo, the de-novo synthesis of NAD⁺ from tryptophan takes place predominantly in the liver. The liver then excretes NAM which is subsequently taken up by other organs and converted to NAD⁺. NAD⁺ fluxes vary widely across tissues, with high fluxes in the small intestine and spleen and low fluxes in skeletal muscle [9]. Intravenous administration of NR or NMN delivered NAD⁺ to multiple tissues, but the same agents given orally were metabolized to NAM, indicating that the type of administration and the involved organs metabolize NAD⁺ precursors differently [9]. In-depth analysis of NAD⁺ synthesis-breakdown fluxes in different cell lines revealed that the vast majority of immortalized cells lack the enzymes to synthesize NAD⁺ from tryptophan, and thus they depend entirely on NAM [9]. Consequently, most cell lines depend on two enzymes to generate NAD⁺. The first is nicotinamide phosphoribosyltransferase (NAMPT) which catalyzes the conversion of NAM to NMN; this is considered the rate limiting step of the NAD⁺ salvage pathway [31]. The second family of enzymes required are the nicotinamide mononucleotide adenylyltransferases (NMNATs), which convert NMN to NAD⁺. NAMPT is primarily nuclear and cytosolic; however, a small portion co-purifies with mitochondria isolated from the liver [32]. There are both intracellular and extracellular isoforms of NAMPT that are known as iNAMPT and eNAMPT, respectively [33]. Three isoforms of NMNAT have been identified; NMNAT1 localizes to the nucleus, NMNAT2 to the Golgi apparatus and neuronal axons, and NMNAT3 to the mitochondria [34–36]. Given that both NAMPT and NMNAT3 localize to the mitochondria, it has been suggested that mitochondria might recycle their own nicotinamide or

take it up from the cytoplasm. Increasing evidence has revealed that (i) reduced NAD⁺ levels alter metabolic rates and increase age-related disease susceptibility and (ii) restoration of NAD⁺ levels can prevent disease progression [37,38].

1.4. NAD⁺ Signaling

Within a given location, NAD⁺ has two main pools, the *free* pool and *protein-associated* pool. Moreover, the ratio of these pools varies across different organelles, cell types, and tissues [29]. NAD⁺ is predicted to bind >500 proteins involved in the regulation of almost all major biological processes [39,40]. Two principal types of NAD⁺-dependent signaling reactions have recently been described: the generation of second messenger molecules and the modification of proteins [41,42]. NAD⁺ is a direct precursor of various small molecules such as ADP-ribose (ADPr), cyclic ADP-ribose (cADPr), NAADP and O-acetyl-ADP-ribose (OAADPR); all of which are important second messengers that regulate multiple aspects of biology, including cell survival, apoptosis, and inflammation [39,43–48]. In addition, NAD⁺ serves as a substrate for post-translational protein modifications (PTM). In mammals, the two main NAD⁺-dependent enzyme families are the ADP-ribosyltransferases (ARTs) and the sirtuins (SIRTs) [39,49–52]. Both protein families use NAD⁺ as a co-substrate to modify or de-modify target proteins. Interestingly, ADP-ribosylation was the first NAD⁺-dependent PTM identified [53]. ADP-ribosylation involves the attachment of one (mono-ADP-ribosylation; MARylation) or several (poly-ADP-ribosylation; PARylation) moieties of ADPr onto specific amino acid acceptor sites of target proteins or onto ribonucleotides [54]. Depending on their structural similarities to either cholera- or diphtheria toxins, ARTs can be further subdivided into the intracellular ARTDs (Diphtheria toxin-like) and the extracellular or membrane-associated ARTCs (Cholera toxin-like). Despite the very low NAD⁺ concentrations in these compartments [55], ARTCs as well as cADPr hydrolases, such as CD38, can metabolize NAD⁺ in the extracellular space and in serum. Since this review focuses on the influence of ADP-ribosylation on carbohydrate metabolism the reader is referred to other reviews describing extracellular NAD⁺ consumption [45,56]. SIRTs remove acyl marks (most commonly acetylation) from proteins using NAD⁺ as an acceptor, thereby generating OAADPR and NAM [39,57]. SIRTs and ARTs localize to various intracellular compartments and have been experimentally linked to distinct cellular functions. Both enzyme families are associated with the regulation of various physiological processes, including metabolic regulation, DNA damage repair, cell cycle progression, and epithelial-to-mesenchymal transition (EMT) [58–62].

It is important to note that when functioning as a coenzyme, for instance in redox reactions, NAD⁺ is reversibly converted to NADH. In contrast, when NAD⁺ functions as a signaling molecule in NAD⁺-dependent signaling processes, NAD⁺ but not NADH, is used as a substrate [63] and is continuously and irreversibly catabolized by cleavage of the glycosidic bond between NAM and the ADPr moiety. In cells, the involvement of NAD⁺ in deacetylation reactions or ADP-ribosylation requires constant re-synthesis of NAD⁺ to avoid depletion of intracellular NAD⁺ pools, which is particularly challenging for rapidly dividing cells [9].

NAM and ADPr are the common products generated as a result of NAD⁺-dependent signaling conversions. In human cells, ADPr can be cleaved by several members of the Nudix hydrolase family [64]. Among these enzymes, NUDT9 appears to have the highest specificity towards ADPr generating AMP and ribose 5-phosphate [65–67]. The recycling of these degradation products would require three molecules of ATP to regenerate NAD⁺. Therefore, NAD⁺-dependent signaling events, especially PARylation, are rather energy consuming.

1.5. ADP-Ribosylation and Carbohydrate Metabolism

ADP-ribosylation is considered energetically challenging for the cell, as each ADPr moiety attached to a given target protein requires the consumption of one NAD⁺ molecule. Therefore, ADPr-metabolizing enzymes such as ARTs and ADP-ribosylhydrolases (ARHs) have a very strong impact on intracellular NAD⁺ homeostasis [9]. Indeed, an anti-correlation between the activity of

the predominant nuclear ART, ARTD1, and intracellular NAD^+ levels has been the subject of various studies [68–72]. It is, thus, not surprising that ART activation has been linked to cell metabolism in various physiological and pathophysiological processes, including adipogenesis, genotoxicity-induced cell death, immune cell activation, and metabolic disorders [68–73]. In fact, the activity of these enzymes can influence glycolysis and oxidative phosphorylation directly and indirectly at various steps (see below). NAD^+ and NADH are important redox-equivalents for various metabolic reactions, like the conversion of pyruvate to lactate or the conversion of succinate to fumarate and back (Figure 1). In addition, ADP and ATP are important co-factors as their ratio is strongly dependent on oxidative phosphorylation and, thus, NAD^+ . As a consequence, extensive ART activation decreases NAD^+ and ATP levels, and indirectly influences anabolic and catabolic reactions involved in central carbohydrate metabolism. There is, however, growing evidence that ARTs also regulate the carbohydrate metabolism by directly modifying important regulators of the involved metabolic pathways. ADP-ribosylation has also been proposed to play a role in other metabolic processes, such as the lipid metabolism (reviewed in [74]). ADP-ribosylated proteins involved in glucose metabolism with assigned ADPr amino acid acceptor sites are summarized in Table 1.

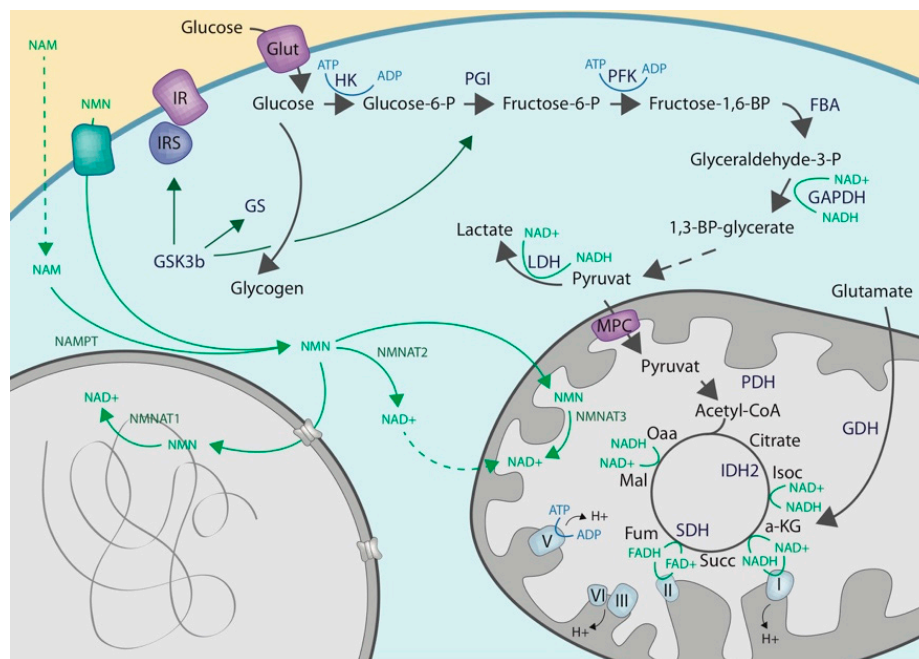


Figure 1. Schematic overview over the main metabolic hubs comprising the central carbohydrate metabolism. Glycolysis, TCA cycle and oxidative phosphorylation are depicted in black (black arrows), while enzymes involved in those processes are depicted in dark blue. NAD^+ metabolism, including synthesis and consumption is depicted in green (green descriptions and arrows), while ADP/ATP conversions are depicted in light blue. Glucose-6-phosphate (Glucose-6-P), fructose-6-phosphate (Fructose-6-P), fructose-1,6-bisphosphate (Fructose-1,6-BP), glyceraldehyde-3-phosphate (Glyceraldehyde-3-P), oxaloacetate (OAA), isocitrate (Isoc), a-ketoglutarate (a-KG), succinate (Succ), fumarate (Fum), malate (Mal), hexokinase (HK), phosphoglucose isomerase (PGI), phosphofructokinase (PFK), aldolase (FBA), lactate dehydrogenase (LDH), pyruvate dehydrogenase (PDH), glutamate dehydrogenase (GDH), insulin receptor (IR), insulin receptor signaling (IRS).

Table 1. Overview of ADP-ribosylated proteins involved in glucose metabolism with assigned ADP-ribose amino acid acceptor site and their function.

Protein	Modification Side	Function	Function Affected by ADPR	Localization	References
ATP5A1	E508, K506 (m) S184, S513 (h)	ATP synthesis	N/A	m	[75,76]
ATP5B	E98, R121 (m)	ATP synthesis	N/A	m	[75]
ATP5F1	H164, S226 (h)	ATP synthesis	N/A	m	[76]
ATP5F1B	S415, R458, H477 (h)	ATP synthesis	N/A	m	[76]
ATP5F1C	S195 (h)	ATP synthesis	N/A	m	[76]
ATP50	R29, S126 (m) S163, S166 (h)	ATP synthesis	N/A	m	[75,76]
C/EBP-b	E135, K133, E139 (m) S141, S257 (h)	Transcription factor, gene expression	Yes (activity dampened)	n	[76,77]
COX4I1	H51 (h)	Electron transport chain	N/A	m	[76]
CYCS	S34 (h)	Electron transport chain	N/A	m	[76]
GAPDH	R198, R232 (m) R80, S122, S192, R197, S210 (h)	Glycolysis	N/A	c, n	[75,76]
MDH1	S241 (h)	Carbohydrate metabolism	N/A	m	[76]
MDH2	S261, S246 (m) S317 (h)	Carbohydrate metabolism	N/A	m	[75,76]
NDUFB1	R49 (m)	Electron transport	N/A	m	[75]
NDUFB5	R93 (m)	Electron transport	N/A	m	[75]
NDUFAF7	R415 (m)	Electron transport	N/A	m	[75]
NDUFC2	R120 (m)	Electron transport	N/A	m	[75]
NDUFAB1	S99 (h)	Electron transport	N/A	m	[76]
NDUFV1	R449 (m)	Electron transport	N/A	m	[75]
PDPR	R866 (m)	Carbohydrate metabolism	N/A	m	[75]
PDHA1	R304 (m)	Carbohydrate metabolism	N/A	m	[75]
PDHX	S131 (m)	Carbohydrate metabolism	N/A	m	[75]
SDHA	S505, H522 (h)	Electron transport	N/A	m	[76]
UQCRC1	S221 (h)	Electron transport	N/A	m	[76]
UQCRC2	R241 (m) S221 (h)	Electron transport	N/A	m	[75,76]
UQCRCF1	E95 (m)	Electron transport	N/A	m	[75]

Abbreviations: R: arginine, S: serine, K: lysine, H: histidine, E: glutamic acid, c: cytoplasm, m: mitochondria, n: nucleus, (m): mouse, (h): human, N/A: not available.

2. Cytoplasmic Crosstalk of ADP-Ribosylation and the Carbohydrate Metabolism

2.1. Cytoplasmic NAD Biosynthesis

Many enzymes involved in NAD⁺ biosynthesis, such as NAMPT and the NMNATs, and breakdown, like the ARTs and SIRT, are highly compartmentalized on the subcellular level, providing yet another layer of NAD⁺ level regulation ([16]; Table 2).

The enzymes involved in the kynurenine and the Preiss-Handler pathway all localize to the cytoplasm. Therefore, de-novo NAD⁺ synthesis predominantly happens in this compartment. With NAMPT and NMNAT2 the cytoplasm also harbors a set of enzymes required for NAD⁺ recycling from NAM and NMN. As mentioned, for most transformed cells the salvage pathway is the main source of NAD⁺ synthesis. In those cells, the levels of NAMPT are highly dynamic and respond to changes in cellular demands for NAD⁺, such as starvation or severe DNA damage, for instance [32]. NMNAT2 catalyzes the final reaction, converting NMN to NAD⁺ in the cytoplasm. In the cytoplasm, NAD⁺ plays an important electron carrier role in the glycolysis, as it is required for the conversion of glyceraldehyde-3-phosphate to 1,3-bisphosphoglycerate, as well as for the conversion of pyruvate to lactate. In fact, reductions in cytoplasmic NAD⁺ levels are associated with reduced glycolysis and the necessity of cells to metabolize non-glucose carbon sources, such as glutamate or pyruvate via the TCA cycle [78].

Table 2. Overview of the localization of enzymes involved in NAD⁺ synthesis and conversion.

Nucleus				
Family	Name	NAD ⁺ Metabolism	ART/ARH Activity	References
ARTs	ARTD1	Consumption	Poly (branching)	[79–81]
	ARTD2	Consumption	Poly (branching)	[79–81]
	ARTD3	Consumption	Mono	[79–81]
	ARTD4	Consumption	Mono	[79–81]
	ARTD5	Consumption	Poly/Oligo	[79–81]
	ARTD6	Consumption	Poly/Oligo	[79–81]
	ARTD8	Consumption	Mono	[79–81]
	ARTD9	Consumption	Inactive/Mono	[79,80,82]
	ARTD10	Consumption	Mono	[79–81]
	ARTD11	Consumption	Mono	[79–81]
ARTD14	Consumption	Mono	[79–81]	
ARHs	PARG		Poly	[83,84]
	ARH3		Poly/Mono	[85–87]
	TARG		Mono	[88–90]
SIRTs	SIRT1	Consumption	N/A	[39]
	SIRT2	Consumption	N/A	[39]
	SIRT6	Consumption	Mono	[39,91]
	SIRT7	Consumption	N/A	[39]
NAMPT	NAMPT	Synthesis	N/A	[2]
NMNAT	NMNAT1	Synthesis	N/A	[2]
Cytoplasm				
Family	Name	NAD ⁺ Metabolism	ART/ARH Activity	References
ARTs	ARTD2	Consumption	Poly (branching)	[79–81]
	ARTD3	Consumption	Mono	[79–81]
	ARTD4	Consumption	Mono	[79–81]
	ARTD5	Consumption	Poly/Oligo	[79–81]
	ARTD6	Consumption	Poly/Oligo	[79–81]
	ARTD7	Consumption	Mono	[79–81]
	ARTD8	Consumption	Mono	[79–81]
	ARTD9	Consumption	Inactive/Mono	[79,80,82]
	ARTD10	Consumption	Mono	[79–81]
	ARTD11	Consumption	Mono	[79–81]
	ARTD12	Consumption	Mono	[79–81]
	ARTD13	Consumption	Inactive	[79–81]
	ARTD14	Consumption	Mono	[79–81]
	ARTD15	Consumption	Mono	[79–81]
	ARTD16	Consumption	Mono	[79–81]
	ARTD17	Consumption	Mono	[79–81]
	ARTD18	Consumption	Mono	[79–81]
	ARHs	PARG		Poly
ARH1			Mono	[85]
ARH2			Inactive	[85]
MacroD2			Mono	[88–90]
SIRTs	SIRT1	Consumption	N/A	[39]
	SIRT2	Consumption	N/A	[39]
NAMPT	NAMPT	Synthesis	N/A	[2]
NMNAT	NMNAT2	Synthesis	N/A	[2]
Mitochondria				
Family	Name	NAD ⁺ Metabolism	ART/ARH Activity	References
ARTs	ARTD1	Consumption	Poly (branching)	[79,80]
ARHs	PARG		Poly	[83,84,92]
	ARH3		Poly/Mono	[85–87]
	MacroD1		Mono	[88–90,93]
SIRTs	SIRT3	Consumption	N/A	[39]
	SIRT4	Consumption	Mono	[39,94]
	SIRT5	Consumption	N/A	[39]
NAMPT	NAMPT	Synthesis	N/A	[2]
NMNAT	NMNAT3	Synthesis	N/A	[2]

Abbreviations: ART: ADP-ribosyltransferase, ARH: ADP-ribosylhydrolase, SIRT: Sirtuin, NAMPT: Nicotinamide phosphoribosyltransferase, NMNAT: Nicotinamide mononucleotide adenylyltransferase, N/A: Not available

2.2. Cytoplasmic ADP-Ribosylation

Many ARTD-family members, especially ARTDs catalyzing MARYlation are described to localize to the cytoplasm (Table 2) [49]. Nonetheless, potential functions of ADP-ribosylation in the cytoplasm are rather poorly described. The best-studied ARTDs localizing to the cytoplasm are ARTD10, ARTD12, ARTD15 and the tankyrases (TNKs) [95–100]. ARTD10 and the TNKs have been linked to cytoplasmic metabolic processes, while the other cytoplasmic ARTDs have thus far not been directly associated with cell/carbohydrate metabolism. ARTD12 localizes to the Golgi and is involved in protein sorting and trafficking [95,96], while ARTD15 localizes to the ER and has been linked to the unfolded protein response [97,98]. For more detailed information about the function of other (cytoplasmic) ARTs, the reader is referred to other reviews [101]. Nothing has so far been reported regarding the regulation of the carbohydrate metabolism by cytoplasmic ARHs.

2.3. Crosstalk with Enzymes of the Carbohydrate Metabolisms

Glut4: Glut4 is one of 14 Glut transporters described in mammals [102]. Glut transporters are transmembrane proteins that are able to transport hexoses from the extracellular space into cells. Specifically, Glut4 transports glucose and glucosamine, and is predominantly expressed in adipocytes, skeletal muscle and cardiomyocytes [102,103]. While some glucose transporters, such as Glut1, constitutively localize to the cell membrane, Glut4 translocates to the plasma membrane in response to insulin stimulation [103]. Both TNK1 and TNK2 have been shown to influence Glut4 mediated glucose transport in an ADP-ribosylation-dependent manner [99,104]. While in adipocytes, *tnks* deficiency increased Glut4 levels, TNKs inhibition in myocytes destabilized Glut4 regulatory proteins, resulting in decreased Glut4 translocation to the plasma membrane [99,104]. However, in both studies *tnks* deficiency/inhibition affected insulin sensitivity. Furthermore, the use of TNK inhibitors supported the notion that this process is dependent on ADP-ribosylation; specific target proteins of TNK1/2, however, have not yet been identified.

Hexokinase 1: Hexokinases catalyze the initial step of glycolysis by converting glucose to glucose-6-phosphate (Figure 2). In mammals, 4 isoenzymes, HK1, 2, 3, and 4 have been identified [105]. HK1 was shown to contain a PAR binding motif (PBM) that enables the enzyme to interact with PAR in a non-covalent manner [106]. Under normal conditions HK1 localizes to the mitochondrial membrane. Binding of HK1 to free cytoplasmic PAR, e.g., in the context of ARTD1 hyperactivation upon severe genotoxic stress, was proposed to lead to the segregation of HK1 from the mitochondria and to reduce its enzymatic activity in a dose-dependent manner, consequently slowing down glycolysis and oxidative phosphorylation [106]. This observation was strengthened by the observation that inhibition of PAR release by PARG knockdown rescued HK1 activity and the cells' metabolic capacity. Together, the above-discussed study proposes a mechanism in which extensive nuclear ARTD1-mediated PAR formation results in a reduction in the enzymatic activity of HK1, which is independent on direct ADP-ribosylation, but mediated via PAR binding.

Glycogen synthase kinase 3b: The glycogen synthase kinase 3b (GSK3b) has also been proposed to be regulated by ADP-ribosylation [100]. GSK3b is a serine-threonine-kinase that regulates glucose metabolism by controlling the activity of various enzymes involved in anabolic and catabolic carbohydrate metabolism, including the glycogen synthase, the insulin receptor, the glucose-6-phosphatase, and/or the phosphoenolpyruvate carboxykinase [107,108] (Figure 2). A recent study identified GSK3b as a direct ADP-ribosylation target of ARTD10 [100]. Intriguingly, studies also suggest that ARTD10 negatively regulates cell metabolism and mitochondrial function, as ARTD10 knockdown increased the respiratory capacities of various cell lines and reduced reactive oxygen species susceptibility via induction of antioxidant gene expression [109]. In addition, ARTD10 levels were also shown to change in response to fasting-induced metabolic stimulation [109]. A direct link between ARTD10-mediated ADP-ribosylation and cell metabolism, however, remains to be elucidated.

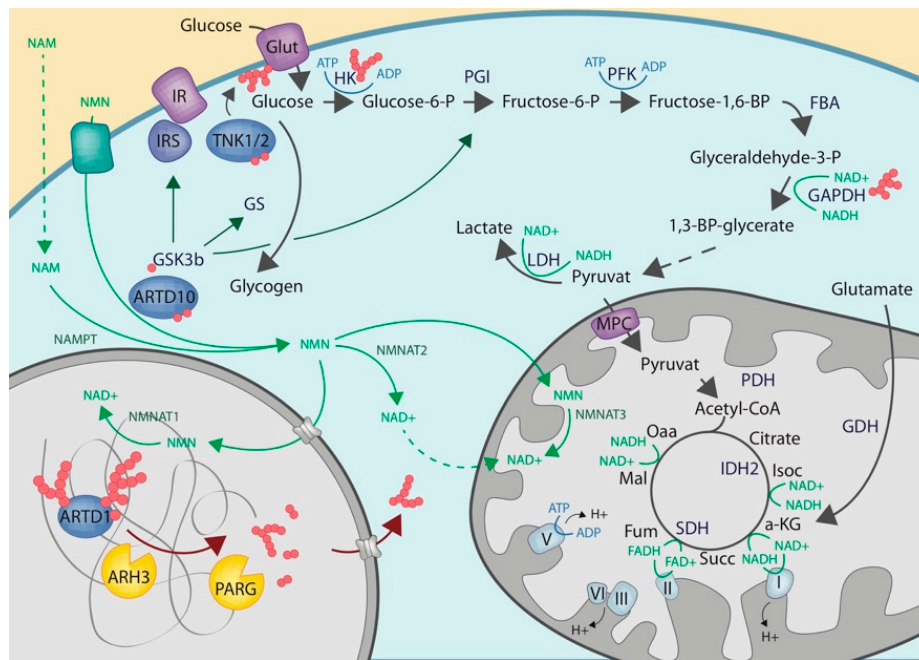


Figure 2. Schematic overview on the functional contribution of cytoplasmic ARTs to carbohydrate metabolism. Metabolic pathways are shown as in Figure 1. ADP-ribosyltransferases are depicted in blue, while ADP-ribosylhydrolases are depicted in yellow.

Glyceraldehyde 3-phosphate dehydrogenase: Another key-metabolic enzyme directly regulated via ADP-ribosylation is the glyceraldehyde 3-phosphate dehydrogenase (GAPDH). GAPDH possesses glyceraldehyde 3-phosphate dehydrogenase activity as well as nitrosylase activity. The enzyme plays an important role in glucose breakdown by converting glyceraldehyde-3-phosphate to 1,3-bisphosphoglycerate; a reaction that requires the reduction of NAD^+ to NADH (Figures 1 and 2). Since the second activity is linked to nuclear processes, the enzyme is considered a moonlighting enzyme [110,111]. Comparable to GSK3b, ADP-ribosylation of GAPDH has been shown to decrease its enzymatic activity. While initial studies suggested that GAPDH auto-ADP-ribosylates itself [112], more recent studies suggest GAPDH to rather be trans-ADP-ribosylated by either ARTD1 or ARTD10 [113,114]. ARTD1-mediated inhibition of GAPDH has been demonstrated in the context of hyperglycemia, where increased glucose concentrations in the cell culture medium resulted in increased DNA double-strand breaks as a consequence of an increase in mitochondrial superoxide production [113]. GAPDH inhibition could further be associated with an increase in PKC activity, as well as $\text{NF-}\kappa\text{B}$ activation; both of which are likely indirect responses to the increase in DNA-damage. In addition, increased metabolic flux into the hexosamine pathway was also observed as a possible consequence to reduced glycolysis rates [113]. Decreasing mitochondrial superoxide production (e.g., via overexpression of UCP-1 or mtSOD2) rescued all of the above-described consequences of hyperglycemia. Besides ARTD1, also ARTD10 was shown to interact with GAPDH [114]. Overexpression of full-length ARTD10 or its catalytic domain lead to an ADP-ribosylation-dependent sequestering of GAPDH in membrane-free, cytoplasmic cell bodies [114]. The functional consequence of this ARTD10-mediated ADP-ribosylation, as well as compartmentalization of GAPDH, has however so far not been further investigated.

3. Mitochondrial NAD Biosynthesis, ADP-Ribosylation Crosstalk and Carbohydrate Metabolism

3.1. Mitochondrial NAD Biosynthesis

Supplementing cell culture medium with NAD^+ or NR has been shown to increase mitochondrial NAD^+ levels in cultured cells [36,115,116]. Intra-mitochondrial NAD^+ has been described to be synthesized from NAM [32,117] or NMN via NMNAT3 [118]. In agreement with this hypothesis,

a mitochondrial localized splice variant of NMNAT3 has been identified and ectopically expressed NMNAT3 shown to localize to the mitochondria [36]. Although one study suggested that NMNAT3 overexpression increased mitochondrial NAD⁺ [119], other studies reported that NMNAT3 was not required for mitochondrial NAD⁺ level maintenance [10,120]. Mitochondrial NAD⁺ levels vary in a circadian fashion and have been shown to directly influence fuel selection [121] and regulate cell survival under stress conditions [32]. Although cytoplasmic and mitochondrial NAD⁺ pools are indirectly connected via glycolysis and oxidative phosphorylation [122], very recent evidence has been presented using isotopic labeling of NR and NAR which suggest a direct NAD⁺ transport between these two intra-cellular spaces [123]. It is, therefore, possible that mitochondrial NAD⁺ comes from both, local synthesis and transport from the cytoplasm; each of these sources are likely to serve as the backup for the other, with both coordinately contributing to the tightly regulated and maintained NAD⁺ concentrations within mitochondria.

3.2. Mitochondrial ADP-Ribosylation

High NAD⁺ concentrations within the mitochondria suggest that these organelles harbor ideal conditions for protein ADP-ribosylation to take place [29]. In fact, the concept of mitochondrial ADP-ribosylation was proposed over 30 years ago when a macromolecular enzymatic product of NAD⁺ was detected in mitochondria isolated from rat livers [117]. This, as well as several follow-up studies, further described ARTs and ADP-ribosylhydrolase (ARHs) activities in mitochondrial lysates [117,124,125]. Unfortunately, these studies failed to identify the corresponding enzymes that mediated these activities. Even the recent identification of the whole mitochondrial proteome did not result in the clear detection of any known ARTs in this organelle [126]. While the distribution of ARTs and ARHs in nucleus and cytoplasm has been quite well described, the existence and distribution of ADP-ribosylating enzymes in mitochondria remains thus controversial. Five enzymes able to regulate ADP-ribosylation have so far been associated with mitochondria: SIRT4 [94,127], ARTD1, PARG, ARH3 [92,128], and MacroD1 [93,129]. ART activity has, in fact, been detected in the mitochondrial matrix, as well as the inner- and outer mitochondrial membranes [130]. Due to the lack of clear evidence regarding mitochondrial localized ARTs and ARHs, this review briefly summarizes what is known about the five enzymes SIRT4, ARTD1, PARG, ARH3, and MacroD1 with respect to mitochondria:

SIRT4: In mammals, 7 SIRT proteins have been described, 3 of which (namely, SIRT3, 4, and 5) localize to the mitochondria (Table 2) [131–133]. Interestingly, SIRT4 has been described to not only hydrolyze protein bound acyl-groups, but to also possess ADP-ribosylation activity [94]. Indeed, SIRT4 was described to ADP-ribosylate and thus negatively regulate the activity of the glutamate dehydrogenase (GDH) in-vitro and in pancreatic β -cells, suggesting SIRT4 as an important regulator of insulin secretion [94,127]. GDH is, however, the only protein thus far described to be ADP-ribosylated by SIRT4 (Figure 3). Recent mass-spectrometry-based analyses identified various ADP-ribosylated mitochondria-localized proteins, thus further strengthening the idea that the mitochondria harbor several enzymes with ADP-ribosylating activities [75,76,134].

ARTD1: ARTD1 mitochondrial localization is the subject of controversial discussions in the field of ADP-ribosylation. This stems from the fact that ARTD1 does not contain a classical mitochondrial localization sequence and, thus, it has been proposed that its translocation is dependent on an association with mitofillin [135]. This association, however, would not explain how ARTD1 would be able to enter mitochondria. While some studies detected ARTD1 in mitochondria, mostly via western-blot analysis; other studies clearly only detect ARTD1 in the nucleus [135–144]. Due to the high abundance of ARTD1, western-blot analysis after cell fractionation (e.g., mitochondrial isolation) should be taken with caution. Moreover, given that the proposed protein targets of mitochondria-localized ARTD1 (mtPARP1) mainly localize to the mitochondrial matrix, more thorough mitochondrial fractionations (including detergent lysis) are required to clarify if and where ARTD1 localizes within the mitochondria. Mitochondrial localization of ARTD1 has been proposed to regulate the mitochondrial energy homeostasis [92,139] (i.e., mitochondrial oxidative capacity) and mitochondrial DNA damage repair [145]. PARP inhibition or

deletion of ARTD1 in different cell lines and systems has been shown to increase mitochondrial oxygen consumption rate [92]. Nevertheless, determining whether ARTD1 needs to localize to mitochondria for these effects requires further experimental investigation. In fact, since nuclear ARTD1 has also been demonstrated to affect whole cell NAD^+ homeostasis [106], all metabolic phenotypes attributed to mitochondrial-localized ARTD1 might well be indirect consequences of the nuclear ARTD1 (see below, nuclear compartment).

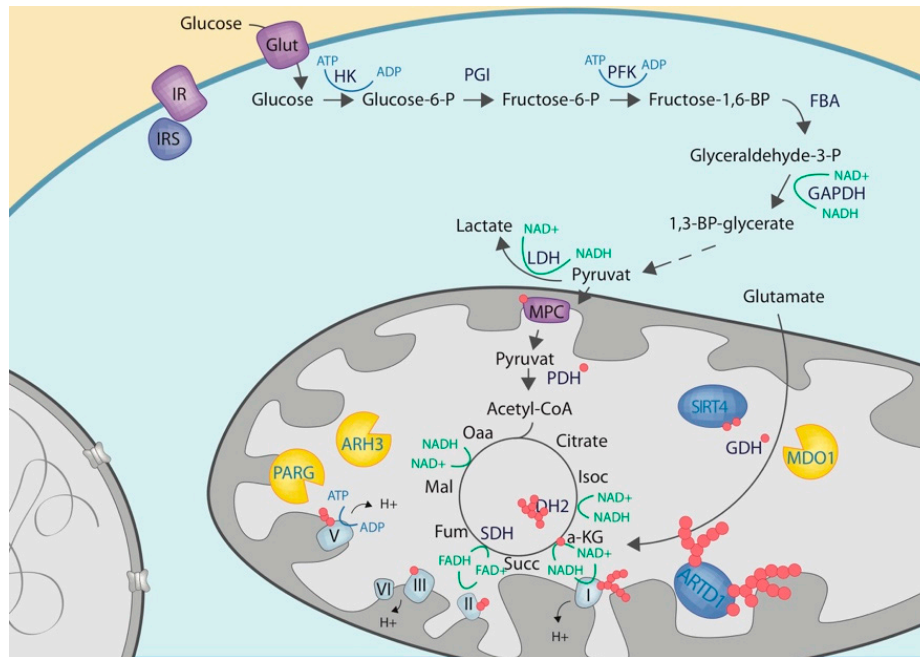


Figure 3. Schematic overview on the functional contribution of mitochondrial ARTs to carbohydrate metabolism. Metabolic pathways are shown as in Figure 1. ADP-ribosyltransferases are depicted in blue, while ADP-ribosylhydrolases are depicted in yellow.

PARG: PARG is able to degrade poly-ADP-ribose [146]. Two shorter splice variants of PARG have been described to reside in the extra-nuclear space [92]. One of these shorter forms contains a predicted N-terminal mitochondrial localization sequence, but direct evidence demonstrating that this isoform of PARG localizes to the mitochondria remains missing. Nevertheless, overexpression studies targeting exogenous PARG ($\Delta 1-460$) to mitochondria indicated that this PARG splice variant is inactive [92,128].

ARH3: ARH3 has recently been shown to hydrolyze both PARylation and MARylation [86,87]. While the majority of ARH3 localizes to the nucleus, a small fraction of the hydrolase has also been detected in mitochondria [86,87]. In contrast to PARG, overexpression of mitochondria-targeted ARH3 reduced mitochondrial protein ADPr metabolism, thus suggesting that ARH3 might indeed be involved in mitochondrial ADPr metabolism [92,128]. Interestingly, while ARH3 could functionally be associated to metabolic diseases in human and in mice [147,148], direct evidence for endogenous mitochondrial ARH3, comparable to PARG, is still missing.

MacroD1: MacroD1 has been shown to be active towards mono-ADP-ribosylated proteins, as well as toward free OAADPR, the by-product of SIRT-catalyzed de-acetylation [88,89,129]. In contrast to the four enzymes discussed above, MacroD1 has been clearly shown to predominantly localize to mitochondria [93,129]. Available transcriptome data from humans, mice, and rats [149] suggest that MacroD1 expression correlates with the energetic status of a given cell or tissue type. While the expression of MacroD1 is rather moderate in most tissues, tissues with a high energetic turnover, like the heart or skeletal muscle, express high levels of this hydrolase. In addition, several studies suggest that MacroD1 plays a role in the regulation of adipogenesis and insulin receptor signaling [150,151]. Upon knockdown of MacroD1 in adipocytes, an increase in glucose uptake and $\text{PPAR}\gamma$ expression on

both mRNA and protein levels was observed; which suggests that it could negatively affect adipocyte differentiation. In addition, this knockdown was also shown to promote insulin signaling, which resulted in increased Akt phosphorylation [150]. In contrast, overexpression of MacroD1 increased ERK1/2- and Akt phosphorylation and insulin content in the pancreatic b-cell-derived MIN6 cell line [151]. While the precise mode of action of MacroD1 remains elusive, it is possible that MacroD1 influences the activity of Akt and insulin signaling in a cell and stimuli specific manner. In fact, MacroD1 has been shown to alter cell proliferation and apoptosis [152], and has been linked to the degree of differentiation and the invasiveness of various types of cancer [153]. The interesting associations of MacroD1 with cell metabolism and metabolism-related diseases make this enzyme an attractive target of further studies.

The function and relevance of mitochondrial ADP-ribosylated proteins is currently under extensive debate. While some labs suggest mitochondrial ADP-ribosylation to be involved in the regulation of metabolic enzymes, others propose that the modification regulate mitochondrial DNA damage repair or oxidative stress responses [154]. Several recent proteomic studies identified various mitochondria-localized proteins to be ADP-ribosylated in lysates from both cells and mouse organs [75,76,134]. The amount of ADP-ribosylated mitochondrial proteins seemed especially pronounced in mitochondria-rich tissues with high metabolic activities such as heart, skeletal muscle, and brown adipose tissue. Nevertheless, detection of endogenous mitochondrial ADP-ribosylation via other methods has thus far proven challenging. Various ADP-ribosylated mitochondrial proteins are implicated in many different biological processes, including mtDNA repair, protein-, metabolite- and ion transport, protein folding and synthesis, and mitochondrial metabolism. The function of ADP-ribosylation in mtDNA repair, has however thoroughly been addressed in previous reviews [154]. As this review focuses mainly on carbohydrate metabolism, proteins involved in non-metabolic processes will not further be discussed.

3.3. Crosstalk with Proteins or Enzymes of the Carbohydrate Metabolisms

Complex I, II, III, and the ATP-synthase: A prominent cluster of mitochondrial ADP-ribosylated proteins is involved in the mitochondrial respiratory chain. Indeed, complex I, II, III, and the ATP-synthase have been shown to be ADP-ribosylated and harbor various potential ADPr acceptor sites [75] (Figure 3). Several studies, including an in-vivo ischemia reperfusion (or traumatic brain injury model) and in vitro activation of the nitric-oxide synthase, provided evidence that the induction of ADP-ribosylation dampens mitochondrial oxidative phosphorylation [136,137,139,155]. While some of the studies directly showed different sub-units of different respiratory chain complexes to be ADP-ribosylated upon induction of the respective stress, other studies only observed an increase in (mitochondrial) ADP-ribosylation in general.

Malate dehydrogenase (MDH), pyruvate dehydrogenase (PDH) and glutamate dehydrogenase (GDH): In addition to the respiratory chain components, other mitochondrial metabolic enzymes involved in carbohydrate metabolism, such as MDH, PDH or GDH were also found to be ADP-ribosylated [75,94,134]. While the functional contribution of ADP-ribosylation to MDH or PDH activities has not yet been experimentally addressed, ADP-ribosylation of the GDH by SIRT4 has been proposed to dampen its activity (see above) [94].

4. Crosstalk between Nuclear ADP-Ribosylation and the Carbohydrate Metabolism

4.1. Nuclear NAD Biosynthesis

The cytoplasmic NAD⁺ pool serves as a central hub that connects and regulates other NAD⁺ pools (Figure 1). Cytosolic and nuclear NAD⁺ pools are closely connected as indicated by studies using the small molecule NAMPT inhibitor FK866, which demonstrated comparable nuclear and cytoplasmic reductions in NAD⁺ following treatment [13]. It is also expected that the NAD⁺ precursors, NAM and NMN, are shared between nucleus and cytosol as they are able to freely diffuse through the nuclear

pore complex. The final step, converting NMN to NAD^+ , is believed to be locally catalyzed. While cytoplasmic NAD^+ is mainly synthesized by NMNAT2, which localizes to the Golgi apparatus [35,156], nuclear NAD^+ is thought to be mainly synthesized by nuclear NMNAT1, the most ubiquitously expressed NMNAT isoform [157,158]. Recent studies found that depleting NMNAT2 significantly reduced cytoplasmic NAD^+ but minimally affected nuclear NAD^+ . Furthermore, nuclear NAD^+ could not completely replenish cytoplasmic NAD^+ following NMNAT2 depletion [13]. Additionally, NMNAT1 has been shown to be recruited to gene promoters targeted by ARTD1, to locally generate NAD^+ and support the catalytic activity of ARTD1 [69,158]. These observations suggest that while nuclear and cytoplasmic NAD^+ pools are interconnected, local regulation by NMNAT1 and NMNAT2, respectively, still plays an important role in NAD^+ homeostasis. This compartment-specific local regulation of NAD^+ biosynthesis seems highly important for controlling the enzymatic activities of ARTs and SIRT.

4.2. Nuclear ADP-Ribosylation

The nuclear role of NAD^+ is mainly centered on ARTD-mediated ADP-ribosylation and SIRT-mediated deacetylation. As ARTDs and SIRTs are NAD^+ -consuming enzymes, their activities require quasi constant replenishment of nuclear NAD^+ , mainly through nuclear NMNAT1-mediated NAD^+ synthesis [9]. Nuclear ARTDs can dampen the activity of SIRTs either directly via ADP-ribosylation or indirectly via substrate competition. Given that nuclear ARTs, especially ARTD1, are able to consume tremendous amounts of NAD^+ upon hyperactivation, they can reduce the substrates/co-factors (NAD^+ , ATP) of many metabolic enzymes. Three of the known ARTDs localize predominantly to the nucleus (ARTD1, ARTD2 and ARTD3), while others shuttle between the cytoplasm and nucleus (ARTD4-6, ARTD8-12, ARTD14) [159]. ARTD1 is the most well-characterized member of the ARTD family and is best-known for its involvement in genotoxic stress responses and as a cleaved marker during apoptosis [160]. ARTD1 however also regulates transcription by different mechanisms including the binding and/or modification of transcription factors (see below), transcription co-factors or histones and thus influencing highly organized chromatin structures [161–165]. Of the seven mammalian SIRT, SIRT1, 6 and 7 have all been shown to localized to the nucleus [57,166]. SIRT6 has been described to be an ADP-ribosyltransferase [91] that directly modifies ARTD1 and BAF170 [167,168], the latter of which is a subunit of BAF chromatin remodeling complex required for activation of a subset of NRF2 responsive genes following oxidative stress [167]. The extensive network of ARTD and SIRT proteins and their regulatory targets indicates the profound impact that nuclear NAD^+ can have on various cellular processes [29]. A direct link between the enzymatic activities of nuclear ARHs and the metabolic state of the cell remains to be explored.

4.3. Crosstalk with Proteins Regulating the Expression of Enzymes Involved in the Carbohydrate Metabolisms

As most enzymes involved in carbohydrate metabolism reside in either the cytoplasm or the mitochondria, nuclear ARTDs have a limited ability to influence their activity directly via ADP-ribosylation. As discussed above, the exception is GAPDH, which is believed to shuttle between the cytoplasm and nucleus in response to metabolic stimulation. Nevertheless, nuclear ARTDs may still indirectly affect various metabolic enzymes; indeed, it is possible that they regulate the expression of genes involved in metabolic processes, potentially via transcription factor ADP-ribosylation (see above).

C/EBP-b: C/EBP-b is an important transcription factor involved in various physiological processes including embryonic development, immune responses, hematopoiesis, adipogenesis, and glucose metabolism [169–171]. C/EBP-b is a direct target of ARTD1-mediated poly-ADP-ribosylation during adipocyte differentiation [69] (Figure 4). In fact, ADP-ribosylation of C/EBP-b during adipogenesis initiation results in its eviction from the DNA. In order to allow C/EBP-b to bind to and activate its target genes during the initial phase of adipogenesis, the activity of ARTD1 had to be dampened via reduction of local NAD^+ [69]. Adipogenic signaling rapidly induces cytoplasmic NMNAT-2, which competed with nuclear NMNAT-1 for the NMN leading to a reduction in nuclear NAD^+ levels. The whole process,

including the changes in cytoplasmic and nuclear NAD^+ levels was dependent on glucose-uptake. Interestingly, cells would still differentiate in a glucose-free medium if ARTD1 was knocked down, pointing towards an interplay between ARTD1 activity, NAD^+ availability, and glucose metabolism.

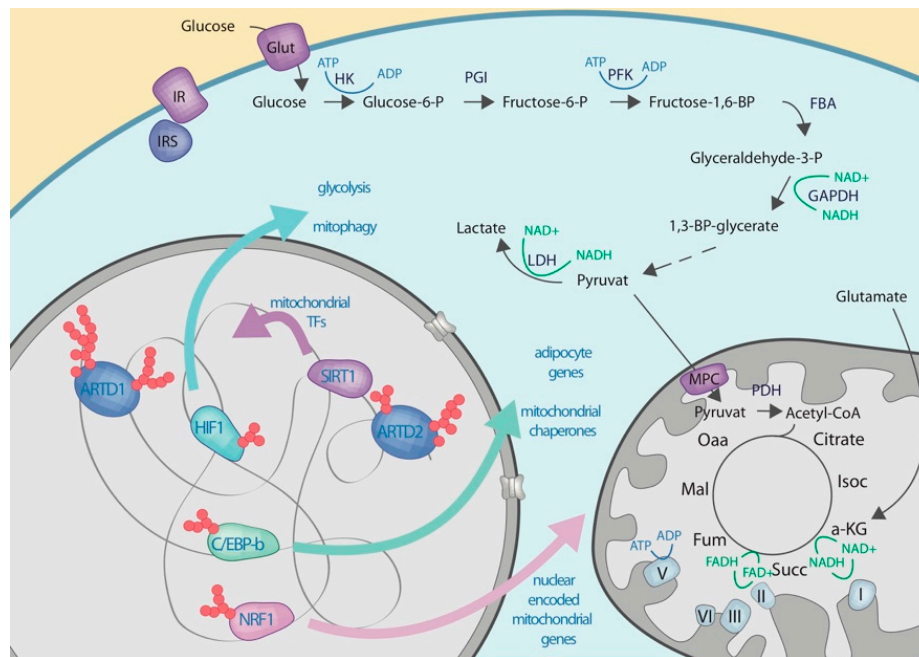


Figure 4. Schematic overview on the functional contribution of nuclear ARTs to carbohydrate metabolism. Metabolic pathways are shown as in Figure 1.

NRF1: Nuclear respiratory factor 1 (NRF1) is a transcription factor that regulates cell growth and metabolism by activating genes involved in mitochondrial biogenesis and function [172]. Together with NRF2, NRF1 mediates nuclear- and the mitochondrial genome coordination by regulating the expression of a cluster of nuclear DNA-encoded genes that localize to the mitochondria [172]. ARTD1 has been shown to form a complex with NRF1 in an ADPr-independent manner facilitating the regulation of its target genes [173] (Figure 4). Although modification is not required for the binding of NRF1 to ARTD1, its enzymatic activity seems to be important for NRF1 transcriptional activity. While the functional contribution of NRF1 in the regulation of mitochondrial dynamics and function is well described, the precise contribution of ARTD1 remains to be investigated. Nonetheless, as ARTD1 activity does affect mitochondrial function and cell metabolism, it is interesting to speculate that ARTD1 interacts with NRF1 in an NAD^+ -dependent manner, and that this interaction transmits information about the metabolic status (i.e., NAD^+ concentration/availability) of the cell between nucleus and mitochondria.

HIF1: Hypoxia-inducible factor (HIF) 1 is the key transcription factor that regulates gene expression in response to hypoxia [174]. The family of human HIF transcription factors is composed of six members, all of which respond to changes in oxygen availability [174]. Upon oxygen deprivation, HIF1 is stabilized and regulates the adaptation of the cells toward hypoxia on the transcriptional level. Many genes involved in this adaptation are metabolic, and this is because hypoxic conditions force cells to rely on glycolysis rather than oxidative phosphorylation [174]. Several studies have demonstrated that ARTD1 forms a complex with HIF1 and acts as a transcriptional co-activator for HIF1 target genes [175,176] (Figure 4). Although HIF1 does not seem to be a direct target of ARTD1, this co-activation has been shown to be ADP-ribosylation dependent, as treatment with PARP inhibitors impaired HIF1 target gene activation [175,176].

SIRT1: SIRT1 is a nuclear de-acetylase involved in the transcriptional regulation of various genes [177]. Expression and activation of SIRT1 is generally positively associated with whole cell

NAD⁺ levels and longevity [177,178]. *ARTD2* deficiency resulted in an increase in SIRT1 on mRNA and protein level, suggesting that *ARTD2* negatively regulates the activity of SIRT1 in cell lines and mice [179–182] (Figure 4). In line with this, *ARTD2*-deficient cells and mice showed higher expression levels of SIRT1 regulated genes, such as PGC-1 α , and displayed increased mitochondrial biogenesis and function [179]. Further, *ARTD2*-deficient animals were protected against diet-induced obesity [179]. Inversely, overexpression of *ARTD2* decreased SIRT1 levels and mitochondrial biogenesis [180]. The effect of *ARTD2* on SIRT1 was found to be NAD⁺ independent, thus excluding a NAD⁺ substrate competition.

KAP1: KRAB-associated protein 1 (KAP1) is an important regulator of chromatin organization involved in numerous processes including embryogenesis, development, cell cycle progression, and apoptosis [183]. KAP1 also plays an important role in the repression of LINE1 transposons that depends on SIRT6-mediated ADP-ribosylation of KAP1 [184]. The association of SIRT6 to LINE1 loci and the modification of KAP1 declines with age. However, the mechanism how KAP1ADP-ribosylation is regulating cell metabolism remains to be investigated.

5. Intercompartmental NAD⁺ Cross-Talks after Genotoxic Stress

Many enzymes involved in glycolysis and oxidative phosphorylation require NAD⁺ or ATP as co-factors for their reactions (Figures 1–3). Consequently, a decrease in cytoplasmic and/or mitochondrial NAD⁺ levels, and subsequently a decrease in whole cell ATP levels, blocks various anabolic and catabolic reactions due to co-factor deficiencies. Studies using either PARP-inhibitors (PARPi) or *ARTD1*-deficient cells revealed an anti-correlation between *ARTD1* activity and intracellular NAD⁺ levels under basal conditions. Thus, implying that *ARTD1* also consumes NAD⁺ under physiological conditions [9]. Hyperactivation of *ARTD1*, as observed following severe genotoxic stress, has also been associated with a substrate competition-mediated decrease in SIRT1 activity [185]. While some studies mainly observed a considerable drop in nuclear and cytoplasmic NAD⁺ levels following genotoxic stress, other studies have also found that mitochondrial NAD⁺ levels drop under these conditions [68,72,106,186]. Several studies have further associated genotoxic stress induced *ARTD1* activation with a drop in ATP levels and glycolysis [68,72,186]. It is important to note that while some studies observed mainly a decrease in glycolysis, other studies also observed reduced mitochondrial oxidative phosphorylation [106]. Both phenotypes could not only be rescued via *ARTD1* inhibition/knockout, but also via NAD⁺-supplementation [68,72,186]. The fact that PARP inhibitor increased NAD⁺ levels in mice, improved mitochondrial function, and protected against the effects of a high fat diet [187,188], provides strong evidence for an intercompartmental cross-talk. Thus, two main hypotheses exist that could explain how hyper-activation of *ARTD1* affects cell metabolism: (i) studies describing a drop in glycolysis suggest that severe genotoxic stress results in a metabolic switch from glycolysis to oxidative phosphorylation, making cells dependent on mitochondria for energy/ATP production, or (ii) studies demonstrating a reduction in both the glycolytic and TCA capacity point towards a general reduction in cell metabolism following *ARTD1* activation and subsequent NAD⁺ depletion. Along with the first possibility, supplementation with non-glucose carbon sources, such as pyruvate or glutamate to directly fuel the TCA cycle, rescued astrocyte cell death following MNNG treatment [68]. In fact, the two hypotheses presented here are not mutually exclusive as the reduced cellular metabolic capacities could be a direct function of the degree of *ARTD1* activation. While mild activation of *ARTD1* might predominantly result in the depletion of cytoplasmic and nuclear NAD⁺ pools, more pronounced activation might also considerably deplete mitochondrial NAD⁺. As a consequence, in the first instance, cells would mainly experience a decrease in glycolysis as they would run out of cytoplasmic NAD⁺, which is required for the conversion of glucose-3-phosphate to 1,3-bisphosphoglycerate. Following a more pronounced drop in NAD⁺ levels, cells would experience further reductions in TCA capacity, as well as in oxidative phosphorylation. Pyruvate and glutamate could reduce the severity of the phenotype in both cases, as both metabolites are not exclusively metabolized within the mitochondria, but can also be utilized by enzymes in the cytoplasm [78].

Whether mitochondria, with their high NAD⁺ concentrations, also contribute to NAD⁺-regulated processes in other cellular compartments requires additional investigation. However, increased mitochondrial NAD⁺ promoted cell survival during genotoxic stress, which was regulated by the mitochondrial SIRT3 and SIRT4 [32]. These data show that mitochondrial NAD⁺ is a major determinant of apoptosis and shed new light on the influence of diet on organ physiology and disease.

Author Contributions: Writing, review & editing of MS: M.O.H., A.-K.H.; Figures: A.-K.H.; Table 1: A.-K.H.; Table 2: A.-K.H. and P.G.

Funding: ADP-ribosylation research in the laboratory of MOH is funded by the Kanton of Zurich and the Swiss National Science Foundation (grant 310030_157019 and 31003A_176177).

Acknowledgments: We thank Deena M. Leslie Pedrioli and Tobias Suter (both University of Zurich) for the helpful discussions and for providing editorial assistance.

Conflicts of Interest: The authors declare no financial interest.

References

1. Verdin, E. NAD⁺ in aging, metabolism, and neurodegeneration. *Science* **2015**, *350*, 1208–1213. [[CrossRef](#)]
2. Katsyuba, E.; Auwerx, J. Modulating NAD⁺ metabolism, from bench to bedside. *EMBO J.* **2017**, *36*, 2670–2683. [[CrossRef](#)]
3. Chiarugi, A.; Dölle, C.; Felici, R.; Ziegler, M. The NAD metabolome—A key determinant of cancer cell biology. *Nat. Rev. Cancer* **2012**, *12*, 741–752. [[CrossRef](#)]
4. Pearce, E.L.; Poffenberger, M.C.; Chang, C.H.; Jones, R.G. Fueling immunity: Insights into metabolism and lymphocyte function. *Science* **2013**. [[CrossRef](#)]
5. Van der Heiden, M.G.; Cantley, L.C.; Thompson, C.B. Understanding the warburg effect: The metabolic requirements of cell proliferation. *Science* **2009**, *324*, 1029–1033. [[CrossRef](#)]
6. MacDonald, M.J. High content of mitochondrial glycerol-3-phosphate dehydrogenase in pancreatic islets and its inhibition by diazoxide. *J. Biol. Chem.* **1981**, *256*, 8287–8290.
7. MacDonald, M.J. Evidence for the malate aspartate shuttle in pancreatic islets. *Arch. Biochem. Biophys.* **1982**, *213*, 643–649. [[CrossRef](#)]
8. Pollak, N.; Dölle, C.; Ziegler, M. The power to reduce: Pyridine nucleotides—Small molecules with a multitude of functions. *Biochem. J.* **2007**. [[CrossRef](#)]
9. Liu, L.; Su, X.; Quinn, W.J.; Hui, S.; Krukenberg, K.; Frederick, D.W.; Redpath, P.; Zhan, L.; Chellappa, K.; White, E.; et al. Quantitative Analysis of NAD Synthesis-Breakdown Fluxes. *Cell Metab.* **2018**. [[CrossRef](#)]
10. Hikosaka, K.; Ikutani, M.; Shito, M.; Kazuma, K.; Gulshan, M.; Nagai, Y.; Takatsu, K.; Konno, K.; Tobe, K.; Kanno, H.; et al. Deficiency of nicotinamide mononucleotide adenylyltransferase 3 (Nmnat3) causes hemolytic anemia by altering the glycolytic flow in mature erythrocytes. *J. Biol. Chem.* **2014**. [[CrossRef](#)]
11. Dölle, C.; Niere, M.; Lohndal, E.; Ziegler, M. Visualization of subcellular NAD pools and intra-organellar protein localization by poly-ADP-ribose formation. *Cell. Mol. Life Sci.* **2010**, *67*, 433–443. [[CrossRef](#)]
12. VanLinden, M.R.; Niere, M.; Nikiforov, A.A.; Ziegler, M.; Dölle, C. Compartment-specific poly-ADP-ribose formation as a biosensor for subcellular NAD pools. *Method. Mol. Biol.* **2017**, *1608*, 45–56.
13. Cambronne, X.A.; Stewart, M.L.; Kim, D.; Jones-Brunette, A.M.; Morgan, R.K.; Farrens, D.L.; Cohen, M.S.; Goodman, R.H. Biosensor reveals multiple sources for mitochondrial NAD⁺. *Science* **2016**. [[CrossRef](#)]
14. Cameron, W.D.; Bui, C.V.; Hutchinson, A.; Loppnau, P.; Gräslund, S.; Rocheleau, J.V. Apollo-NADP⁺: A spectrally tunable family of genetically encoded sensors for NADP⁺. *Nat. Methods* **2016**, *13*, 352–358. [[CrossRef](#)]
15. Sallin, O.; Reymond, L.; Gondrand, C.; Raith, F.; Koch, B.; Johnsson, K. Semisynthetic biosensors for mapping cellular concentrations of nicotinamide adenine dinucleotides. *Elife* **2018**. [[CrossRef](#)]
16. Koch-Nolte, F.; Fischer, S.; Haag, F.; Ziegler, M. Compartmentation of NAD⁺-dependent signalling. *FEBS Lett.* **2011**. [[CrossRef](#)]
17. Alano, C.C.; Tran, A.; Tao, R.; Ying, W.; Karliner, J.S.; Swanson, R.A. Differences among cell types in NAD⁺ compartmentalization: A comparison of neurons, astrocytes, and cardiac myocytes. *J. Neurosci. Res.* **2007**. [[CrossRef](#)]
18. Di Lisa, F.; Menabò, R.; Canton, M.; Barile, M.; Bernardi, P. Opening of the Mitochondrial Permeability Transition Pore Causes Depletion of Mitochondrial and Cytosolic NAD⁺ and Is a Causative Event in the Death of Myocytes in Postischemic Reperfusion of the Heart. *J. Biol. Chem.* **2001**. [[CrossRef](#)]

19. Fjeld, C.C.; Birdsong, W.T.; Goodman, R.H. Differential binding of NAD⁺ and NADH allows the transcriptional corepressor carboxyl-terminal binding protein to serve as a metabolic sensor. *Proc. Natl. Acad. Sci. USA* **2003**. [[CrossRef](#)]
20. Veech, R.L.; Eggleston, L.V.; Krebs, H.A. The redox state of free nicotinamide-adenine dinucleotide phosphate in the cytoplasm of rat liver. *Biochem. J.* **1969**. [[CrossRef](#)]
21. Zhang, Q.; Piston, D.W.; Goodman, R.H. Regulation of corepressor function by nuclear NADH. *Science* **2002**. [[CrossRef](#)]
22. Stein, L.R.; Imai, S.I. The dynamic regulation of NAD metabolism in mitochondria. *Trends Endocrinol. Metab.* **2012**. [[CrossRef](#)]
23. Anderson, R.M.; Bitterman, K.J.; Wood, J.G.; Medvedik, O.; Sinclair, D.A. Nicotinamide and PNC1 govern lifespan extension by calorie restriction in *Saccharomyces cerevisiae*. *Nature* **2003**, *423*, 181–185. [[CrossRef](#)]
24. Nikiforov, A.; Kulikova, V.; Ziegler, M. The human NAD metabolome: Functions, metabolism and compartmentalization. *Crit. Rev. Biochem. Mol. Biol.* **2015**. [[CrossRef](#)]
25. Hassa, P.O.; Haenni, S.S.; Elser, M.; Hottiger, M.O. Nuclear ADP-Ribosylation Reactions in Mammalian Cells: Where Are We Today and Where Are We Going? *Microbiol. Mol. Biol. Rev.* **2006**. [[CrossRef](#)]
26. Bogan, K.L.; Brenner, C. Nicotinic Acid, Nicotinamide, and Nicotinamide Riboside: A Molecular Evaluation of NAD⁺ Precursor Vitamins in Human Nutrition. *Annu. Rev. Nutr.* **2008**, *28*, 115–130. [[CrossRef](#)]
27. Mori, V.; Amici, A.; Mazzola, F.; Di Stefano, M.; Conforti, L.; Magni, G.; Ruggieri, S.; Raffaelli, N.; Orsomando, G. Metabolic profiling of alternative NAD biosynthetic routes in mouse tissues. *PLoS ONE* **2014**. [[CrossRef](#)]
28. Strømmand, Ø.; Niere, M.; Nikiforov, A.A.; VanLinden, M.R.; Heiland, I.; Ziegler, M. Keeping the balance in NAD metabolism. *Biochem. Soc. Trans.* **2019**. [[CrossRef](#)]
29. Cantó, C.; Menzies, K.J.; Auwerx, J. NAD⁺ Metabolism and the Control of Energy Homeostasis: A Balancing Act between Mitochondria and the Nucleus. *Cell Metab.* **2015**. [[CrossRef](#)]
30. Dölle, C.; Skoge, R.; VanLinden, M.; Ziegler, M. NAD Biosynthesis in Humans - Enzymes, Metabolites and Therapeutic Aspects. *Curr. Top. Med. Chem.* **2013**, *13*, 2907–2917. [[CrossRef](#)]
31. Revollo, J.R.; Grimm, A.A.; Imai, S.I. The NAD biosynthesis pathway mediated by nicotinamide phosphoribosyltransferase regulates Sir2 activity in mammalian cells. *J. Biol. Chem.* **2004**. [[CrossRef](#)]
32. Yang, H.; Yang, T.; Baur, J.A.; Perez, E.; Matsui, T.; Carmona, J.J.; Lamming, D.W.W.; Souza-Pinto, N.C.; Bohr, V.A.; Rosenzweig, A.; et al. Nutrient-Sensitive Mitochondrial NAD⁺ Levels Dictate Cell Survival. *Cell* **2007**. [[CrossRef](#)]
33. Revollo, J.R.; Körner, A.; Mills, K.F.; Satoh, A.; Wang, T.; Garten, A.; Dasgupta, B.; Sasaki, Y.; Wolberger, C.; Townsend, R.R.; et al. Namp1/PBEF/Visfatin Regulates Insulin Secretion in β Cells as a Systemic NAD Biosynthetic Enzyme. *Cell Metab.* **2007**. [[CrossRef](#)]
34. Brazill, J.M.; Li, C.; Zhu, Y.; Zhai, R.G. NMNAT: It's an NAD⁺ synthase ... It's a chaperone ... It's a neuroprotector. *Curr. Opin. Genet. Dev.* **2017**. [[CrossRef](#)]
35. Berger, F.; Lau, C.; Dahlmann, M.; Ziegler, M. Subcellular compartmentation and differential catalytic properties of the three human nicotinamide mononucleotide adenylyltransferase isoforms. *J. Biol. Chem.* **2005**. [[CrossRef](#)]
36. Felici, R.; Lapucci, A.; Ramazzotti, M.; Chiarugi, A. Insight into Molecular and Functional Properties of NMNAT3 Reveals New Hints of NAD Homeostasis within Human Mitochondria. *PLoS ONE* **2013**. [[CrossRef](#)]
37. Yaku, K.; Okabe, K.; Nakagawa, T. NAD metabolism: Implications in aging and longevity. *Ageing Res. Rev.* **2018**. [[CrossRef](#)]
38. Johnson, S.; Imai, S. NAD⁺ biosynthesis, aging, and disease. *F1000Research* **2018**. [[CrossRef](#)]
39. Haigis, M.C.; Sinclair, D.A. Mammalian Sirtuins: Biological Insights and Disease Relevance. *Annu. Rev. Pathol. Mech. Dis.* **2010**. [[CrossRef](#)]
40. Ansari, H.R.; Raghava, G.P.S. Identification of NAD interacting residues in proteins. *BMC Bioinform.* **2010**. [[CrossRef](#)]
41. Berger, F.; Ramírez-Hernández, M.H.; Ziegler, M. The new life of a centenarian: Signalling functions of NAD(P). *Trends Biochem. Sci.* **2004**. [[CrossRef](#)]
42. Houtkooper, R.H.; Williams, R.W.; Auwerx, J. Metabolic Networks of Longevity. *Cell* **2010**. [[CrossRef](#)]
43. Camacho-Pereira, J.; Tarragó, M.G.; Chini, C.C.S.; Nin, V.; Escande, C.; Warner, G.M.; Puranik, A.S.; Schoon, R.A.; Reid, J.M.; Galina, A.; et al. CD38 Dictates Age-Related NAD Decline and Mitochondrial Dysfunction through an SIRT3-Dependent Mechanism. *Cell Metab.* **2016**. [[CrossRef](#)]

44. Chini, E.N.; Chini, C.C.S.; Espindola Netto, J.M.; de Oliveira, G.C.; van Schooten, W. The Pharmacology of CD38/NADase: An Emerging Target in Cancer and Diseases of Aging. *Trends Pharmacol. Sci.* **2018**. [[CrossRef](#)]
45. Malavasi, F.; Deaglio, S.; Funaro, A.; Ferrero, E.; Horenstein, A.L.; Ortolan, E.; Vaisitti, T.; Aydin, S. Evolution and function of the ADP ribosyl cyclase/CD38 gene family in physiology and pathology. *Physiol. Rev.* **2008**. [[CrossRef](#)]
46. Opitz, C.A.; Heiland, I. Dynamics of NAD-metabolism: Everything but constant. *Biochem. Soc. Trans.* **2015**. [[CrossRef](#)]
47. Kirchberger, T.; Guse, A.H. Measuring CD38 (ADP-ribosyl Cyclase/Cyclic ADP-ribose hydrolase) activity by reverse-phase HPLC. *Cold Spring Harb. Protoc.* **2013**. [[CrossRef](#)]
48. Horenstein, A.L.; Sizzano, F.; Lusso, R.; Besso, F.G.; Ferrero, E.; Deaglio, S.; Corno, F.; Malavasi, F. CD38 and CD157 Ectoenzymes Mark Cell Subsets in the Human Corneal Limbus. *Mol. Med.* **2009**. [[CrossRef](#)]
49. Hottiger, M.O. Nuclear ADP-Ribosylation and Its Role in Chromatin Plasticity, Cell Differentiation, and Epigenetics. *Annu. Rev. Biochem.* **2015**. [[CrossRef](#)]
50. Gupte, R.; Liu, Z.; Kraus, W.L. Parps and adp-ribosylation: Recent advances linking molecular functions to biological outcomes. *Genes Dev.* **2017**. [[CrossRef](#)]
51. Grube, K.; Bürkle, A. Poly(ADP-ribose) polymerase activity in mononuclear leukocytes of 13 mammalian species correlates with species-specific life span. *Proc. Natl. Acad. Sci. USA* **1992**. [[CrossRef](#)]
52. Guarente, L. Sirtuins, aging, and metabolism. *Cold Spring Harb. Symp. Quant. Biol.* **2011**. [[CrossRef](#)]
53. Chambon, P.; Weill, J.D.; Mandel, P. Nicotinamide mononucleotide activation of a new DNA-dependent polyadenylic acid synthesizing nuclear enzyme. *Biochem. Biophys. Res. Commun.* **1963**. [[CrossRef](#)]
54. Leung, A.K.L. PARPs. *Curr. Biol.* **2017**, *27*, R1249–R1267. [[CrossRef](#)]
55. Haag, F.; Adriouch, S.; Braß, A.; Jung, C.; Möller, S.; Scheuplein, F.; Bannas, P.; Seman, M.; Koch-Nolte, F. Extracellular NAD and ATP: Partners in immune cell modulation. *Purinergic Signal.* **2007**. [[CrossRef](#)]
56. Di Girolamo, M.; Fabrizio, G. Overview of the Mammalian ADP-Ribosyl-Transferases Clostridia Toxin-Like (ARTCs) Family. *Biochem. Pharmacol.* **2019**. [[CrossRef](#)]
57. Imai, S.I.; Guarente, L. It takes two to tango: Nad⁺ and sirtuins in aging/longevity control. *npj Aging Mech. Dis.* **2016**. [[CrossRef](#)]
58. Schreiber, V.; Dantzer, F.; Amé, J.C.; De Murcia, G. Poly(ADP-ribose): Novel functions for an old molecule. *Nat. Rev. Mol. Cell Biol.* **2006**. [[CrossRef](#)]
59. Imai, S.I.; Armstrong, C.M.; Kaerberlein, M.; Guarente, L. Transcriptional silencing and longevity protein Sir2 is an NAD-dependent histone deacetylase. *Nature* **2000**. [[CrossRef](#)]
60. Rajman, L.; Chwalek, K.; Sinclair, D.A. Therapeutic Potential of NAD-Boosting Molecules: The In Vivo Evidence. *Cell Metab.* **2018**. [[CrossRef](#)]
61. Zhu, Y.; Liu, J.; Park, J.; Rai, P.; Zhai, R.G. Subcellular compartmentalization of NAD⁺ and its role in cancer: A sereNADe of metabolic melodies. *Pharmacol. Ther.* **2019**. [[CrossRef](#)]
62. Hottiger, M.O.; Hassa, P.O.; Lüscher, B.; Schüler, H.; Koch-Nolte, F. Toward a unified nomenclature for mammalian ADP-ribosyltransferases. *Trends Biochem. Sci.* **2010**. [[CrossRef](#)]
63. Dölle, C.; Rack, J.G.M.; Ziegler, M. NAD and ADP-ribose metabolism in mitochondria. *FEBS J.* **2013**. [[CrossRef](#)]
64. McLennan, A.G. The Nudix hydrolase superfamily. *Cell. Mol. Life Sci.* **2006**. [[CrossRef](#)]
65. Lin, S.; Gasmí, L.; Xie, Y.; Ying, K.; Gu, S.; Wang, Z.; Jin, H.; Chao, Y.; Wu, C.; Zhou, Z.; et al. Cloning, expression and characterisation of a human Nudix hydrolase specific for adenosine 5'-diphosphoribose (ADP-ribose). *Biochim. Biophys. Acta* **2002**. [[CrossRef](#)]
66. Perraud, A.L.; Fleig, A.; Dunn, C.A.; Bagley, L.A.; Launay, P.; Schmitz, C.; Stokes, A.J.; Zhu, Q.; Bessman, M.J.; Penner, R.; et al. ADP-ribose gating of the calcium-permeable LTRPC2 channel revealed by Nudix motif homology. *Nature* **2001**. [[CrossRef](#)]
67. Perraud, A.L.; Shen, B.; Dunn, C.A.; Rippe, K.; Smith, M.K.; Bessman, M.J.; Stoddard, B.L.; Scharenberg, A.M. NUDT9, a member of the Nudix hydrolase family, is an evolutionarily conserved mitochondrial ADP-ribose pyrophosphatase. *J. Biol. Chem.* **2003**. [[CrossRef](#)]
68. Ying, W.; Alano, C.C.; Garnier, P.; Swanson, R.A. NAD⁺ as a metabolic link between DNA damage and cell death. *J. Neurosci. Res.* **2005**. [[CrossRef](#)]
69. Ryu, K.W.; Nandu, T.; Kim, J.; Challa, S.; DeBerardinis, R.J.; Lee Kraus, W. Metabolic regulation of transcription through compartmentalized NAD⁺ biosynthesis. *Science* **2018**. [[CrossRef](#)]

70. Cameron, A.M.; Castoldi, A.; Sanin, D.E.; Flachsmann, L.J.; Field, C.S.; Puleston, D.J.; Kyle, R.L.; Patterson, A.E.; Hässler, F.; Buescher, J.M.; et al. Inflammatory macrophage dependence on NAD⁺ salvage is a consequence of reactive oxygen species-mediated DNA damage. *Nat. Immunol.* **2019**. [[CrossRef](#)]
71. Minhas, P.S.; Liu, L.; Moon, P.K.; Joshi, A.U.; Dove, C.; Mhatre, S.; Contrepolis, K.; Wang, Q.; Lee, B.A.; Coronado, M.; et al. Macrophage de novo NAD⁺ synthesis specifies immune function in aging and inflammation. *Nat. Immunol.* **2019**. [[CrossRef](#)]
72. Alano, C.C.; Ying, W.; Swanson, R.A. Poly(ADP-ribose) Polymerase-1-mediated Cell Death in Astrocytes Requires NAD⁺ Depletion and Mitochondrial Permeability Transition. *J. Biol. Chem.* **2004**. [[CrossRef](#)]
73. Erener, S.; Hesse, M.; Kostadinova, R.; Hottiger, M.O. Poly(ADP-Ribose) Polymerase-1 (PARP1) Controls Adipogenic Gene Expression and Adipocyte Function. *Mol. Endocrinol.* **2011**. [[CrossRef](#)]
74. Bai, P. Biology of Poly(ADP-Ribose) Polymerases: The Factotums of Cell Maintenance. *Mol. Cell* **2015**. [[CrossRef](#)]
75. Akram, M. Citric Acid Cycle and Role of its Intermediates in Metabolism. *Cell Biochem. Biophys.* **2014**. [[CrossRef](#)]
76. Grimaldi, G.; Corda, D. ADP-ribosylation and intracellular traffic: An emerging role for PARP enzymes. *Biochem. Soc. Trans.* **2019**. [[CrossRef](#)]
77. Catara, G.; Grimaldi, G.; Schembri, L.; Spano, D.; Turacchio, G.; Lo Monte, M.; Beccari, A.R.; Valente, C.; Corda, D. PARP1-produced poly-ADP-ribose causes the PARP12 translocation to stress granules and impairment of Golgi complex functions. *Sci. Rep.* **2017**. [[CrossRef](#)]
78. Jwa, M.; Chang, P. PARP16 is a tail-anchored endoplasmic reticulum protein required for the PERK-and IRE1 α -mediated unfolded protein response. *Nat. Cell Biol.* **2012**. [[CrossRef](#)]
79. Di Paola, S.; Micaroni, M.; Di Tullio, G.; Buccione, R.; Di Girolamo, M. PARP16/ARTD15 is a novel endoplasmic-reticulum-associated mono-ADP-ribosyltransferase that interacts with, and modifies karyopherin- β 1. *PLoS ONE* **2012**. [[CrossRef](#)]
80. Yeh, T.Y.J.; Beiswenger, K.K.; Li, P.; Bolin, K.E.; Lee, R.M.; Tsao, T.S.; Murphy, A.N.; Hevener, A.L.; Chi, N.W. Hypermetabolism, hyperphagia, and reduced adiposity in tankyrase-deficient mice. *Diabetes* **2009**. [[CrossRef](#)]
81. Feijs, K.L.; Kleine, H.; Braczynski, A.; Forst, A.H.; Herzog, N.; Verheugd, P.; Linzen, U.; Kremmer, E.; Lüscher, B. ARTD10 substrate identification on protein microarrays: Regulation of GSK3 β by mono-ADP-ribosylation. *Cell Commun. Signal.* **2013**. [[CrossRef](#)]
82. Lüscher, B.; Bütepage, M.; Ecke, L.; Krieg, S.; Verheugd, P.; Shilton, B.H. ADP-Ribosylation, a Multifaceted Posttranslational Modification Involved in the Control of Cell Physiology in Health and Disease. *Chem. Rev.* **2018**. [[CrossRef](#)]
83. Mueckler, M.; Thorens, B. The SLC2 (GLUT) family of membrane transporters. *Mol. Asp. Med.* **2013**. [[CrossRef](#)]
84. Huang, S.; Czech, M.P. The GLUT4 Glucose Transporter. *Cell Metab.* **2007**. [[CrossRef](#)]
85. Su, Z.; Deshpande, V.; James, D.E.; Stöckli, J. Tankyrase modulates insulin sensitivity in skeletal muscle cells by regulating the stability of GLUT4 vesicle proteins. *J. Biol. Chem.* **2018**. [[CrossRef](#)]
86. Robey, R.B.; Hay, N. Mitochondrial hexokinases, novel mediators of the antiapoptotic effects of growth factors and Akt. *Oncogene* **2006**. [[CrossRef](#)]
87. Fouquerel, E.; Goellner, E.M.; Yu, Z.; Gagné, J.P.; de Moura, M.B.; Feinstein, T.; Wheeler, D.; Redpath, P.; Li, J.; Romero, G.; et al. ARTD1/PARP1 negatively regulates glycolysis by inhibiting hexokinase 1 independent of NAD⁺ depletion. *Cell Rep.* **2014**. [[CrossRef](#)]
88. Lochhead, P.A.; Coghlan, M.; Rice, S.Q.J.; Sutherland, C. Inhibition of GSK-3 selectively reduces glucose-6-phosphatase and phosphoenolpyruvate carboxykinase gene expression. *Diabetes* **2001**. [[CrossRef](#)]
89. Liberman, Z.; Eldar-Finkelman, H. Serine 332 phosphorylation of insulin receptor substrate-1 by glycogen synthase kinase-3 attenuates insulin signaling. *J. Biol. Chem.* **2005**. [[CrossRef](#)]
90. Márton, J.; Fodor, T.; Nagy, L.; Vida, A.; Kis, G.; Brunyánszki, A.; Antal, M.; Lüscher, B.; Bai, P. PARP10 (ARTD10) modulates mitochondrial function. *PLoS ONE* **2018**. [[CrossRef](#)]
91. Nicholls, C.; Li, H.; Liu, J.P. GAPDH: A common enzyme with uncommon functions. *Clin. Exp. Pharmacol. Physiol.* **2012**. [[CrossRef](#)]
92. Sirover, M.A. Pleiotropic effects of moonlighting glyceraldehyde-3-phosphate dehydrogenase (GAPDH) in cancer progression, invasiveness, and metastases. *Cancer Metastasis Rev.* **2018**. [[CrossRef](#)]
93. Kots, A.Y.; Sergienko, E.A.; Bulargina, T.V.; Severin, E.S. Glyceraldehyde-3-phosphate activates auto-ADP-ribosylation of glyceraldehyde-3-phosphate dehydrogenase. *FEBS Lett.* **1993**. [[CrossRef](#)]

94. Du, X.; Matsumura, T.; Edelstein, D.; Rossetti, L.; Zsengellér, Z.; Szabó, C.; Brownlee, M. Inhibition of GAPDH activity by poly(ADP-ribose) polymerase activates three major pathways of hyperglycemic damage in endothelial cells. *J. Clin. Investig.* **2003**. [[CrossRef](#)]
95. Mayo, E.; Fabrizio, G.; Scarpa, E.; Stilla, A.; Dani, N.; Chiacchiera, F.; Kleine, H.; Attanasio, F.; Lüscher, B.; Di Girolamo, M. ARTD10/PARP10 Induces ADP-Ribosylation of GAPDH and Recruits GAPDH into Cytosolic Membrane-Free Cell Bodies When Overexpressed in Mammalian Cells. *Challenges* **2018**. [[CrossRef](#)]
96. Cantó, C.; Houtkooper, R.H.; Pirinen, E.; Youn, D.Y.; Oosterveer, M.H.; Cen, Y.; Fernandez-Marcos, P.J.; Yamamoto, H.; Andreux, P.A.; Cettour-Rose, P.; et al. The NAD⁺ precursor nicotinamide riboside enhances oxidative metabolism and protects against high-fat diet-induced obesity. *Cell Metab.* **2012**. [[CrossRef](#)]
97. Pittelli, M.; Felici, R.; Pitozzi, V.; Giovannelli, L.; Bigagli, E.; Cialdai, F.; Romano, G.; Moroni, F.; Chiarugi, A. Pharmacological Effects of Exogenous NAD on Mitochondrial Bioenergetics, DNA Repair, and Apoptosis. *Mol. Pharmacol.* **2011**. [[CrossRef](#)]
98. Kun, E.; Zimber, P.H.; Chang, A.C.; Puschendorf, B.; Grunicke, H. Macromolecular enzymatic product of NAD⁺ in liver mitochondria. *Proc. Natl. Acad. Sci. USA* **1975**. [[CrossRef](#)]
99. Nikiforov, A.; Dölle, C.; Niere, M.; Ziegler, M. Pathways and subcellular compartmentation of NAD biosynthesis in human cells: From entry of extracellular precursors to mitochondrial NAD generation. *J. Biol. Chem.* **2011**. [[CrossRef](#)]
100. Son, M.J.; Kwon, Y.; Son, T.; Cho, Y.S. Restoration of Mitochondrial NAD⁺ Levels Delays Stem Cell Senescence and Facilitates Reprogramming of Aged Somatic Cells. *Stem Cells* **2016**. [[CrossRef](#)]
101. Yamamoto, M.; Hikosaka, K.; Mahmood, A.; Tobe, K.; Shojaku, H.; Inohara, H.; Nakagawa, T. Nmnat3 is dispensable in mitochondrial NAD level maintenance in vivo. *PLoS ONE* **2016**. [[CrossRef](#)]
102. Peek, C.B.; Affinati, A.H.; Ramsey, K.M.; Kuo, H.Y.; Yu, W.; Sena, L.A.; Ilkayeva, O.; Marcheva, B.; Kobayashi, Y.; Omura, C.; et al. Circadian clock NAD⁺ cycle drives mitochondrial oxidative metabolism in mice. *Science* **2013**. [[CrossRef](#)]
103. Ying, W. NAD⁺/NADH and NADP⁺/NADPH in Cellular Functions and Cell Death: Regulation and Biological Consequences. *Antioxid. Redox Signal.* **2008**. [[CrossRef](#)]
104. Davila, A.; Liu, L.; Chellappa, K.; Redpath, P.; Nakamaru-Ogiso, E.; Paoletta, L.M.; Zhang, Z.; Migaud, M.E.; Rabinowitz, J.D.; Baur, J.A. Nicotinamide adenine dinucleotide is transported into mammalian mitochondria. *Elife* **2018**. [[CrossRef](#)]
105. Roberts, J.H.; Stark, P.; Giri, C.P.; Smulson, M. Cytoplasmic poly(ADP-ribose) polymerase during the HeLa cell cycle. *Arch. Biochem. Biophys.* **1975**. [[CrossRef](#)]
106. Burzio, L.O.; Sáez, L.; Cornejo, R. Poly (ADP-ribose) synthetase activity in rat testis mitochondria. *Biochem. Biophys. Res. Commun.* **1981**. [[CrossRef](#)]
107. Williams, E.G.; Wu, Y.; Wolski, W.; Kim, J.Y.; Lan, J.; Hasan, M.; Halter, C.; Jha, P.; Ryu, D.; Auwerx, J.; et al. Quantifying and Localizing the Mitochondrial Proteome Across Five Tissues in A Mouse Population. *Mol. Cell. Proteom.* **2018**. [[CrossRef](#)]
108. Haigis, M.C.; Mostoslavsky, R.; Haigis, K.M.; Fahie, K.; Christodoulou, D.C.; Murphy, A.J.J.; Valenzuela, D.M.; Yancopoulos, G.D.; Karow, M.; Blander, G.; et al. SIRT4 Inhibits Glutamate Dehydrogenase and Opposes the Effects of Calorie Restriction in Pancreatic β Cells. *Cell* **2006**. [[CrossRef](#)]
109. Ahuja, N.; Schwer, B.; Carobbio, S.; Waltregny, D.; North, B.J.; Castronovo, V.; Maechler, P.; Verdin, E. Regulation of insulin secretion by SIRT4, a mitochondrial ADP-ribosyltransferase. *J. Biol. Chem.* **2007**. [[CrossRef](#)]
110. Niere, M.; Kernstock, S.; Koch-Nolte, F.; Ziegler, M. Functional Localization of Two Poly(ADP-Ribose)-Degrading Enzymes to the Mitochondrial Matrix. *Mol. Cell. Biol.* **2008**. [[CrossRef](#)]
111. Niere, M.; Mashimo, M.; Agledal, L.; Dölle, C.; Kasamatsu, A.; Kato, J.; Moss, J.; Ziegler, M. ADP-ribosylhydrolase 3 (ARH3), not poly(ADP-ribose) glycohydrolase (PARG) isoforms, is responsible for degradation of mitochondrial matrix-associated poly(ADP-ribose). *J. Biol. Chem.* **2012**. [[CrossRef](#)]
112. Agnew, T.; Munnur, D.; Crawford, K.; Palazzo, L.; Mikoc, A.; Ahel, I. MacroD1 is a promiscuous ADP-ribosyl hydrolase localized to mitochondria. *Front. Microbiol.* **2018**. [[CrossRef](#)]
113. Neuvonen, M.; Ahola, T. Differential Activities of Cellular and Viral Macro Domain Proteins in Binding of ADP-Ribose Metabolites. *J. Mol. Biol.* **2009**. [[CrossRef](#)]
114. Richter, C.; Winterhalter, K.H.; Baumhuter, S.; Lotscher, H.R.; Moser, B. ADP-ribosylation in inner membrane of rat liver mitochondria. *Proc. Natl. Acad. Sci. USA* **2006**. [[CrossRef](#)]

115. Schwer, B.; North, B.J.; Frye, R.A.; Ott, M.; Verdin, E. The human silent information regulator (Sir)2 homologue hSIRT3 is a mitochondrial nicotinamide adenine dinucleotide-dependent deacetylase. *J. Cell Biol.* **2002**. [[CrossRef](#)]
116. Choudhary, C.; Weinert, B.T.; Nishida, Y.; Verdin, E.; Mann, M. The growing landscape of lysine acetylation links metabolism and cell signalling. *Nat. Rev. Mol. Cell Biol.* **2014**. [[CrossRef](#)]
117. Onyango, P.; Celic, I.; McCaffery, J.M.; Boeke, J.D.; Feinberg, A.P. SIRT3, a human SIR2 homologue, is an NAD-dependent deacetylase localized to mitochondria. *Proc. Natl. Acad. Sci. USA* **2002**. [[CrossRef](#)]
118. Leutert, M.; Menzel, S.; Braren, R.; Rissiek, B.; Hopp, A.K.; Nowak, K.; Bisceglie, L.; Gehrig, P.; Li, H.; Zolkiewska, A.; et al. Proteomic Characterization of the Heart and Skeletal Muscle Reveals Widespread Arginine ADP-Ribosylation by the ARTC1 Ectoenzyme. *Cell Rep.* **2018**. [[CrossRef](#)]
119. Martello, R.; Leutert, M.; Jungmichel, S.; Bilan, V.; Larsen, S.C.; Young, C.; Hottiger, M.O.; Nielsen, M.L. Proteome-wide identification of the endogenous ADP-ribosylome of mammalian cells and tissue. *Nat. Commun.* **2016**. [[CrossRef](#)]
120. Larsen, S.C.; Hendriks, I.A.; Lyon, D.; Jensen, L.J.; Nielsen, M.L. Systems-wide Analysis of Serine ADP-Ribosylation Reveals Widespread Occurrence and Site-Specific Overlap with Phosphorylation. *Cell Rep.* **2018**. [[CrossRef](#)]
121. Rossi, M.N.; Carbone, M.; Mostocotto, C.; Mancone, C.; Tripodi, M.; Malone, R.; Amati, P. Mitochondrial localization of PARP-1 requires interaction with mitofilin and is involved in the maintenance of mitochondrial DNA integrity. *J. Biol. Chem.* **2009**. [[CrossRef](#)]
122. Lai, Y.; Chen, Y.; Watkins, S.C.; Nathaniel, P.D.; Guo, F.; Kochanek, P.M.; Jenkins, L.W.; Szabó, C.; Clark, R.S.B. Identification of poly-ADP-ribosylated mitochondrial proteins after traumatic brain injury. *J. Neurochem.* **2008**. [[CrossRef](#)]
123. Du, L.; Zhang, X.; Han, Y.Y.; Burke, N.A.; Kochanek, P.M.; Watkins, S.C.; Graham, S.H.; Carcillo, J.A.; Szabó, C.; Clark, R.S.B. Intra-mitochondrial poly(ADP-ribosylation) contributes to NAD⁺ depletion and cell death induced by oxidative stress. *J. Biol. Chem.* **2003**. [[CrossRef](#)]
124. Pankotai, E.; Lacza, Z.; Murányi, M.; Szabó, C. Intra-mitochondrial poly(ADP-ribosylation): Potential role for alpha-ketoglutarate dehydrogenase. *Mitochondrion* **2009**. [[CrossRef](#)]
125. Módis, K.; Gerő, D.; Erdélyi, K.; Szoleczky, P.; Dewitt, D.; Szabo, C. Cellular bioenergetics is regulated by PARP1 under resting conditions and during oxidative stress. *Biochem. Pharmacol.* **2012**. [[CrossRef](#)]
126. Brunyanski, A.; Olah, G.; Coletta, C.; Szczesny, B.; Szabo, C. Regulation of Mitochondrial Poly(ADP-Ribose) Polymerase Activation by the -Adrenoceptor/cAMP/Protein Kinase A Axis during Oxidative Stress. *Mol. Pharmacol.* **2014**. [[CrossRef](#)]
127. Yu, S.W.; Wang, H.; Poitras, M.F.; Coombs, C.; Bowers, W.J.; Federoff, H.J.; Poirier, G.G.; Dawson, T.M.; Dawson, V.L. Mediation of poly(ADP-ribose) polymerase-1 - Dependent cell death by apoptosis-inducing factor. *Science* **2002**. [[CrossRef](#)]
128. Cipriani, G.; Rapizzi, E.; Vannacci, A.; Rizzuto, R.; Moroni, F.; Chiarugi, A. Nuclear poly(ADP-ribose) polymerase-1 rapidly triggers mitochondrial dysfunction. *J. Biol. Chem.* **2005**. [[CrossRef](#)]
129. Poitras, M.F.; Koh, D.W.; Yu, S.W.; Andrabi, S.A.; Mandir, A.S.; Poirier, G.G.; Dawson, V.L.; Dawson, T.M. Spatial and functional relationship between poly(ADP-ribose) polymerase-1 and poly(ADP-ribose) glycohydrolase in the brain. *Neuroscience* **2007**. [[CrossRef](#)]
130. Lapucci, A.; Pittelli, M.; Rapizzi, E.; Felici, R.; Moroni, F.; Chiarugi, A. Poly(ADP-ribose) Polymerase-1 Is a Nuclear Epigenetic Regulator of Mitochondrial DNA Repair and Transcription. *Mol. Pharmacol.* **2011**. [[CrossRef](#)]
131. Druzhyina, N.; Smulson, M.E.; LeDoux, S.P.; Wilson, G.L. Poly(ADP-ribose) polymerase facilitates the repair of N-methylpurines in mitochondrial DNA. *Diabetes* **2000**. [[CrossRef](#)]
132. Ueda, K.; Oka, J.; Narumiya, S.; Miyakawa, N.; Hayaishi, O. Poly ADP-ribose glycohydrolase from rat liver nuclei, a novel enzyme degrading the polymer. *Biochem. Biophys. Res. Commun.* **1972**. [[CrossRef](#)]
133. Abplanalp, J.; Leutert, M.; Frugier, E.; Nowak, K.; Feurer, R.; Kato, J.; Kistemaker, H.V.A.; Filippov, D.V.; Moss, J.; Cafilisch, A.; et al. Proteomic analyses identify ARH3 as a serine mono-ADP-ribosylhydrolase. *Nat. Commun.* **2017**. [[CrossRef](#)]
134. Fontana, P.; Bonfiglio, J.J.; Palazzo, L.; Bartlett, E.; Matic, I.; Ahel, I. Serine ADP-ribosylation reversal by the hydrolase ARH3. *Elife* **2017**. [[CrossRef](#)]

135. Mashimo, M.; Bu, X.; Aoyama, K.; Kato, J.; Ishiwata-Endo, H.; Stevens, L.A.; Kasamatsu, A.; Wolfe, L.A.; Toro, C.; Adams, D.; et al. PARP1 inhibition alleviates injury in ARH3-deficient mice and human cells. *JCI Insight* **2019**. [[CrossRef](#)]
136. Danhauser, K.; Alhaddad, B.; Makowski, C.; Piekutowska-Abramczuk, D.; Syrbe, S.; Gomez-Ospina, N.; Manning, M.A.; Kostera-Pruszczyk, A.; Krahn-Peper, C.; Berutti, R.; et al. Bi-allelic ADPRHL2 Mutations Cause Neurodegeneration with Developmental Delay, Ataxia, and Axonal Neuropathy. *Am. J. Hum. Genet.* **2018**. [[CrossRef](#)]
137. Rosenthal, F.; Feijs, K.L.H.; Frugier, E.; Bonalli, M.; Forst, A.H.; Imhof, R.; Winkler, H.C.; Fischer, D.; Caflich, A.; Hassa, P.O.; et al. Macrodomain-containing proteins are new mono-ADP-ribosylhydrolases. *Nat. Struct. Mol. Biol.* **2013**. [[CrossRef](#)]
138. Jankevicius, G.; Hassler, M.; Golia, B.; Rybin, V.; Zacharias, M.; Timinszky, G.; Ladurner, A.G. A family of macrodomain proteins reverses cellular mono-ADP-ribosylation. *Nat. Struct. Mol. Biol.* **2013**. [[CrossRef](#)]
139. Lattin, J.E.; Schroder, K.; Su, A.I.; Walker, J.R.; Zhang, J.; Wiltshire, T.; Saijo, K.; Glass, C.K.; Hume, D.A.; Kellie, S.; et al. Expression analysis of G Protein-Coupled Receptors in mouse macrophages. *Immunome Res.* **2008**. [[CrossRef](#)]
140. Zang, L.; Xue, B.; Lu, Z.; Li, X.; Yang, G.; Guo, Q.; Ba, J.; Zou, X.; Dou, J.; Lu, J.; et al. Identification of LRP16 as a negative regulator of insulin action and adipogenesis in 3T3-L1 adipocytes. *Horm. Metab. Res.* **2013**. [[CrossRef](#)]
141. Li, X.; Xue, B.; Wang, X.; Sun, L.; Zhang, T.; Qu, L.; Zou, X.; Mu, Y. Reduced expression of the LRP16 gene in mouse insulinoma (MIN6) cells exerts multiple effects on insulin content, proliferation and apoptosis. *J. Huazhong Univ. Sci. Technol. Med. Sci.* **2012**. [[CrossRef](#)]
142. Li, X.J.; Guo, Q.H.; Wang, X.; Xue, B.; Sun, L.Q.; Meng, Q.T.; Lu, J.M.; Mu, Y.M. LRP16 gene protects mouse insulinoma MIN6 cells against fatty acid-induced apoptosis through Akt/FoxO1 signaling. *Chin. Med. J. (Engl.)* **2012**. [[CrossRef](#)]
143. Li, Y.Z.; Zhao, P.; Han, W.D. Clinicopathological significance of LRP16 protein in 336 gastric carcinoma patients. *World J. Gastroenterol.* **2009**. [[CrossRef](#)]
144. Brunyanszki, A.; Szczesny, B.; Virág, L.; Szabo, C. Mitochondrial poly(ADP-ribose) polymerase: The Wizard of Oz at work. *Free Radic. Biol. Med.* **2016**. [[CrossRef](#)]
145. Zhou, H.-Z.; Swanson, R.A.; Simonis, U.; Ma, X.; Cecchini, G.; Gray, M.O. Poly(ADP-ribose) polymerase-1 hyperactivation and impairment of mitochondrial respiratory chain complex I function in reperfused mouse hearts. *Am. J. Physiol. Heart Circ. Physiol.* **2006**. [[CrossRef](#)]
146. Mayer, P.R.; Huang, N.; Dewey, C.M.; Dries, D.R.; Zhang, H.; Yu, G. Expression, localization, and biochemical characterization of nicotinamide mononucleotide adenylyltransferase 2. *J. Biol. Chem.* **2010**. [[CrossRef](#)]
147. Lau, C. The NMN/NaMN adenylyltransferase (NMNAT) protein family. *Front. Biosci.* **2009**, *14*, 410–431. [[CrossRef](#)]
148. Zhang, T.; Berrocal, J.G.; Yao, J.; DuMond, M.E.; Krishnakumar, R.; Ruhl, D.D.; Ryu, K.W.; Gamble, M.J.; Kraus, W.L. Regulation of poly(ADP-ribose) polymerase-1-dependent gene expression through promoter-directed recruitment of a nuclear NAD⁺ synthase. *J. Biol. Chem.* **2012**. [[CrossRef](#)]
149. Hottiger, M.O. SnapShot: ADP-Ribosylation Signaling. *Mol. Cell* **2015**. [[CrossRef](#)]
150. Fouquerel, E.; Sobol, R.W. ARTD1 (PARP1) activation and NAD⁺ in DNA repair and cell death. *DNA Repair (Amst.)* **2014**. [[CrossRef](#)]
151. Kraus, W.L.; Hottiger, M.O. PARP-1 and gene regulation: Progress and puzzles. *Mol. Asp. Med.* **2013**. [[CrossRef](#)]
152. Posavec Marjanović, M.; Crawford, K.; Ahel, I. PARP, transcription and chromatin modeling. *Semin. Cell Dev. Biol.* **2017**. [[CrossRef](#)]
153. Jubin, T.; Kadam, A.; Gani, A.R.; Singh, M.; Dwivedi, M.; Begum, R. Poly ADP-ribose polymerase-1: Beyond transcription and towards differentiation. *Semin. Cell Dev. Biol.* **2017**. [[CrossRef](#)]
154. Schiewer, M.J.; Knudsen, K.E. Transcriptional Roles of PARP1 in Cancer. *Mol. Cancer Res.* **2014**. [[CrossRef](#)]
155. Abplanalp, J.; Hottiger, M.O. Cell fate regulation by chromatin ADP-ribosylation. *Semin. Cell Dev. Biol.* **2017**. [[CrossRef](#)]
156. Chalkiadaki, A.; Guarente, L. The multifaceted functions of sirtuins in cancer. *Nat. Rev. Cancer* **2015**. [[CrossRef](#)]
157. Liszt, G.; Ford, E.; Kurtev, M.; Guarente, L. Mouse Sir2 homolog SIRT6 is a nuclear ADP-ribosyltransferase. *J. Biol. Chem.* **2005**. [[CrossRef](#)]
158. Mao, Z.; Hine, C.; Tian, X.; Van Meter, M.; Au, M.; Vaidya, A.; Seluanov, A.; Gorbunova, V. SIRT6 promotes DNA repair under stress by activating PARP1. *Science* **2011**. [[CrossRef](#)]

159. Rezazadeh, S.; Yang, D.; Tomblin, G.; Simon, M.; Regan, S.P.; Seluanov, A.; Gorbunova, V. SIRT6 promotes transcription of a subset of NRF2 targets by mono-ADP-ribosylating BAF170. *Nucleic Acids Res.* **2019**. [[CrossRef](#)]
160. Darlington, G.J.; Ross, S.E.; MacDougald, O.A. The role of C/EBP genes in adipocyte differentiation. *J. Biol. Chem.* **1998**. [[CrossRef](#)]
161. Poli, V. The role of C/EBP isoforms in the control of inflammatory and native immunity functions. *J. Biol. Chem.* **1998**. [[CrossRef](#)]
162. Westmacott, A.; Burke, Z.D.; Oliver, G.; Slack, J.M.W.; Tosh, D. C/EBP α and C/EBP β are markers of early liver development. *Int. J. Dev. Biol.* **2006**. [[CrossRef](#)]
163. Tiranti, V.; Rossi, E.; Rocchi, M.; DiDonato, S.; Zuffardi, O.; Zeviani, M. The gene (nfe2l1) for human nrf-1, an activator involved in nuclear- mitochondrial interactions, maps to 7q32. *Genomics* **1995**. [[CrossRef](#)]
164. Hossain, M.B.; Ji, P.; Anish, R.; Jacobson, R.H.; Takada, S. Poly(ADP-ribose) polymerase 1 interacts with nuclear respiratory factor 1 (NRF-1) and plays a role in NRF-1 transcriptional regulation. *J. Biol. Chem.* **2009**. [[CrossRef](#)]
165. Smith, T.G.; Robbins, P.A.; Ratcliffe, P.J. The human side of hypoxia-inducible factor. *Br. J. Haematol.* **2008**. [[CrossRef](#)]
166. Hulse, M.; Caruso, L.B.; Madzo, J.; Tan, Y.; Johnson, S.; Tempera, I. Poly(ADP-ribose) polymerase 1 is necessary for coactivating hypoxia-inducible factor-1-dependent gene expression by Epstein-Barr virus latent membrane protein 1. *PLoS Pathog.* **2018**. [[CrossRef](#)]
167. Martínez-Romero, R.; Martínez-Lara, E.; Aguilar-Quesada, R.; Peralta, A.; Oliver, F.J.; Siles, E. PARP-1 modulates deferoxamine-induced HIF-1 α accumulation through the regulation of nitric oxide and oxidative stress. *J. Cell. Biochem.* **2008**. [[CrossRef](#)]
168. Rahman, S.; Islam, R. Mammalian Sirt1: Insights on its biological functions. *Cell Commun. Signal.* **2011**. [[CrossRef](#)]
169. Satoh, A.; Brace, C.S.; Rensing, N.; Cliften, P.; Wozniak, D.F.; Herzog, E.D.; Yamada, K.A.; Imai, S.I. Sirt1 extends life span and delays aging in mice through the regulation of Nk2 Homeobox 1 in the DMH and LH. *Cell Metab.* **2013**. [[CrossRef](#)]
170. Bai, P.; Canto, C.; Brunyánszki, A.; Huber, A.; Szántó, M.; Cen, Y.; Yamamoto, H.; Houten, S.M.; Kiss, B.; Oudart, H.; et al. PARP-2 regulates SIRT1 expression and whole-body energy expenditure. *Cell Metab.* **2011**. [[CrossRef](#)]
171. Mohamed, J.S.; Hajira, A.; Pardo, P.S.; Boriek, A.M. MicroRNA-149 inhibits PARP-2 and promotes mitochondrial biogenesis via SIRT-1/PGC-1 α network in skeletal muscle. *Diabetes* **2014**. [[CrossRef](#)]
172. Szántó, M.; Rutkai, I.; Hegedus, C.; Czikora, Á.; Rózsahegy, M.; Kiss, B.; Virág, L.; Gergely, P.; Tóth, A.; Bai, P. Poly(ADP-ribose) polymerase-2 depletion reduces doxorubicin-induced damage through SIRT1 induction. *Cardiovasc. Res.* **2011**. [[CrossRef](#)]
173. Geng, B.; Cai, Y.; Gao, S.; Lu, J.; Zhang, L.; Zou, J.; Liu, M.; Yu, S.; Ye, J.; Liu, P. PARP-2 knockdown protects cardiomyocytes from hypertrophy via activation of SIRT1. *Biochem. Biophys. Res. Commun.* **2013**. [[CrossRef](#)]
174. Iyengar, S.; Farnham, P.J. KAP1 protein: An enigmatic master regulator of the genome. *J. Biol. Chem.* **2011**. [[CrossRef](#)]
175. Van Meter, M.; Kashyap, M.; Rezazadeh, S.; Geneva, A.J.; Morello, T.D.; Seluanov, A.; Gorbunova, V. SIRT6 represses LINE1 retrotransposons by ribosylating KAP1 but this repression fails with stress and age. *Nat. Commun.* **2014**. [[CrossRef](#)]
176. Cantó, C.; Sauve, A.A.; Bai, P. Crosstalk between poly(ADP-ribose) polymerase and sirtuin enzymes. *Mol. Asp. Med.* **2013**. [[CrossRef](#)]
177. Zong, W.X.; Ditsworth, D.; Bauer, D.E.; Wang, Z.Q.; Thompson, C.B. Alkylating DNA damage stimulates a regulated form of necrotic cell death. *Genes Dev.* **2004**. [[CrossRef](#)]
178. Bai, P.; Cantó, C.; Oudart, H.; Brunyánszki, A.; Cen, Y.; Thomas, C.; Yamamoto, H.; Huber, A.; Kiss, B.; Houtkooper, R.H.; et al. PARP-1 inhibition increases mitochondrial metabolism through SIRT1 activation. *Cell Metab.* **2011**. [[CrossRef](#)]
179. Pirinen, E.; Cantó, C.; Jo, Y.S.; Morato, L.; Zhang, H.; Menzies, K.J.; Williams, E.G.; Mouchiroud, L.; Moullan, N.; Hagberg, C.; et al. Pharmacological inhibition of poly(ADP-ribose) polymerases improves fitness and mitochondrial function in skeletal muscle. *Cell Metab.* **2014**. [[CrossRef](#)]

180. Luo, X.; Ryu, K.W.; Kim, D.S.; Nandu, T.; Medina, C.J.; Gupte, R.; Gibson, B.A.; Soccio, R.E.; Yu, Y.; Gupta, R.K.; et al. PARP-1 Controls the Adipogenic Transcriptional Program by PARylating C/EBP β and Modulating Its Transcriptional Activity. *Mol. Cell* **2017**. [[CrossRef](#)]
181. Vyas, S.; Matic, I.; Uchima, L.; Rood, J.; Zaja, R.; Hay, R.T.; Ahel, I.; Chang, P. Family-wide analysis of poly(ADP-ribose) polymerase activity. *Nat. Commun.* **2014**. [[CrossRef](#)]
182. Vyas, S.; Chesarone-Cataldo, M.; Todorova, T.; Huang, Y.H.; Chang, P. A systematic analysis of the PARP protein family identifies new functions critical for cell physiology. *Nat. Commun.* **2013**. [[CrossRef](#)]
183. Verheugd, P.; Bütepage, M.; Ecke, L.; Lüscher, B. Players in ADP-ribosylation: Readers and Erasers. *Curr. Protein Pept. Sci.* **2016**, *17*, 654–667. [[CrossRef](#)]
184. Yang, C.S.; Jividen, K.; Spencer, A.; Dworak, N.; Ni, L.; Oostdyk, L.T.; Chatterjee, M.; Kušmider, B.; Reon, B.; Parlak, M.; et al. Ubiquitin Modification by the E3 Ligase/ADP-Ribosyltransferase Dtx3L/Parp9. *Mol. Cell* **2017**. [[CrossRef](#)]
185. Slade, D.; Dunstan, M.S.; Barkauskaite, E.; Weston, R.; Lafite, P.; Dixon, N.; Ahel, M.; Leys, D.; Ahel, I. The structure and catalytic mechanism of a poly(ADP-ribose) glycohydrolase. *Nature* **2011**. [[CrossRef](#)]
186. Ohashi, S.; Kanai, M.; Hanai, S.; Uchiumi, F.; Maruta, H.; Tanuma, S.I.; Miwa, M. Subcellular localization of poly(ADP-ribose) glycohydrolase in mammalian cells. *Biochem. Biophys. Res. Commun.* **2003**. [[CrossRef](#)]
187. Mashimo, M.; Kato, J.; Moss, J. Structure and function of the ARH family of ADP-ribosyl-acceptor hydrolases. *DNA Repair (Amst)*. **2014**. [[CrossRef](#)]
188. Feijs, K.L.H.; Forst, A.H.; Verheugd, P.; Lüscher, B. Macrodomein-containing proteins: Regulating new intracellular functions of mono(ADP-ribosyl)ation. *Nat. Rev. Mol. Cell Biol.* **2013**. [[CrossRef](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).