



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2013

**Advances in ancient DNA research can help radiological interpretations of
archaeological diseases**

Bouwman, Abigail S ; Rühli, Frank J

DOI: <https://doi.org/10.1007/s00256-012-1568-1>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-93808>

Journal Article

Published Version

Originally published at:

Bouwman, Abigail S; Rühli, Frank J (2013). Advances in ancient DNA research can help radiological interpretations of archaeological diseases. *Skeletal Radiology*, 42(6):751-752.

DOI: <https://doi.org/10.1007/s00256-012-1568-1>

Advances in ancient DNA research can help radiological interpretations of archaeological diseases

Abigail Bouwman · Frank Rühli

Received: 9 December 2012 / Accepted: 16 December 2012 / Published online: 12 January 2013
© ISS 2013

Evolutionary medicine explores among other things the human vulnerability towards pathogens. To increase the diagnostic level of evidence, alongside radiological investigations, molecular techniques (such as ancient DNA analyses) are often being applied.

Ancient DNA (aDNA) is DNA recovered from archaeological and historical specimens. This DNA is highly degraded and fragmented by biological and environmental factors and is therefore difficult to analyse. However, sequences retrieved from ancient material are highly useful in many ways, including for medical purposes.

The first aDNA experiment was conducted in 1984 to determine the biological classification of the extinct quagga, revealed to be a close relative of the zebra [1]. In the 1980s and 1990s there were multiple publications in which the authors claimed to have extracted aDNA from a variety of samples. However, some of these sequences were later determined to be the result of modern contamination. This led to the introduction of vigorous anti-contamination procedures in the field of aDNA research. At this time a greater understanding of the need for hypothesis-led research allowed for a great diversity of targets. Ancient DNA has been used extensively, for example, to clarify the domestication events of animals and plants, to understand the movements of humans, and to examine the history of modern and ancient disease.

Among the first pathogens targeted by ancient DNA researchers were the *Mycobacterium tuberculosis* complex (MTBC) organisms, and this DNA has been proven to be one of the most robust of all the pathogens studied in archaeological remains. Tuberculosis is a chronic infection caused by any of the members of the MTBC, which are very similar genetically.

Tuberculosis can cause numerous bone changes, most significantly in Pott's disease, where the vertebrae are pitted and gradually undergo extensive osteoblastic activity, resulting in spinal collapse, and cause a hunched posture. Whilst tuberculosis can be inferred from visual (including radiological) inspection of archaeological remains, bone changes only occur in a minority of cases. In addition, the actual causative bacteria cannot be known without microbiological testing.

Many groups have amplified and sequenced archaeological MTBC DNA, and increasingly try to identify the members of the complex; strain identification is easier than type identification as less genetic information is required. Interestingly, so far, only one archaeological tuberculosis case has been shown to have been caused by the *Mycobacterium bovis* strain, despite the close bovine–human habitation of the past.

Modern geneticists are increasingly studying the phylogeography of the MTBC, i.e. the correlation between the strain and its location. This involves examining multiple mutations from one sample. Traditional polymerase chain reaction (PCR) methods work by targeting specific DNA using primers (short pieces of nucleotides that attach to the complementary sequence) and copying the DNA in-between. The DNA is then sequenced in a separate experiment. Multiple primers can be used in one reaction, but generally this only amplifies a few targets, resulting in up to only a few thousand bases of DNA from a limited number of sequences. Therefore, it is highly unlikely that enough informative information could be gathered from one degraded archaeological sample to properly place an ancient bacterium into a phylogeographic type.

Recently introduced into the research field of aDNA studies, next generation sequencing (NGS) works by taking the DNA and adding short sequences to either end that are complementary to universal primers, eliminating the specificity of conventional PCR. Each piece of DNA is then amplified and sequenced individually, so that thousands of strands can be

A. Bouwman · F. Rühli (✉)
Centre for Evolutionary Medicine, Institute of Anatomy,
University of Zurich, Winterthurerstrasse 190,
8057 Zürich, Switzerland
e-mail: frank.ruhli@anatom.uzh.ch

analysed simultaneously (see Mardis [2] for an overview). Therefore, the development of NGS systems allows for thousands of targets and millions of bases to be sequenced at once.

Using one of these NGS systems (SOLiD), MTBC DNA from a 19th century British individual has been typed and identified as a similar, but different type from that isolated in North America in 1905, known as H37Rv [3]. One hundred other individuals from the last 2,000 years across Europe are still being analysed; the bone changes and strain identification are being compared to see if there is any correlation between the strain and the pathology.

Next-generation sequencing has opened up a new field of research to the evolutionary medicine investigator and will continue—alongside radiological diagnostics—to expand our understanding of our past and present health.

Acknowledgment The authors acknowledge funding by the Mäxi Foundation, Zurich.

Conflict of interest None.

References

1. Higuchi R, Bowman B, Freiberger M, Ryder OA, Wilson AC. DNA sequences from the quagga, an extinct member of the horse family. *Nature*. 1984;312:282–4.
2. Mardis ER. Next-generation DNA sequencing methods. *Annu Rev Genomics Hum Genet*. 2008;9:387–402.
3. Bouwman AS, Kennedy SL, Müller R, et al. Genotype of a historic strain of *Mycobacterium tuberculosis*. *Proc Natl Acad Sci U S A*. 2012;109:18511–6.